

2 Pines

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2.1 Introduction

2.1.1 History of the Genus

Origin and Distribution

Pinus is the most important genus within the Family *Pinaceae* and also within the gymnosperms by the number of species (109 species recognized by Farjon 2001) and by its contribution to forest ecosystems. All pine species are evergreen trees or shrubs. They are widely distributed in the northern hemisphere, from tropical areas to northern areas in America and Eurasia. Their natural range reaches the equator only in Southeast Asia. In Africa, natural occurrences are confined to the Mediterranean basin. Pines grow at various elevations from sea level (not usual in tropical areas) to highlands. Two main regions of diversity are recorded, the most important one in Central America (43 species found in Mexico) and a secondary one in China. Some species have a very wide natural range

(e.g., *P. ponderosa*, *P. sylvestris*). Pines are adapted to a wide range of ecological conditions: from tropical (e.g., *P. merkusii*, *P. kesiya*, *P. tropicalis*), temperate (e.g., *P. pungens*, *P. thunbergii*), and subalpine (e.g., *P. albicaulis*, *P. cembra*) to boreal (e.g., *P. pumila*) climates (Richardson and Rundel 1998, Burdon 2002). They can grow in quite pure stands or in mixed forest with other conifers or broadleaved trees. Some species are especially adapted to forest fires, e.g., *P. banksiana*, in which fire is virtually essential for cone opening and seed dispersal. They can grow in arid conditions, on alluvial plain soils, on sandy soils, on rocky soils, or on marsh soils. Trees of some species can have a very long life as in *P. longaeva* (more than 3,000 years).

Botanical Descriptions

The genus is distinguished from other members of the *Pinaceae* family by its needlelike secondary leaves, borne commonly in fascicles of 1 to 8 on dwarf shoots, with a fascicle sheath of bud scales. The leaves of pines are of four types encompassing the complete plant development: cotyledons, juvenile leaves, scale leaves

(cataphylls), and secondary leaves. Cotyledons vary in number from 4 up to 24 in *P. maximartinezii*. There are up to three resin ducts in the cotyledons and either one or two vascular bundles (Farjon 1984). Primary leaves are single, generally helically arranged, and acicular, and they are produced in most species only during the first growth season but for a longer period in a few species. Cataphylls, the nonchlorophyllous primary leaves produced on shoots, occur in an extension of the helical arrangement of the primary leaves and subtend all shoot structures, but they are typically small and subulate or lanceolate. Secondary leaves, the needles, appear by the end of the first growing season, or later in some species. They are the most common pine leaves, permanently green, and metabolically active, ranging in length from 2 to 50 cm (generally smaller ones occur in subalpine or aridity-adapted species). They are borne on dwarf shoots axillary to cataphylls, in fascicles of one (*P. monophylla*, with circular section needles) to eight needles, usual numbers per fascicle being two, three, or five. A fascicle is initially bound together by a basal sheath that may then fall off or persist, but actual leaf fall involves the entire fascicle structure. The number of leaves in the fascicles determines the transverse leaf shape (Farjon and Styles 1997). Stomata are arranged in several longitudinal lines along the entire leaf length. There are generally two or more resin ducts in needles. The number of vascular bundles (one or two as in cotyledons) is the major trait for the identification of the main divisions of genus *Pinus*. The trunk is usually single, erect, and columnar. The branches are grouped into pseudowhorls (often called clusters), at least when young. Bark patterns in pines result mainly from fissuring due to expansion growth and to the formation of scales that eventually fall off.

Pine species are monoecious. Pollen cones are relatively small and soft; these ephemeral structures consist of an axis with many helically arranged microsporophylls. Two microsporangia are attached to the underside of each microsporophyll. These cones open in spring, at least for temperate-climate species, and release large quantities of pollen into the air. Pollen is of the bisaccate type giving it great buoyancy in the air. Pine pollen can be blown over long distances. Its morphology is very similar for all pine species. Seed conelets are found in most species at the ends of new twigs, taking the position of a lateral bud. They are usually located on the higher branches. They consist of scales, the megasporophylls, arranged around an axis; on each megasporophyll lie two separate ovules,

each consisting of a cell mass protected by an integument. The micropyle through which the pollen tube penetrates is turned toward the axis. After pollination the seed cone closes its scales by expansion. The seed cone then grows rapidly. Pollen germinates and produces the male gametophyte, with two sperm nuclei. Fertilization takes place later, about one year after pollination in temperate pine species. Seed cone maturation requires one growing season after fertilization for many species, and even a third for some species. In tropical climates the cycle can be shortened because of the lack of winter dormancy. Individual seed cones thus persist for 2, up to even 3, years on the same tree for most pine species. Seeds contain an embryo embedded in the remaining megagametophyte and the seed coat. The seed wings derive from tissue on the adaxial face of the seed scale. Even in species with vestigial or absent wings, remnants of basal wing tissue are present on the seed scale, on the seed, or on both. Seeds are mostly wind dispersed. In some species, birds are important seed-dispersal vectors (*P. albicaulis*). Seed cones are serotinous in some species and open only following exposure to fire (*P. banksiana*).

Systematics and Phylogeny

The genus is divided into subgenera, sections, and subsections. Various classifications have been proposed in this genus since Linnaeus. Recent ones obtain support from DNA phylogenetics to identify related species. Many phylogenetic studies have been carried out in pines. Some of the first studies involved restriction patterns of the chloroplast genome (Strauss and Doerksen 1990; Govindaraju et al. 1992; Krupkin et al. 1996). More recent classifications, including a large number of species, were established from nuclear sequences (ITS, Liston et al. 1999) and chloroplast sequences (*rbcl*: Gernandt et al. 2005), the chloroplast genome being paternally inherited in the genus *Pinus* (Neale and Sederoff 1989). Some studies have also focused on subsets of the genus *Pinus*: subgenus *Pinus* (Geda López et al. 2002), section *Parrya* (Gernandt et al. 2003), and Eurasian species (Wang et al. 1999). Comprehensive classifications of the genus *Pinus* were earlier established by Gaussen (1960) and Van der Burgh (1973) using morphological and anatomical traits. Later, Price et al. (1998) and Gernandt et al. (2005) also included molecular data and identified monophyletic subgenera, sections, and subsections. Some features are consistent, but variations are noticed between the classical and

molecular approaches. The main division into two subgenera according to the number (one or two) of leaf vascular bundles has been recognized by these authors with various subgenera names (*Haploxylon* and *Diploxylon*, sometimes called, respectively, soft pines and hard pines, and, more recently, *Strobos* and *Pinus* named from type species as recommended by botanical nomenclature code; <http://tolweb.org/tree?group=Pinus&contgroup=Pinaceae>). The taxonomic position of a singular species with flat secondary leaves *P. krempfii* is not fully agreed. It has been considered as a third monospecific subgenus by Gaussen (1960), while molecular data place it as a member of the subgenus *Strobos* (Wang et al. 2000; Gernandt et al. 2005). Most species belong to the subgenus *Pinus*. Subgenus *Pinus* species are characterized by thick seed-cone scales and persistent fascicle sheaths. The numerous sections proposed by Van der Burgh (1973) for this subgenus have subsequently been grouped into two sections *Pinus* and *Trifoliae* (Gernandt et al. 2005), the latter being called New World diploxylon pines by Price et al. (1998). The section *Trifoliae* consists of American species distributed into subsections *Austerales* (septal, internal, or medial needle resin ducts), *Ponderosae* (internal or medial needle resin ducts), and *Contortae* (medial needle resin ducts); most of these species are characterized by three-needled fascicles (Table 1).

Each of these subsections groups two or more previously described subsections. *P. leiophylla* and *P. lumholtzii* are now clustered within subsection *Austerales* and are not further differentiated. The section *Pinus* is divided into subsections *Pinus* and *Pinaster*. Species within this section, with few exceptions, grow in Eurasia and northern Africa. Subsection *Pinaster*, characterized by the lack of a spine on the umbo of the cone scale, includes *P. pinaster* as the type. More recently, all of the other species included in the section *Pinaster* as defined by Van der Burgh (1973) were found to cluster within the *Austerales* subsection of the genus (Gernandt et al. 2005). Species of the subsection *Pinus* including the type species of the genus, *P. sylvestris*, were previously grouped into a section called *Sylvestres* by Van der Burgh (1973). The subgenus *Strobos* has been divided into two sections: *Quinquefoliae* and *Parrya*. They differ from the sections *Strobos* and *Parrya* of Van der Burgh (1973) and Price et al. (1998) by the transfer of the subsections *Krempfianae* and *Gerardianae* from the section *Parrya* into the section *Quinquefoliae*, which also includes the subsection *Strobos*. The subsection *Strobos*

consists of species with five-needled fascicles, thin cone scales, terminal position of spines on seed cone, and several other features absent in the subsections *Krempfianae* and *Gerardianae* of Southeast Asia already differentiated by Van der Burgh (1973) and Price et al. (1998). The three subsections of the *Quinquefoliae* section share a deciduous fascicle sheath. The section *Parrya* consists of the subsections *Cembroides*, *Nelsoniae*, and *Balfourianae*; they share an American distribution, the external position of resin ducts (as in subsections *Krempfianae* and *Gerardianae*), and thick cone scales (again as in subsections *Krempfianae* and *Gerardianae*). Subsection *Nelsoniae* shows persistent fascicle sheath not found in other species of subgenus *Strobos*. Most monophyletic groups cannot be identified from unique morphoanatomical traits.

Two of the 11 subsections consist of American species and Eurasian species. Sections *Pinus* and *Quinquefoliae* have an Asiatic origin according to chloroplast data. The subsection *Strobos* lineage would have then evolved in America before coming back to Eurasia. Few dispersal events to eastern North America have probably occurred to explain the presence there of the limited number of species of the subsection *Pinus*. The development and utilization of low-copy-number nuclear genes (Syring et al. 2005) should provide new insights to solve remaining classification problems. Most ancient pine fossils have been dated to the early Cretaceous (Millar 1998). They have been found in China, North America, and Europe (which was very close to eastern North America at that time). They did not further refine the putative geographic origin of genus *Pinus*.

Hybridization

Interspecific hybridization occurs in pines but is limited mostly to related species within a subsection. Some species such as *P. engelmannii*, *P. jeffreyi*, and *P. ponderosa* are compatible in a number of different combinations (Liston et al. 1999). Natural hybridizations are often indicated by the introgression of the paternally inherited chloroplast genome. Barriers occur at different stages, from the failure of pollen germination to failure at embryogenesis (Ledig 1998). A few species are postulated to have been derived from interspecific hybridization. They also exhibit the highly conserved chromosome number in pines ($2n = 24$). This is the situation for *P. densata* that has been shown to combine nuclear polymorphisms of *P. tabulaeformis* and *P. yunnanensis* with the chloroplast genome of the latter (Wang et al. 2001) and probably several other

Table 1. Systematics of the genus *Pinus*, according to Gernandt et al. (2005)

Subgenus	Section	Subsection	Number of species	Best-known species	Distribution
<i>Pinus</i>	<i>Pinus</i>	<i>Pinus</i>	17	<i>P. sylvestris</i> , <i>P. kesiya</i> , <i>P. merkusii</i>	Eurasia, North America
		<i>Pinaster</i>	7	<i>P. pinaster</i>	Mediterranean, Asia
	<i>Trifoliae</i>	<i>Contortae</i>	4	<i>P. banksiana</i> , <i>P. contorta</i>	America
		<i>Australes</i>	26	<i>P. elliotii</i> , <i>P. radiata</i> , <i>P. taeda</i>	America
		<i>Ponderosae</i>	17	<i>P. jeffreyi</i> , <i>P. ponderosa</i>	America
<i>Strobis</i>	<i>Parrya</i>	<i>Balfourianae</i>	3	<i>P. balfouriana</i>	America
		<i>Cembroides</i>	11	<i>P. cembroides</i> , <i>P. culminicola</i>	America
		<i>Nelsoniae</i>	1	<i>P. nelsonii</i>	Central America
	<i>Quinquefoliae</i>	<i>Gerardianae</i>	3	<i>P. bungeana</i> <i>P. gerardiana</i>	Asia
		<i>Krempfianae</i>	1	<i>P. krempfii</i>	Asia
		<i>Strobis</i>	21	<i>P. cembra</i> , <i>P. lambertiana</i> , <i>P. strobus</i>	America, Eurasia

species. *P. densata* exchanged genes with ancestral populations prior to its isolation with local differentiation (Ma et al. 2006). Combinations of parental traits and selection for adaptation to new conditions favored colonization of new territories by the hybrid species.

2.1.2

Cytogenetics, DNA Content, and Genome Composition

Sax and Sax (1933), Mergen (1958), and Khoshoo (1961) were the earliest to describe the karyotypes of various conifer species. They found that species of the genus *Pinus* were diploid with 24 chromosomes ($2n = 2x = 24$). The chromosomal complements generally consist of 10 or 11 pairs of large homobrachial (metacentric) chromosomes and one or two pairs of smaller heterobrachial (submetacentric) chromosomes (Saylor 1961, 1964, 1972, 1983). Several attempts have been made to construct chromosome-specific karyotypes for various pine species using traditional cytogenetics techniques, viz., C-banding, Giemsa, and fluorescent banding (Borzan and Papes 1978; MacPherson and Filion 1981; Drewry 1982; Saylor 1983; Hizume et al. 1989, 1990). More recently, fluorescent in situ hybridization (FISH) has been utilized in several pine species (Doudrick et al. 1995; Lubaretz et al. 1996; Ja-

cobs et al. 2000; Hizume et al. 2002; Liu et al. 2003; Cai et al. 2006). Doudrick et al. (1995) developed a FISH-based karyotype for *P. elliotii* var. *elliotii* using 18S-25S and 5S rDNA probes and CMA (chromomycin A₃) and DAPI (4',6-diamidino-2-phenylindole) banding that distinguished all 12 homologous pairs of chromosomes. They went further to suggest that the presented karyotype might be useful as a standard or reference karyotype for *Pinus*. Lubaretz et al. (1996) used computer-aided chromosome analysis on the basis of chromosome length, chromosome arm length ratio, and the positions of rDNA (18S-28S and 5S) and telomere (*Arabidopsis*-type telomere repeat sequence or A-type TRS) detected with FISH to discriminate three chromosomes of *P. sylvestris*. Hizume et al. (2002) used four probes [45S rDNA, 5S rDNA, PCSR (CMA-band specific repeat), and A-type TRS] in developing FISH-based karyotypes for four different pine species (*P. densiflora*, *P. thunbergii*, *P. sylvestris*, and *P. nigra*). Liu et al. (2003) and Cai et al. (2006) used FISH to establish rDNA positions in several species of the *Pinus* and *Strobis* subgenera, respectively. Current work in *P. taeda* (subgenera *Pinus*, section *Pinus*, subsection *Australes*) (Fig. 1) emphasizes an improved chromosome preparation technique (based on Jewell and Islam-Faridi 1994 and Islam-Faridi and Mujeeb-Kazi 1995) and statistical analyses of chromosome arm lengths and FISH signal positions and intensities to develop a reference karyotype and cy-

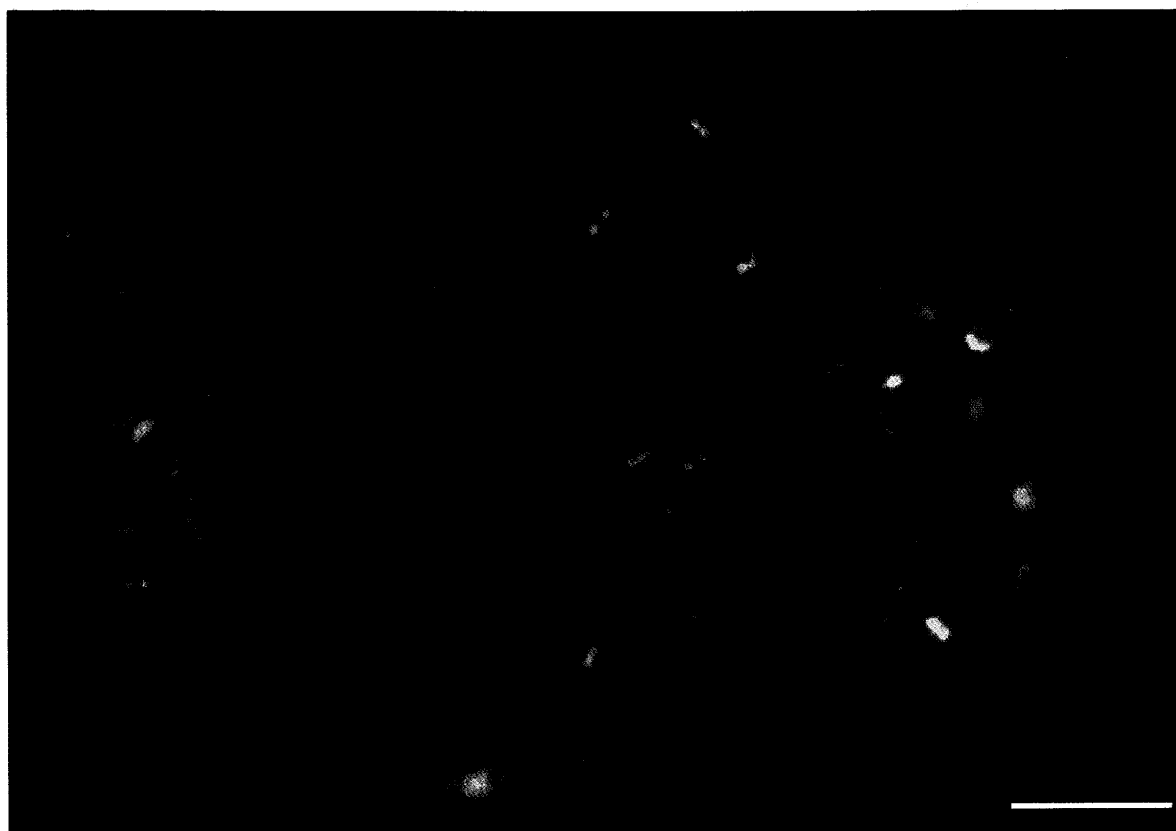


Fig. 1. A fluorescent in situ hybridization (FISH) photomicrograph of *Pinus taeda* metaphase chromosomes showing 23 of the 24 chromosomes (bar = 10 μ m). The red signals detect the 18S-28S rDNA sites, and the green signals detect the *Arabidopsis*-type telomere repeat sequence (A-type TRS) sites

togenetic map for use in physical genome mapping in the subsection *Australes* (Islam-Faridi et al. 2003, Islam-Faridi et al. 2007). Comparison of these results with those obtained in other subsections suggests that a subsection-specific karyotype may be required for more robust physical mapping across the entire genus as a whole (Islam-Faridi et al. 2007).

Another feature of the pine genome is its large physical genome size expressed in DNA content. Pines exhibit some of the largest DNA contents per diploid cell in the plant kingdom ranging from ca. 44 pg (*P. banksiana*) to 75 pg (*P. gerardiana*) (1 pg = 960 Mb; Arumuganathan and Earle 1991) based on laser flow cytometry (Grotkopp et al. 2004). For instance, the size of the maritime pine (*P. pinaster*) genome has been estimated to be between 51 and 60 pg/2C (Chagné et al. 2002; Grotkopp et al. 2004), which is about seven times the size of the human genome (7 pg/2C; Morton 1991) and 170-fold larger than the genome of model plant *Arabidopsis thaliana* (0.3 pg/2C). The DNA amount varies according to the

subgenus and the section. The genome of subgenus *Pinus* is lower (54.0 pg/2C) than that of subgenus *Strobus* (65.6 pg/2C). This variation has been related to seed mass. Relationships of such variation with ecological conditions, such as drought tolerance, have also been investigated (Wakamiya et al. 1996). Various environmental conditions appear to be related to DNA content, including latitude of range and invasiveness (Grotkopp et al. 2004). An extremely large genome size is common to other gymnosperms (reviewed by Ohri and Khoshoo 1986; Murray 1998; Leitch et al. 2001; Grotkopp et al. 2004).

At the whole-genome level, reassociation kinetics data (i.e., Cot analysis) indicates that 25 to 30% of the pine genome corresponds to low- to single-copy sequences, while 70 to 75% corresponds to highly repeated sequences (Miksche and Hotta 1973; Rake et al. 1980; Kriebel 1985; Peterson et al. 2006). The repetitive sequences of pine have not been studied in much detail. However, it appears that the majority of repetitive DNA consists of repeats of low sequence complex-

ity (Schmidt et al. 2000), retrotransposons (Kamm et al. 1996; Kossack and Kinlaw 1999; Friesen et al. 2001), and 18S-5.8S-25S rDNA genes. As shown by molecular, cytological, and linkage-mapping studies (Friesen et al. 2001; Scotti et al. 2005), most of these repeat sequences are present at multiple loci and are presumably interspersed among other sequences, although they tend to form loose clusters that surround gene-rich islands. The remainder of the genome is composed of low-copy sequences of which an unknown portion are genes. If the pine genome (1C ~ 25 to 30 pg; see above) is similar to *Arabidopsis* in having about 30,000 expressed genes and an average gene size of 2,000 bp including introns and UTRs (The *Arabidopsis* Genome Initiative 2000), only 0.5% of its genome is likely to be transcribed. In comparison, 54% of the *Arabidopsis* sequence and 5 to 6% of mammalian genomes are transcribed (Rat Genome Sequencing Project Consortium 2004).

Gene duplication and the formation of complex gene families have been widely cited as a potential cause of the abundance of low-copy DNA in the pine genome. Kinlaw and Neale (1997) suggested that levels of multiplication were greater in conifers than in other plant species, and transcriptional profiling studies have noted surprising levels of transcriptome complexity (Lorenz and Dean 2002). However, considering the relatively low proportion of a conifer genome represented by expressed sequences, it is unlikely that the evolution of multigene families alone can explain the enormous size of pine nuclear genomes. It is possible that a relatively large proportion of low-copy sequences in pine are pseudogenes and/or highly diverged repeat sequences, and indeed there is evidence supporting both possibilities (Elsik and Williams 2000; Rabinowicz et al. 2005).

2.1.3

Economic Importance

Pine species play an especially important role in modern plantation forestry worldwide and now form a large part of both the annual wood harvest and the immature plantation forests that will provide wood in the future (Burdon 2002). Pines enjoy such great popularity because the large number of species allows choice for widely varying site and environmental conditions; the volume of production of some species can be high to very high, even under unfavorable site conditions; they are well suited for reforestation and for

simple silviculture (monocultures and clear-felling); their wood is easily processed and utilized for a wide variety of end uses (lumber, pulp and paper, particleboard, etc.); and even if species lack naturally durable heartwood, treatment with preservatives is easy.

Various minor forest products come from pines (Burdon 2002). Some pine plantations, especially *P. pinaster* in the Landes of southwest France, were established largely for resin production. Resin production from pines was also a major economic activity in the southeast of the USA, Mediterranean basin, northern India, and east and Southeast Asia. Resin products are still recovered from chemical pulping of pines. Foliage, in the form of litterfall, has been used for a range of purposes. The bark is used for a variety of purposes, and that of many species has a high tannin content. For a few species with large seeds, notably *P. pinea*, *P. edulis*, and *P. sibirica*, seeds have been a prized item of the human diet. Edible fungi, representing the fruiting bodies of mycorrhizal symbionts, are often collected from pine stands.

Pine species are also used widely for shelter and the interlinked purposes of revegetation, soil stabilization, and soil conservation, as befits their tolerance of exposure and degraded soils in certain species. Most of the widely planted species of pines are used in some degree for one or more of these purposes, often with timber production as a major bonus. Ornamental and festive use is common, with *P. sylvestris* and *P. virginiana* being very widely grown for Christmas trees.

2.1.4

Classical Breeding Objectives

In most pine breeding programs, the setting of goals was originally done subjectively, based on perceptions of the main traits limiting profitability, of the variability and heritability of the traits concerned, and of the genetic correlations (both favorable and adverse) among traits (Burdon 2004). The setting of breeding goals is crucial to any tree-improvement program, and yet most pine tree breeding programs do not have formally defined breeding objectives. Various reasons for this include complexity of the forest processing industry, difficulties in determining the relationships between selection traits and end uses, and long rotation ages that create uncertainty about their use (Apiolaza and Greaves 2001). Earlier attempts of most improvement programs have been to improve growth, form,

climatic adaptation, and resistance to certain diseases (Cahalan 1981; Danjon 1995; Paul et al. 1997; Shelbourne et al. 1997).

Most recent efforts have been diverted at improving traits related to pulp and paper and solid-wood products (Shelbourne et al. 1997; Chambers and Borralho 1999; Greaves 1999; Lowe et al. 1999; Pot et al. 2002; Kumar 2004). Wood density and fiber morphology (e.g., tracheid length and coarseness) have been reported to be crucial for pulp yield and quality. Wood properties such as wood density and stiffness are crucial selection traits for the improvement of wood stiffness, while compression wood, spiral grain, and microfibril angle are candidate traits for reducing in-service instability (Shelbourne et al. 1997; Ivkovic et al. 2006). Heartwood, resinous defects, and internal checking are also being considered as selection traits to develop germplasm for appearance-grade products (Shelbourne et al. 1997). There are some situations, especially involving exotic species and disease resistance, etc., where hybrids between selective species are desirable to make genetic improvement in the breeding-objective traits (Hyun 1976; Byun et al. 1989; Blada 1994; Nikles 2000; Shelbourne 2000). Pine species vary widely in their amenability to various forms of vegetative propagation (Hartmann et al. 1990). Ease of propagation was generally a minor criterion for species selection in breeding programs of pines. Currently with improved technology, selection is based more on silvicultural performance and wood quality.

2.1.5 Classical Breeding Achievements

Various selection and deployment strategies are being used for different pine species. Species selection followed by provenance and family-within-provenance selection, and establishment of seed orchards are quite common first steps across various species including *P. radiata* (Falkenhagen 1991; Matziris 1995), *P. oocarpa* (Moura et al. 1998), *P. caribaea* (Zheng et al. 1994), *P. strobus* (Beaulieu et al. 1996), *P. sylvestris* (Quencez and Bastien 2001), *P. pinaster* (Alía et al. 1995, 1997; Danjon 1995), *P. taeda* (Jiang et al. 1999; Lopez-Upton et al. 2000), *P. tecunumanii* (Hodge and Dvorak 1999), and *P. contorta* (Cahalan 1981). In advanced-generation breeding programs, forward selections and/or combined selection (among- and within-family) are the major sources of

genetic gain (Wei et al. 1997; Rosvall et al. 1998; Lambeth 2000; Alazard 2001; Olsson et al. 2001; Plomion et al. 2001; Burdon and Kumar 2004). In the species that are easy to propagate, the concept of using clonal replication of individuals within families as a means of genetic testing is being implemented in order to increase the efficiency of genetic improvement. Significant genetic gains from such a strategy have been reported for *P. radiata* (Matheson and Lindgren 1985; Shelbourne 1992) and *P. taeda* (Isik et al. 2004).

Pinus patula, *P. taeda*, and *P. elliottii*, are planted in South Africa in summer rainfall zones, and their average productivity is $15 \text{ m}^3 \text{ ha}^{-1} \text{ yr}^{-1}$ (Du Toit et al. 1998). Brazil, with its humid summers, largely uses *P. caribaea*, *P. taeda*, and *P. elliottii*. Pines in Brazil produce 8 to $30 \text{ m}^3 \text{ ha}^{-1} \text{ yr}^{-1}$ on rotations of 20 to 25 years. *P. radiata* is grown as large plantations in the temperate southern hemisphere countries of Australia, Chile, New Zealand, and South Africa. The mean annual increment over 25 years is often 25 to $30 \text{ m}^3 \text{ ha}^{-1} \text{ yr}^{-1}$ (Lamprecht 1990). However in New Zealand, growth rates of up to $50 \text{ m}^3 \text{ ha}^{-1} \text{ yr}^{-1}$ have been recorded on the best sites and as low as $11 \text{ m}^3 \text{ ha}^{-1} \text{ yr}^{-1}$ on very dry sites (Burdon and Miller 1992). The average productivity of *P. pinaster* in southern France is about $10 \text{ m}^3 \text{ ha}^{-1} \text{ yr}^{-1}$ but could reach 20 to $25 \text{ m}^3 \text{ ha}^{-1} \text{ yr}^{-1}$ on the best sites. Deployment of genetically improved loblolly pine in the USA has been reported to yield up to about $21 \text{ m}^3 \text{ ha}^{-1} \text{ yr}^{-1}$ (McKeand et al. 2003). Dhakal et al. (1996) reported a realized gain in volume of about 22% in a slash pine improvement program in the USA. Wood-quality traits that are currently being included, in addition to growth, form, and health traits, in the breeding objectives of various pines species appear to be under moderate to strong genetic control (Burdon and Low 1992; Hannrup et al. 2000; Atwood et al. 2002; Gwaze et al. 2002; Pot et al. 2002; Kumar 2004), and predicted genetic gains from selection appeared to be in the order of about 10% for traits such as wood density and stiffness (Kumar 2004).

Clonal forestry (CLF) represents the large-scale propagation and deployment of selected clones that have been clonally tested. Deployment of tested clones by CLF is being increasingly employed with *P. radiata* in New Zealand (Sorensson and Shelbourne 2005), *P. taeda* in the USA (Stelzer and Goldfarb 1997), and pine hybrids in Australia (Walker et al. 1996). In principle, CLF offers additional genetic gains from capturing nonadditive effects, which are not captured via sexual propagation, plus the benefits of greater uni-

formity and predictability in performance resulting from a lack of genetic segregation. Genetic gains from CLF have been predicted to be considerably higher than those from family forestry in *P. radiata* (Aimers-Halliday et al. 1997), *P. taeda* (Stelzer and Goldfarb 1997), and *P. strobus* (Park 2002).

2.1.6

Molecular Diversity

The majority of studies aiming to monitor the level and distribution of genetic diversity in *Pinus* natural populations were based on the use of neutral markers. Indeed, molecular markers, such as microsatellites, provided useful information on historical demography and population evolution.

The use of isozyme electrophoresis significantly increased the amount of data on the genetic structure of populations (Petit et al. 2005). These data revealed that pine species had high genetic diversity within populations and only low levels of differentiation among populations. For 28 north temperate pines, genetic differentiation (G_{ST}) averaged 0.076 (Ledig 1998). Exceptions are represented by *P. pinea* (G.G. Vendramin et al. personal communication), *Pinus resinosa*, a species that has a vast range across northeastern North America (Echt et al. 1998; Walter and Epperson 2001), and *P. torreyana* (Ledig and Conkle 1983; Provan et al. 1999) where near absence of variation was observed, and by Mexican pine species where higher differentiation among populations was observed, probably because their natural distributions are more highly fragmented by physiography than those of species at more northerly latitudes. Other pine species with disjunct populations and restricted gene flow also showed higher differentiation among populations: for example, 16 to 27% for *P. radiata* and 22% for *P. muricata* (Wu et al. 1999). On the other hand, experimental evidence indicates that seeds of some pines (e.g., *P. palustris*; Grace et al. 2004) have the potential to disperse greater distances than previously reported, which partly contributes to the low levels of genetic differentiation observed in these species.

In general, the typical distribution of the genetic diversity within and among populations of *Pinus* species is correlated with their mating system and life history (pines, for example, are wind-pollinated and tend to be predominately outcrossing) (Hamrick and Godt 1996; J. Duminil et al. personal communication)

and biogeographic history (the distributions of many species have been affected by Pleistocene glacial advances), even if, in some cases, human activities also played a relevant role (e.g., *P. pinaster* in Portugal; Ribeiro et al. 2001).

Self-fertilization in pines generally occurs at a low level (Muona and Harju 1989), and a high outcrossing at the mature seed stage is maintained. There is evidence of selection at the embryonic stage so that the number of inbreds is already low at the seedling stage (Kärkkäinen and Savolainen 1993). Selection after the seedling stage is still severe. This has been shown by observing the survival of selfed seedlings in *P. sylvestris* (Muona et al. 1987) and in *P. leucodermis* (Morgante et al. 1993).

The mode of inheritance has a major effect on the partitioning of genetic diversity in pines, with studies based on maternally inherited markers (transmitted by seeds only) having significantly higher G_{ST} values than those based on paternally or biparentally inherited markers for pine (Burban and Petit 2003; Petit et al. 2005). In fact, the chloroplast and mitochondrial genomes are generally paternally and maternally inherited in pines, respectively (Petit and Vendramin 2006). In pines, G_{ST} is nearly always larger at mitochondrial DNA markers than at chloroplast DNA markers. On the other hand, there is no significant difference between G_{ST} at biparentally inherited markers and at paternally inherited markers in pines (Petit et al. 2005). This is expected considering that both the cpDNA and half the nuclear genomic complement are dispersed by pollen and by seeds, i.e., they use the same vehicles to achieve gene flow.

Chloroplast and mitochondrial DNA markers allow describing phylogeographic structure in pines. While chloroplast DNA generally exhibits the highest diversity, phylogeographic inferences from these markers can be blurred by extensive pollen flow. Low population structure due to extensive pollen flow has been inferred in *P. pinaster* (Burban and Petit 2003), *P. sylvestris* (Robledo-Arnuncio et al. 2005), *P. canariensis* (Gómez et al. 2003), and *P. albicaulis* (Richardson et al. 2002). In contrast, the mitochondrial markers, despite their generally lower level of diversity in pines (but this holds for all conifers; Soranzo et al. 2000; Gugerli et al. 2001), generally provide a clear picture of nonoverlapping areas colonized from different refugia (e.g., in *P. pinaster*; Burban and Petit 2003).

Neutral markers also have allowed investigating spatial genetic structure (SGS) in natural pine popu-

lations. A generally weak within-population structure has been described. *P. pinaster* showed a fine-scale structure at the seedling stage with a patch size of ca. 10 m that seems to be produced by restricted seed flow (González-Martínez et al. 2002). Pines with a heavy seed (differences in seed dispersion capability play an important role), such as *P. pinaster*, are expected to have a short dispersal distance, thus producing a fine-scale structure. However, fine-scale structure often does not persist as stands mature. For example, within-population genetic structure in Mediterranean pines may be affected by postdispersal events (e.g., mortality due to the severity of the Mediterranean climate and animal-mediated secondary dispersal during the summer period) that may modify the original spatial structure (González-Martínez et al. 2002). Logging can also play a role in decreasing spatial structuring, as observed in *P. strobus* (Marquardt and Epperson 2004), suggesting that management practices can alter natural spatial patterns, too.

It should be stressed that the presence of fine-scale structure is uncommon in *Pinus* species. Epperson and Allard (1989), studying the spatial pattern of allozyme alleles within *P. contorta* ssp. *latifolia* stands, found a lack of structure in the distributions of most genotypes. Neutral markers by definition do not reflect selective processes and therefore are not used as an indicator of the population adaptive potential to a changing environment (Morin et al. 2004). Recent and well-established markers to detect functional genetic variation are single nucleotide polymorphisms (SNPs), which are particularly useful for finding genes under selection and studying the dynamics of these genes in natural populations.

The availability of high-density markers, such as SNPs, opens the possibility of studying, by association genetics, the molecular basis of complex quantitative traits in natural populations, taking advantage of the fact that genetic markers in close proximity to causal polymorphisms may be in linkage disequilibrium (LD) to them. The magnitude and distribution of LD determine the choice of association mapping methodology. Extension and distribution of LD depend on many factors including population history (e.g., the presence of population bottlenecks or admixture) and the frequency of recombination.

In order to avoid false associations, the optimization of LD mapping requires a detailed knowledge of

basic population genetic parameters such as the pattern of nucleotide diversity and LD for each particular species and candidate gene set.

First estimates indicate that nucleotide diversity varies considerably between plant species. Interestingly, the pines (e.g., *P. sylvestris*, Dvornyk et al. 2002; García-Gil et al. 2003; *P. taeda*, Brown et al. 2004; González-Martínez et al. 2006a; *P. pinaster*, Pot et al. 2005a) are not among the most variable species, contradicting expectations from the results obtained using neutral markers and their life history characteristics. First evidences seem to show that broadleaved species (e.g., *Populus*, Ingvarsson 2005; *Quercus*, Table 2.7 in Pot et al. 2005a) display higher levels of nucleotide diversity than pines.

Markers in specific functional regions of the genome need to be statistically analyzed in order to test for the possibility that these regions might have experienced different selective pressures. In unstructured populations, standard neutrality tests might be applied. When variation is structured in populations, a relatively easy approach is the comparison of genetic differentiation estimates, such as Wright's *F*-statistics, among markers tagging a putative gene under selection and neutral markers or expected distributions computed using coalescence theory (see reviews in van Tienderen et al. 2002; Luikart et al. 2003). If population divergence (F_{ST}) is higher for the gene-targeted marker with respect to divergence estimates obtained from random markers, this might indicate divergent selection and local adaptation for the tagged gene (van Tienderen et al. 2002). Pot et al. (2005a) found a high differentiation among populations at *Pp1* (*glycine-rich protein* homolog) gene in *P. pinaster*, higher than at the neutral level. This result is consistent with diversifying selection acting at this locus in this species. On the other hand, the absence of differentiation observed for the gene *CeA3* (*cellulose synthase*), compared with the significant level observed at neutral markers may indicate balancing selection acting on this gene.

Recent studies on pines reveal a rapid decay in LD with physical distance. LD declines very rapidly within 200 to 2,000 bp in *Pinus taeda* (Brown et al. 2004; González-Martínez et al. 2006a), *Pinus sylvestris* (Dvornyk et al. 2002; García-Gil et al. 2003). A rapid decay of LD in pines is consistent with what is expected from outcrossing species with large effective population size.

2.2

Construction of Genetic Maps

In genetics, mapping is defined as the process of deducing schematic representations of DNA. Three types of DNA maps can be constructed depending on the landmarks on which they are based:

- Physical maps, whose highest resolution would be the complete nucleotide sequence of the genome
- Genetic maps, which describe the relative positions of specific DNA markers along the chromosomes, determined on the basis of how often these loci are inherited together
- Cytogenetic maps, a visual appearance of a chromosome when stained and examined under a microscope

To provide a first glimpse of the pine genome, high-resolution genetic maps have been established for several pine species using different types of molecular markers and following different strategies that are reviewed in the following sections.

2.2.1

Development of Molecular Markers in Pines

The construction of a linkage map relies on the availability of enough molecular markers to detect linkage between them. Each type of marker technology has advantages and limitations. Many factors (e.g., polymorphism information content, level of polymorphism exhibited for the mapping progeny, mode of inheritance, genome size) can influence the development of a particular technique and the choice of a marker system for a given purpose (e.g., genetic mapping, quantitative trait loci analysis, survey of genetic diversity, forensic applications). The purpose of this section is to briefly review the different types of molecular marker techniques that have been developed in pines and used for genetic mapping applications. We will not present the details of each technique. Both the review by Cervera et al. (2000a) and the references cited in Table 2 will provide the reader with the necessary information for understanding the scientific basis of each technique.

Isozymes and Proteins

The first markers developed for pine were isozymes. Linkage studies were carried out on more than 10 species for about 15 loci (reviewed by Tulsieram et al.

1992). Conkle (1981) located more loci, but still not enough to cover the pine genome. Proteins revealed by two-dimensional polyacrylamide gel electrophoresis (2D-PAGE; O'Farrell 1975) presented the advantage of being multiplexed compared to isozymes. Importantly, proteins can be easily characterized by mass spectrometry (e.g., Gion et al. 2005) and may be recognizable by sequence similarity to others proteins published in sequence databases, therefore providing functional markers expressed in the tissues analyzed. Two-dimensional protein markers were developed only in *P. pinaster* (reviewed in Cánovas et al. 2004 and Plomion et al. 2004). Although proteins provided physiologically relevant markers to map the expressed genome, this time-consuming technique failed to provide enough markers for genetic application, which requires full genome coverage, such as linkage mapping and quantitative trait loci (QTL) detection.

RFLPs

Restriction fragment length polymorphism (RFLP) markers were developed in the early 1990s for *P. taeda* (Neale and Williams 1991). They offered a sufficient number of markers for high-density genome mapping in pine. However, this labor-intensive and time-consuming technique was only applied to *P. taeda* (Devey et al. 1994) and *P. radiata* (Devey et al. 1996).

RAPDs and AFLPs

In the mid-1990s PCR-based multiplex DNA fingerprinting techniques provided very powerful tools to generate dense linkage maps in a short period of time. Random amplified polymorphic DNA (RAPD; Williams et al. 1990) and then amplified fragment length polymorphism (AFLP; Vos et al. 1995) became the most popular marker technologies in conifers. Despite their biallelic nature and dominant mode of inheritance (which was actually not an issue for mapping with haploid megagametophyte, see Sect. 2.2.2.), these markers tremendously boosted up genetic analysis in most forest tree species including pines (reviewed in Cervera et al. 2000b).

Nuclear Microsatellites

In contrast to other plant species, few polymorphic single-copy nuclear microsatellite markers or simple sequence repeats (SSRs) have been reported in pines (reviewed in Chagné et al. 2004). The genome

Table 2. Characteristics and applications of molecular markers

Feature	Protein markers		DNA markers				
	Isozyme	2D-PAGE	Hybridization-based RFLP	PCR-based RAPD	AFLP	SSR	SNP
Tissue quantity (g)	0.5–2	1	na	na	na	na	na
DNA quantity (mg)	na	na	10	0.02	0.25–0.5	0.02	0.005–0.02
DNA quality	na	na	high	Medium	Medium	Medium	Medium
Allelism and mode of inheritance	Multiallelic codominant	biallelic, codominant, or dominant	Biallelic codominant	Biallelic dominant	Biallelic dominant	Multiallelic codominant	Biallelic codominant
Number loci analysed per assay	1–5	several tens	1–6	2–30	20–100	1–10 (multiplexing)	1–several hundred (multiplexing)
Polymorphism	Low	Low	Medium	Medium	Medium	High	Medium ^a
Development cost	Low	Low	High	Low	Medium	High	High
Cost per analysis	Low	Medium	High	Low	Medium	Low	Low
Amenable to automation	Low	Low	Low	Medium	Medium	High	High
Technical demand	Low	High	High	Low	Medium	Low	Medium
Reproducibility	High ^b	Medium ^b	High	Low–high ^c	High	High	High
Review	(Müller–Starck 1998)	(Plomion et al. 2004)	(Brettschneider 1998)	(Rafalski 1998)	(Cervera et al. 2000b)	(Echt and Burns 1999)	(Syvanen 2001)
Applications							
Certification	-	-	-	-	+	+++	++
Diversity	+	+	++	+	+	+++	+++
Phylogeny	++	+	++	++	++	++	+++
Mapping	-	-	++	++ ^d	++ ^d	+++	+++
Comparative mapping	+	+	++	-	-	+++	+++
QTL analysis	+	+	++	+	+	+++	+++
Association studies	+	+	+	-	-	++ ^e	+++

^a Sequence dependent (exon vs UTR and intron)^b Under the same environmental conditions, type of tissue, and age^c Between laboratories vs single laboratory^d To construct dense maps^e Used as control loci

structure of these species, characterized by a large physical size, with a large amount of repeated sequence (Sect. 2.1.2), has been the main obstacle to the development of useful markers using classical SSR-enriched library approaches (e.g., Auckland et al. 2002; Guevara et al. 2005a; C.S. Echt and C.D. Nelson, unpublished results). In addition, the ancient divergence time between coniferous species (Price et al. 1998) and the complexity of their genomes means that transferability of single-copy SSRs among genera and even within *Pinus* is generally poor, resulting in a large proportion of amplification failure, nonspecific amplification, multibanding patterns, or lack of polymorphism (Echt and Nelson 1997; Mariette et al. 2001). In an attempt to circumvent these genome-related problems, Elsik and Williams (2001) removed most of the repetitive portion of the genome using a DNA reassociation kinetics-based method, and Zhou et al. (2002) targeted the low-copy portion of the genome using an undermethylated region enrichment method. Both approaches yielded remarkable enrichment for useful SSR markers in *P. taeda*. SSRs made from low-copy, undermethylated, and total genomic DNA yielded mappable markers (Nelson et al. 2003; Zhou et al. 2003). *P. taeda* SSRs developed by Elsik and Williams (2001) and Zhou et al. (2002) transferred quite well between American hard pines (Shepherd et al. 2002a) but were shown to be less transferable in the phylogenetically divergent Mediterranean hard pines (Chagné et al. 2004; González-Martínez et al. 2004). Interestingly, perfect trinucleotide SSRs transferred from American to Mediterranean pines better than other motifs (Kutil and Williams 2001). A number of nuclear SSR markers have been developed for *P. radiata*, almost all of which are based on the more frequently polymorphic dinucleotide repeat motifs (Smith and Devey 1994; Fisher et al. 1996, 1998; Devey et al. 2003), and used in a number of applications. Polymorphic chloroplast microsatellite loci have also been identified and applied (Cato and Richardson 1996; Kent and Richardson 1997). More recent SSR discovery efforts have been undertaken in both New Zealand and Australia and have been most commonly applied to fingerprinting (Kent and Richardson 1997; Bell et al. 2004) and QTL mapping applications (Devey et al. 2004a).

EST Polymorphisms

With the availability of sequence data obtained by random sequencing of pine cDNAs (Sect. 2.5.2), there is now a clear trend toward the development of gene-

based markers (ESTP: EST polymorphisms). There are basically two groups of technologies used to detect nucleotide polymorphisms (SNPs and INDELs), either based on the knowledge of nucleotide variants or not. Up to now, techniques based on the detection of differences in the DNA stability (denaturing gradient gel electrophoresis, DGGE; Myers et al. 1987), conformation (single-strand conformation polymorphisms, SSCPs; Orita et al. 1989) under specific polyacrylamide gel conditions, or heteroduplex cleavage (TILLING, targeting induced local lesions in genomes; Colbert et al. 2001) have been successfully applied in pines (Plomion et al. 1999; Temesgen et al. 2001; Chagné et al. 2003; Ritland et al. 2006). With the decrease of sequencing costs and the availability of pine cDNA sequences, more targeted and precise approaches are now possible (Pot et al. 2005b). The bioinformatics assembly of ESTs into large contigs (i.e., unigenes) has also made it possible to identify putative SNPs. Le Dantec et al. (2004) identified a set of 1,400 candidate SNPs in *P. pinaster* contigs containing between 4 and 20 sequence reads. This represents a great resource of molecular markers for this species that can be used to map candidate genes, study LD, and develop comparative orthologous sequence markers for comparative genome mapping. In addition to SNPs and INDELs, a large set of microsatellite markers have been developed from *P. taeda* and *P. pinaster* expressed sequence tags (ESTs) (Echt and Burns 1999; Chagné et al. 2004; Echt et al. 2006). These markers present the advantage of being highly polymorphic and located in coding regions.

2.2.2

Haploid- and Diploid-Based Mapping Strategies

The construction of a genetic map requires two components: first, a segregating population (mapping pedigree) derived from a cross between parental trees that are heterozygous for many loci and, second, a set of molecular markers segregating in the progeny according to Mendelian ratios. Linkage map construction is based on the statistical analysis of polymorphic markers in the mapping population, considering that the distance between two loci is related to the probability of observing a recombination event between them. There is a number of mapping software to facilitate automated analysis (<http://linkage.rockefeller.edu/soft/list.html>).

Haploid- or Half-Sib-Based Mapping Strategy

In conifers, the haploid megagametophyte constitutes an ideal plant material for genetic mapping. This nutritive tissue surrounding the embryo is derived from the same megaspore that gives rise to the maternal gamete. Therefore, it represents a single meiotic event in the parent tree that is genetically equivalent to a maternal gamete. The dominant nature and biallelic mode of inheritance of RAPD and AFLP is not an issue for genetic mapping with haploid megagametophytes. However, quantitative traits can only be measured on half-sib seedlings, limiting the detection of QTLs at the first stages of tree development. Thus, this approach is not applicable to the analysis of QTLs for economically important traits in well-established plantations, unless the megagametophytes were collected and saved, which has generally not been the case.

Diploid- or Full-Sib-Based Mapping Strategy

Different strategies have been followed to construct genetic linkage maps of *Pinus*: the “pseudotestcross strategy,” the “F₂ inbred model,” and the “three-generation outbred model.” The pseudotestcross strategy is mainly based on selection of single-copy polymorphic markers heterozygous in one parent and homozygous null in the other parent and therefore segregating 1:1 in their F₁ progeny as in a testcross. Grattapaglia and Sederoff (1994) introduced the term two-way pseudotestcross to define this mapping strategy, where two independent genetic linkage maps are constructed by analyzing the cosegregation of markers in each progenitor. The efficiency of this strategy, as well as for the haploid mapping strategy, depends on finding individual trees that are heterozygous for many loci, which is quite easy using arbitrarily primed PCR assays (RAPD and AFLP) in highly heterozygous outcrossed tree species such as pines. The F₂ inbred model is based on a three-generation pedigree for which the grandparents are treated as inbred lines. In the F₂ generation, three genotypes occur at any locus – AA, AB, and BB – segregating 1:2:1. The three-generation outbred model (Sewell et al. 1999) is an extension of the pseudotestcross strategy. Within a single outbred pedigree, any given codominant marker will segregate in one of three different ways. When one parent is heterozygous and the other is homozygous, segregation will be 1:1 (i.e., testcross mating type). When both parents are heterozygous, segregation will be either 1:2:1, if both parents have the same genotype

(i.e., intercross mating type), or 1:1:1:1, if they have different genotypes (i.e., fully informative mating type). These segregation data are then subdivided into two independent data sets that separately contain the meiotic segregation data from each parent, and independent maps are constructed for each parent. A sex-average map is then constructed using an outbred mapping program, which uses fully informative and intercross markers to serve as common anchor points between each parental data set. Compared to “megagametophyte progeny,” full-sibs can be grafted and/or propagated by cuttings, thereby constituting a perpetual population, analogous to recombinant inbred lines in crop plants. The use of such clonally propagated progeny obviously increases the precision of quantitative measurements and therefore enhances the QTL detection power (Bradshaw and Foster 1992).

2.2.3

Genetic Mapping Initiatives in Pines

In this section and Table 3 we summarize what has been done in terms of linkage map construction in the genus *Pinus* with emphasis made on the most studied species. In addition, some maps have been published together with QTL studies and will be found in the references cited in Sect. 2.3.

Maritime Pine

Linkage maps of the Maritime pine (*Pinus pinaster* Ait.) genome were first constructed by analyzing the cosegregation of proteins extracted from megagametophytes collected during the germination of the embryo. Bahrman and Damerval (1989) were the first to report a linkage analysis for 119 protein loci using 56 megagametophytes of a single tree. Extending this approach, Gerber et al. (1993) reported a 65-locus linkage map covering one fourth of the pine genome, using 18 maritime pine trees with an average of 12 megagametophytes per tree. A more conventional pedigree (inbred F₂) was used to map 61 proteins using haploid (Plomion et al. 1995a) and diploid (Plomion et al. 1997; Costa et al. 2000) tissues of the same seedlings. In the latter case, protein loci were found on each chromosome (Thiellement et al. 2001). As stated above, the advance of PCR-based markers has allowed the construction of saturated linkage maps, in a short period of time, with no prior knowledge of DNA sequence (Plomion et al. 1995a, b; Costa et al. 2000).

Table 3. Summary of genetic linkage maps of *Pinus* species

Species pedigree	Number of LGs (+pairs, triplets)	Total number of linked marker loci	RAPD, AFLP	SSR	EST-based	Others	Total genetic distance (cM)	Genome saturation	References
<i>Pinus pinaster</i>									
Haploid (1 tree)	Not estimated	119	–	–	–	119 (2D proteins)	Not estimated	Not estimated	Bahrman and Damerval 1989
Haploid (8 trees)	Not estimated	65	–	–	–	65 (2D proteins)	530	25%	Gerber et al. 1993
Haploid (H12 self)	13(+5)	263	251	–	–	–	1223	90%	Plomion et al. 1995b
Haploid (H12 open pollinated)							1236	90%	
Haploid (H12 self)	12	463	436	–	–	27 (2D proteins)	1860	100%	Plomion et al. 1995a
Haploid (H12self)	11	94	94	–	–	–	1169	Not estimated	Plomion and O'Malley 1996
Diploid (H12 self)	11	94	94	–	–	–	1354	Not estimated	
Haploid (H12 self)	13	398	235+127	–	–	36 (2D protein)	1873	93.4%	Costa et al. 2000
F1 - AFOCEL	12	759	738	14	7	–	1994	Not estimated	Ritter et al. 2002
9.103.3 × 10.159.3 (consensus)	12	620	620	–	–	–	1441	Not estimated	Chagné et al. 2002
9.103.3 × 10.159.3 (consensus)	12	326	276	–	50	–	1638	Not estimated	Chagné et al. 2003
<i>Pinus taeda</i>									
Various pedigrees	Not estimated	–	–	–	–	20 isozymes	Not estimated	Not estimated	Conkle et al. 1981
Base pedigree	20	75	–	–	90	6	Not estimated	Not estimated	Devey et al. 1994
Haploid (Tree 10-5)	16	458	458	–	–	–	1727	97%	Wilcox 1995
Base and QTL pedigrees	18	357	67	–	257	12	1300	76.4%	Sewell et al. 1998
Tree 7-56	12	508	508	–	–	–	1741	100%	Remington et al. 1999
Base and QTL pedigrees	20	265	253	12	–	–	1281	75.4%	Devey et al. 1999
Base pedigree	15	51	–	51	–	–	795	46.7%	Zhou et al. 2003
<i>Pinus sylvestris</i>									
49-2	14	261	261	–	–	–	2638		Yazdani et al. 2003
F1 of P315 × E1101	12(+4)	179	179	–	–	–	1000	50%	Hurme et al. 2000
F1 of AC3065 × Y3038	♀: 12	188	188	–	–	–	1645	98%	Yin et al. 2003
	♂: 12	245	245	–	–	–	1681		
F1 of E635 × E1101	12(+3)	260	194	4	61	–	1314	66–85%	Komulainen et al. 2003

Table 3. (continued)

Species pedigree	Number of LGs (+pairs, triplets)	Total number of linked marker loci	RAPD, AFLP	SSR	EST-based	Others	Total genetic distance (cM)	Genome saturation	References
<i>Pinus radiata</i>									
3-generation	22	208	41	2	165	–	1382	Not estimated	Devey et al. 1996
Haploid (full-sib seed)	14	267	267	–	–	–	1665	93%	Emebiri et al. 1998
S1	19	172	168	4	–	–	1117	56%	Kuang et al. 1999b
Pseudotestcross (Parent 850.055)	20	235	224	11	–	–	1414	85%	Wilcox et al. 2001a
Pseudotestcross (Parent 850.096)	21	194	185	9	–	–	1144	77%	Wilcox et al. 2001a
Two full-sib families	12	311	–	213	98	–	1352	Not estimated	Wilcox et al. 2004
<i>Pinus elliottii</i>									
Tree 8-7	13(+9pairs)	73	73	–	–	–	782	64–75%	Nelson et al. 1993
18-62 × 8-7	17(+12pairs)	129	129	–	–	–	1146	Not estimated	Kubisiak et al. 2000
D4PC40 × D4PC13	15	154	63	–	–	41 (RFLPs) 5 isozyme	1115	Not estimated	Brown et al. 2001
<i>Pinus palustris</i>									
Tree 3-356	16(+6pairs)	133	133	–	–	–	1635	85%	Nelson et al. 1994
<i>P. elliotti</i> and <i>P. palustris</i> hybrids									
F1 of - 3-356 × H-28	♀: 18(+3 pairs)	122	122	–	–	–	1368	81%	Kubisiak et al. 1995
	♂: 13(+6 pairs)	91	91	–	–	–	953	62%	
BC1 of 488 × 18-27	♀: 17	133	133	–	–	–	1338	91%	Weng et al. 2002
	♂:19	83	83	–	–	–	995	81%	
<i>P. elliotti</i> and <i>P. caribea</i> var. <i>hondurensis</i> hybrids									
F1 of 2PEE1-102 × 1PCH1-63	♀: 24	125	117	8	–	–	1548	82%	Shepherd et al. 2003
	♂: 25	155	145	10	–	–	1823	88%	

The two-way pseudotestcross mapping strategy was used to construct genetic linkage maps of maritime pine using AFLP markers (Chagné et al. 2002; Ritter et al. 2002) as well as AFLP, SAMPL, SSR, and gene-based markers (N. de María and M.T. Cervera, unpublished results). Comparing the total map distance of genetic maps constructed based on haploid and diploid progeny from the same Maritime pine tree, a higher rate (28%) of recombination in the pollen parent was found (Plomion and O'Malley 1996). Such a significant difference between male and female recombination was also reported in other pine species (Moran et al. 1983; Groover et al. 1995; Sewell et al. 1999).

Loblolly Pine

Loblolly pine (*Pinus taeda* L.) has been used extensively for genetic mapping including the development of an early map based on 20 isozyme loci that included five linkage groups (LGs) (Conkle 1981). More recently, maps have been constructed in several pedigreed populations using several types of DNA-based markers. Devey et al. (1994) published the first map utilizing 90 RFLP and six isozyme loci. The map was based on a three-generation pedigree with 95 progeny and revealed 20 LGs. Genomic mapping was successfully used by Wilcox et al. (1996) to define a single gene locus for resistance to an isolate of the fungus (*Cronartium quercuum* f. sp. *fusiforme*) that causes fusiform rust disease. O'Malley et al. (1996) described a RAPD-based map of clone 7-56, a top *P. taeda* parent. This map had been constructed several years prior to publication. The first consensus map was produced by Sewell et al. (1999) combining data from two three-generation pedigrees, including the pedigree used by Devey. RFLP, RAPD, and isozyme markers were placed on the integrated or consensus map containing 357 loci and covering about 1,300 cM of genetic distance on 18 LGs. The first complete genome map was developed by Remington et al. (1999). They utilized haploid megagametophyte samples from an individual mother tree to develop a map based on 508 AFLP markers. This map revealed 12 LGs equaling the basic number of chromosomes and about 1,700 cM of genetic distance. Their analysis suggested that this distance saturated the genome, in slight contrast to an earlier estimate of 2,000 cM based on data from three species of pines (Echt and Nelson 1997). SSR markers developed in radiata pine (*P. radiata*) were used in a comparative mapping project between radiata and loblolly pine (Devey et al. 1999). Of the 20

SSR markers tested, only 9 were mapped in both species; however, these codominant markers along with several codominant RFLP markers were useful in defining homeologous LGs between the species. Temesgen et al. (2001) added 56 ESTP markers to the consensus map developed earlier by Sewell. The DGGE method proved quite useful for assaying ESTP markers and suggested a general method for placing genes on the maps since the markers were developed from expressed sequences. Additional ESTP markers were developed and used to identify anchored reference loci based on their sequence similarity between species and their nature to map to conserved locations in more than one species (Brown et al. 2001). Zhou et al. (2003) mapped 51 SSR markers, covering 795 cM on 15 LGs, in a three-generation pedigree with 118 progeny. The markers, developed from loblolly pine libraries of three types, were not found to be clustered within the genome, further highlighting the value of SSR markers in genome mapping.

Radiata Pine

Over the past 15 years, a range of DNA-marker-based linkage maps have been constructed for this species. Wilcox (1997) briefly reviewed mapping studies undertaken up till that date. Although results were summarized from seven studies involving construction of eight linkage maps, only one map had actually been published by that date, consisting of 208 (mostly) RFLP, SSR, and RAPD markers (Devey et al. 1996, 1999). This map consisted of 22 LGs and covered 1,382 cM. All of the other maps reviewed by Wilcox (1997) were constructed using RAPD markers, either using haploid megagametophytes, or diploid tissues using a pseudotestcross approach. The number of markers used in these studies ranged from 124 to 290, with only one of the maps having LGs equal to the haploid number of chromosomes and the remainder ranging between 14 and 22 LGs. Total map length estimates were undertaken in three studies ranging between 1,978 and 3,000 cM. Subsequent to these earlier studies a number of maps were published, some of which were included in Wilcox's 1997 review. Using 222 RAPD markers to genotype 93 megagametophytes, Emebiri et al. (1998) constructed a linkage map that covered 14 LGs and spanned a total distance of 1,665 cM. Kuang et al. (1999a) described a map constructed using megagametophytes of 198 S₁ seeds that had been genotyped with 168 RAPD and four microsatellite markers. The resulting map

consisted of 19 LGs, covering 1,116.7 cM, which was estimated to cover 56% of the genome. Because this was constructed using an S_1 family, elevated levels of segregation distortion were observed. Wilcox et al. (2001a) published framework maps of both parents of 93 full-sib progenies based on a total of 429 AFLP, RAPD, and SSR markers. These parent-specific maps were constructed using a pseudotestcross strategy and covered 1,414 and 1,144 cM in 20 and 21 LGs, respectively. These maps have subsequently been added to, using over 300 SSR and EST markers, and have been reduced to 12 LGs (Wilcox et al. 2004). Devvey et al. (1999) published a comparative map of radiata and loblolly pine based on RFLP, SSR, and RAPD loci and showed that the highly syntenic nature of *Pinus* applies to these two economically important species. Overall estimates of map length appear to be similar to that of loblolly pine (Wilcox et al. 2001a).

Scots Pine

Early mapping work in *P. sylvestris* has been based on isozyme loci (Rudin and Ekberg 1978; Szmidi and Muona 1989). These maps contained no more than 20 loci. The number of RFLP markers developed for Scots pine had been very low (Karhu et al. 1996). Thus, the next mapping efforts were based on RAPD makers segregating in haploid megagametophytes. Yazdani et al. (1995) mapped 261 markers in 14 LGs in a tree that is part of the breeding program. Hurme et al. (2000) made a low-coverage RAPD map for an F_1 tree that was a result of north $\times$ south cross, such that alleles for important quantitative traits were assumed to segregate in the same cross. The map with the best genome coverage, so far, was constructed by Yin et al. (2003). The AFLP mapping in a full-sib family resulted in two maps, for each of the breeding program parents. The map lengths for the two parents, based on about 200 framework markers, were about 1,645 and 1,681 cM for the male and female trees, respectively, with very high estimated genome coverage. Most recently, Komulainen et al. (2003) mapped about 60 gene based markers in the F_1 progeny of a north $\times$ south cross. This map also contained markers that had been previously developed for *P. pinaster* (Plomion et al. 1999), and others that had been used for *P. taeda*. Most importantly, the homologous markers allowed defining the correspondence between 12 LGs in the two species. More markers were later added to this map (Pyhäjärvi et al. unpublished).

Longleaf Pine, Slash Pine, Caribbean Pine, and their Hybrids

Longleaf pine (*P. palustris* Mill.), slash pine (*P. elliottii* Engelm. var. *elliottii*), and Caribbean pine (*P. caribaea* Morelet.) are hard pines of subsection *Australes* found along the coastal plains of the southeastern United States, eastern Central America, and the Caribbean islands. Although loblolly pine (*Pinus taeda* L.) has been planted on millions of acres that were once typically occupied by these species, in many situations these pines are proving to be the preferred timber species due to their adaptation to the coastal-plain soils and the associated natural disturbances such as frequent fires and hurricanes (Wahlenberg 1946; Shoulders 1984). A number of genetic maps, consisting primarily of RAPD markers, have been constructed for slash pine, longleaf pine, and their hybrids (Nelson et al. 1993, 1994; Kubisiak et al. 1995), with the main goal being to use these marker maps as a tool for dissecting the inheritance of specific traits of interest and for use in marker-assisted-selection (MAS) strategies within tree-improvement programs (Kubisiak et al. 1997, 2000; Weng et al. 2002). Some markers significantly linked to traits of interest have been converted to more easily scorable markers, such as sequence characterized amplified region (SCAR), to aid selection efforts (Weng et al. 1998). Brown et al. (2001) assembled a genetic linkage map for slash pine using a variety of markers [RAPDs, expressed sequence tag polymorphisms (ESTPs), restriction fragment length polymorphisms (RFLPs), and isozymes]. An additional genetic map for slash pine and one for Caribbean pine, using amplified fragment length polymorphisms (AFLPs) and microsatellite or SSR markers, were created using an F_1 hybrid population (Shepherd et al. 2003). The number of markers mapped and the genetic distances covered by some of the published maps for these species and their hybrids are summarized in Table 3. Unlike loblolly pine, significantly less effort has been focused on comparative mapping across these species. However, studies have shown synteny across slash and longleaf pines using RAPDs (Kubisiak et al. 1995, 1996), slash and loblolly pines using ESTPs as anchored reference loci (Brown et al. 2001), and slash and Caribbean pine using AFLPs (Shepherd et al. 2003). In addition, SSR markers look promising for further comparative analyses across these species (Shepherd et al. 2002a; C.D. Nelson and C.S. Echt, pers. comm.).

Other Pines

Genetic linkage maps have also been constructed in other pine species including *P. brutia* (Kaya and Neale 1995), *P. contorta* (Li and Yeh 2001), *P. edulis* (Travis et al. 1998), *P. massoniana* (Yin et al. 1997), *P. strobus* (Echt and Nelson 1997), *P. thunbergii* (Kondo et al. 2000; Hayashi et al. 2001). These were based mainly on RAPDs and AFLPs.

2.2.4

Genetic vs. Physical Size and Practical Implication

These mapping studies have led to the conclusion that the total genetic distance of the pine genome is around 2,000 cM (Gerber and Rodolphe 1994), i.e., about 167 cM per chromosome. Given a physical size of 25 pg/C, one unit of genetic distance (1 cM) would therefore correspond to 13×10^6 nucleotides (13 Mb), while it represents 0.23 Mb in the model plant species *Arabidopsis*! Such high genetic/physical size ratio obviously hampers the characterization of QTLs by fine-mapping and positional cloning approaches. Hence, as will be discussed in Sects. 2.3 and 2.4, the only way for understanding the molecular basis underlying quantitative trait variation is the candidate-gene approach in which genes are identified a priori as likely candidates for the trait of interest and their polymorphisms tested against quantitative trait variation. Interestingly, despite the 56-fold difference between *Pinus* and *Arabidopsis* chromosomes, the number of crossings over per chromosome was found to be highly conserved between both genera: 2 to 4 chiasmata per bivalent (1 chiasma = 50 cM; Ott 1991). A comparison of genome lengths among evolutionary divergent pines found *P. pinaster*, *P. palustris*, and *P. strobus* to have essentially identical rates of recombination (Echt and Nelson 1997). Thus, genetic mapping studies carried out in pines have clearly demonstrated that the mechanism of crossing over is conserved on a chromosomal basis, and independent of physical map size and the fraction of coding DNA.

2.2.5

Comparative Mapping: Toward the Construction of a Unified Pine Genetic Map

All pine species have the same number of chromosomes (i.e., $n = 12$) as well as a similar genome size. Moreover, they are all diploid, which suggests

that their genome might be relatively well conserved among species. Approaches for evaluating genome similarity have used cytogenetics and linkage map comparison. Although cytogenetics can provide a direct idea about the conservation between different genomes (Hizume et al. 2002), most of the interspecies comparisons have been carried out using genetic maps. The same tools have also been developed for other applications such as gene mapping and QTL detection. Comparative genome mapping aims to measure the conservation of gene content (synteny) and order (colinearity) among chromosomes and uses orthologous loci as anchor points between maps. Comparative genome mapping has been successfully used in grasses to explain the genome evolution of cereals (Moore et al. 1995), for choosing a model species for whole-genome sequencing (rice, Ware et al. 2002; http://www.cns.fr/externe/English/Projets/Projet_CC/organisme_CC.html), and for transferring genetic information between related species, such as the position of candidate genes (Schmidt 2002). The lack of genome sequence for a Pinaceae species has made comparative mapping even more important as the primary tool for integrating genetic information across species.

To define the syntenic relationships among phylogenetically related pine species, orthologous markers (i.e., homologous DNA sequences whose divergence follows a speciation event and whose sequence and genome location is conserved between different species) are used. A first example was provided by Devey et al. (1999), who aligned the genetic maps of *P. taeda* and *P. radiata* using RFLPs and SSRs. This first effort was further consolidated in the frame of the Conifer Comparative Genomics Project (<http://dendrome.ucdavis.edu/ccgp>). Low-copy cDNA PCR-based markers were developed in loblolly pine (Harry et al. 1998; Brown et al. 2001; Temesgen et al. 2001; Krutovsky et al. 2004). These markers showed a relatively good PCR cross-amplification rate between pine species because they target conserved coding regions, showing a relatively high polymorphism rate and a low number of paralogous amplification when PCR primers were chosen carefully. They were used to study the synteny between species belonging to the Family Pinaceae, which included pines, along with other important conifers such as spruces and firs. These markers made it possible to assign LG homologies for 10 out of 12 chromosomes between *P. elliotii* and *P. taeda* (Brown et al. 2001; <http://dendrome.ucdavis.edu/>

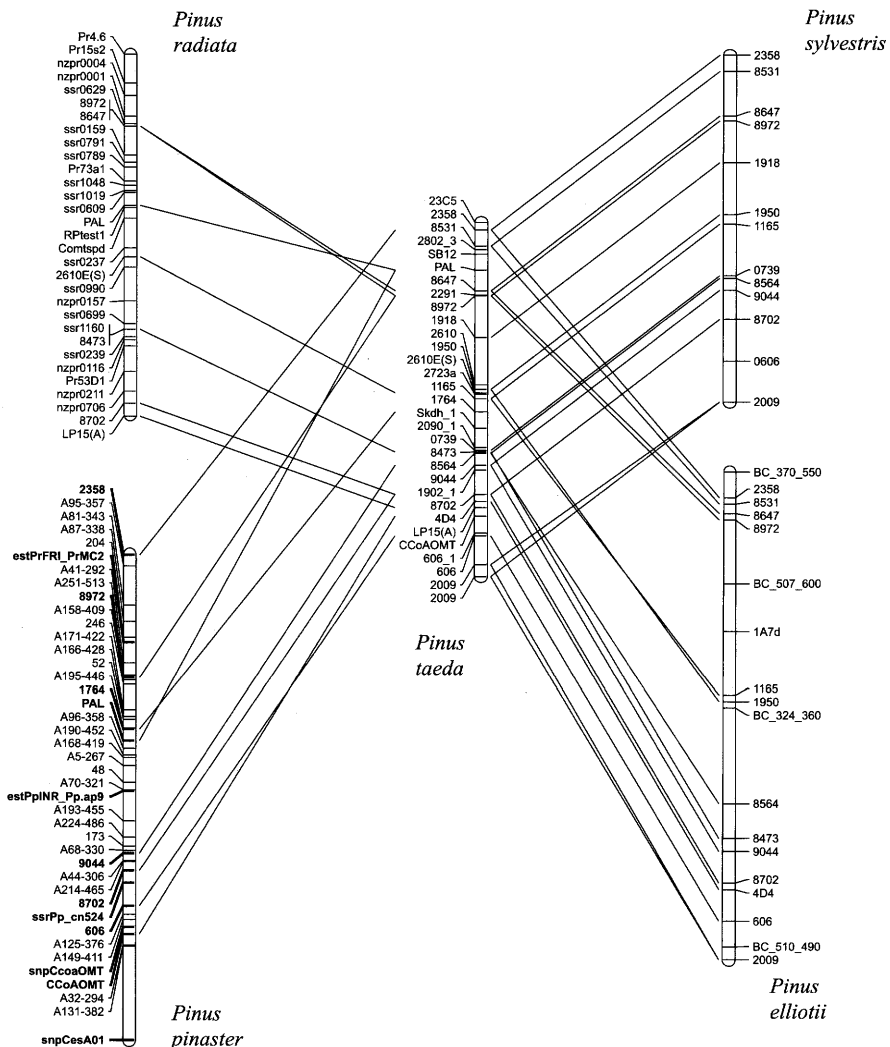
ccgp/compmaps.html), 10 out of 12 between *P. pinaster* and *P. taeda* (Chagné et al. 2003), and 9 out of 12 between *P. sylvestris* and *P. taeda* (Komulainen et al. 2003). About 30 or 40 ESTP markers were proved to be useful in demonstrating large areas of synteny between each species pair. While this comparison was only of low density, these pioneering studies suggested that pine genomes did not show any apparent chromosomal rearrangement. They also provided an indication that gene content and gene order is conserved, as is illustrated for LG 6 in Fig. 2. Current efforts are being expended to add more markers common to *P. radiata* and *P. taeda* (P. Wilcox, personal communication). From an application point of view, these comparisons provide a set of markers that can be used for constructing framework genetic maps

of pine species for which maps have not yet been developed.

As more conifer ESTs become available in public databases (329,531 in *Pinus teada*, 132,531 in *Picea glauca*, 27,283 in *Pinus pinaster*, 28,170 in *Picea engelmannii* × *Picea sitchensis*, 80,789 in *Picea sitchensis*, 6,808 in *Pseudotsuga menziesii*, 7,639 in *Cryptomeria japonica*: EMBL 19 March 2006), a computational approach could be used for *in silico* development of putative orthologous EST-based markers (Fulton et al. 2002), as was recently illustrated between loblolly pine and Douglas fir by Krutovsky et al. (2004). Such resources should help to define the precise syntenic relationship across conifers and establish a framework for comparative genomics in Pinaceae.

The alignment of genetic maps of *P. pinaster* and *P. taeda* made it possible to discover putative con-

Fig. 2. Synteny in the *Pinus* genus (linkage group 6): alignment of genetic maps of five pine species



served QTLs between the two species (Chagné et al. 2003). Those are QTLs for wood density and wood chemical composition traits located on LGs 3 and 8, respectively. The same observation has been made in *P. radiata* (Telfer et al. 2006). Preliminary studies indicate wood-density QTLs are colocating more frequently than would be expected by chance between these two species (Telfer et al. 2006), and analyses are being extended to other wood property traits. Moreover, candidate genes coding for functions that are linked to wood formation have been mapped in the same regions, which indicates that they may be involved in the molecular control of those traits. These first examples of the application of comparative genome mapping in pines show that comparative genome mapping can be used to verify QTLs across species and that the same genes may be involved in the genetic control of the same traits.

2.3

Genetic Architecture of Complex Traits

Pine tree improvement is hampered by different inherent characteristics: (1) the time needed to reach sexual maturity, (2) the time lag required to evaluate field performance (e.g., growth; Kremer 1992), and (3) in some cases, the cost of phenotyping (e.g., wood-quality-related traits). This makes breeding of these species a slow process compared to that of most common crop plants. In addition, selection of these traits remains imprecise because environmental effects are rather high for most traits of interest. Heritabilities for height, diameter, volume, branching traits, and bole taper, i.e., straightness, are in the range of 0.1 to 0.3, and only slightly higher (0.3 to 0.6) for wood and end-use properties (reviewed by Cornelius 1994). In this context, any tool directed toward selection processes that would improve the evaluation of genetic value and also reduce the generation time would be of considerable value.

Most traits important to forestry, such as biomass production, wood quality, and biotic and abiotic stress resistance are complex quantitative traits. In the theory of quantitative genetics it is assumed that the heredity of a quantitative trait can be ascribed to the additive effects of a large number of genes with small and similar actions, modulated by environment. This assumption has been questioned since the early ex-

periments of Sax (1923) and Thoday (1961), and it is now well known that a small number of segregating loci are involved in the genetic control of quantitative trait variation. These genes act together to provide a quantitative difference and are referred to as quantitative trait loci or QTLs (Geldermann 1975). The basic theory of using genetic markers to detect QTLs was introduced by Sax (1923). Initially, the application of this theory was limited by the lack of available segregating markers; however, rapid advances in DNA marker-based technologies since the 1980s have now made it possible to genotype hundreds of genetic markers to construct dense genetic linkage maps (Sect. 2.2.3) and further to carry out a comprehensive search of QTLs along the genome. Sewell and Neale (2000) and Guevara et al. (2005b) recently reviewed the science of QTL mapping in pine trees.

In the first part of this section, devoted to the genetic dissection of agronomically important traits, we will present an update of the studies that were carried out in pines. The specificity of pines regarding the different types of populations and strategies used to detect QTLs will be presented. Then, the main results of these studies will be discussed. In the second part, a perspective on the identification of diagnostic markers for pine-tree breeding will be discussed.

2.3.1

Strategy and Methods Used for QTL Detection in Single Family Pedigrees

Pines are characterized by late maturity (longevity), an outbred mating system, and a high genetic variability (Hamrick and Godt 1990). Their outbred mating system and high genetic load have hampered the development of inbred lines, the material of choice from a QTL mapping perspective. In this context, specific populations and statistical methods were developed specially for forest trees, and pines in particular. Until recently, most of the QTL mapping efforts were focused on single family pedigrees. However, given the high rate of polymorphism encountered in pines and the relative lack of stability of QTLs in different genetic backgrounds, methods aimed at validating markers linked to the traits of interest in more complex pedigrees or even in unrelated genotypes are emerging. In this section, we will review the type of population, statistical methods, and main results that have been obtained using simple mapping pedigrees. The limitations of this approach will be underlined, and the use

of complex pedigree designs or unrelated populations to unravel the genetic architecture of complex traits will be presented. Finally, the relevance of integrative approaches combining classical QTL studies and transcriptome and proteome analysis, together with studies of molecular evolution, will be highlighted.

Single Family Designs

Two-Generation Full-Sib Design The advantage of naturally high levels of heterozygosity in outbred forest trees can be utilized in a two-generation population structure, where it may be expected that trees chosen as potential parents will likely be heterozygous for some number of QTLs, which will then segregate in the F_1 . Typically, “plus trees” are identified and are used as parents of a QTL mapping population. Various studies have taken advantage of this two-generation design to analyze each parent under a pseudotestcross model (Kumar et al. 2000; Lerceteau et al. 2000; Ball 2001; Shepherd et al. 2002b; Weng et al. 2002; Markussen et al. 2003; Yazdani et al. 2003). This model is well suited for dominant markers. However, the main limitation is that the phenotypic effects inherited from each parent are analyzed individually, even though the genetic contribution of each parent simultaneously contributes to the phenotypic variation in the progeny population. Consequently, the genetic information in the four progeny classes of an outbred pedigree is collapsed into only two genotypic classes, thereby reducing the robustness of the analysis. Of course, if codominant markers are used, a consensus map can be built precisely to detect allele effects from both progenitors simultaneously.

Three-Generation Full-Sib Design In the three-generation outbred population structure, two crosses are made among four unrelated grandparents, where each mating pair is selected among individuals displaying divergent phenotypic values for the trait (e.g., Groover et al. 1994). From each grandparental mating, a single phenotypically intermediate individual is chosen as a parent. Presumably, these intermediate parents are heterozygous for both marker and QTL alleles and are potentially heterozygous for different allelic pairs that display a divergent phenotypic effect. This three-generation full-sib structure is typically designed for QTL analysis under an outbred model and has been used extensively (Table 4). Alternatively, Plomion et al. (1996a, b) utilized the selfing ability of *P. pinaster* to design experiments that fit an

inbred F_2 model. Although RAPD markers were used, genetic information from progeny and corresponding megagametophytes were utilized to overcome the limitations associated with dominant markers using an F_2 model. It was also proposed to use trans-dominant-linked markers to overcome the problem of dominant markers on the sporophytic phase of F_2 trees (Plomion et al. 1996c).

Two-Generation Half-Sib Design With this population structure, the effects of two maternal QTL alleles are averaged over a large pollen pool (Hurme et al. 2000). This type of structure allows one to test the stability of the effect of the maternal alleles in different genetic backgrounds. In addition to these classical mapping designs, particular populations (F_1S) were especially developed to analyze inbreeding depression in *P. taeda* and *P. radiata* (Kuang et al. 1999a,b; Remington and O'Malley 2000a,b; Williams et al. 2001).

Two-Generation Full-Sib and Half-Sib Design An extension of the pseudotestcross QTL mapping strategy, in which QTLs are defined in a narrow genetic background, Plomion and Durel (1996) show that a “general” value of a “specific” QTL detected in a full-sib family could be easily evaluated, provided that both parents of the full-sib were involved in maternal half-sib (open-pollinated or polycross) families. Such two-generation pedigrees are widely available in most pine breeding programs that involve the simultaneous estimation of specific and general combining abilities of selected trees. However, this strategy has never been tested experimentally.

Methods Used for QTL Mapping in Single Family Designs

Regardless of the population structure and size (Beavis 1994), several factors must be considered for successful QTL detection. Statistical methodology significantly influences the accuracy of QTL position and effect estimation. Simple statistical methods such as analysis of variance (ANOVA) have opened the way to the development of more powerful QTL detection methods, integrating information available at multiple markers: interval mapping (IM), composite interval mapping (CIM), and multiple interval mapping (MIM).

The first method, called single marker analysis, proposed by Edwards et al. (1987), is the simplest one. ANOVA is performed with one marker at a time, on the genotypic classes defined by a single marker. This

Table 4. QTL and marker-trait association detected in pines: pedigree structure and methods

Species	Objective	Population (size) ^a	Replications ^b	Clonal replicates	Genetic markers	QTL analysis (analytical model) ^c	Software	Reference
<i>Pinus elliottii</i>	Detection	F1OB (186)	1	No	RAPD	SG (54 : 27 genotypes from each tail), SM, MM	SAS	Kubisiak et al. 2000
<i>Pinus elliottii</i> <i>var elliottii</i> × <i>Pinus caribaea</i> <i>var hondurensis</i>	Detection	F1OB (89)	2 sites (60,29)	No	AFLP, SSR	SM, CIM	QTLcartographer	Shepherd et al. 2005
(<i>Pinus palustris</i> Mill × <i>Pinus elliottii</i> Engl) × <i>Pinus elliottii</i> Engl	Detection, stability (age)	F1OB (258)	3 sites (82,83,93)	No	–	SM, IM	SAS, MAPMAKER/QTL	Weng et al. 2002
<i>Pinus sylvestris</i>	Detection	F1OB	1	No	AFLP	IM	Qgene	Lerceteau et al. 2000
	Detection, Stability (time replicate)	F2HS	2	No	RAPD	SG (1994 : 48 vs. 48, 1996 48 vs. 48), SM, Bayesian QTL analysis	SAS, Multimapper/ OUTBRED	Hurme et al. 2000
	Detection	F1OB	1	No	RAPD	SM	SAS	Yazdani et al. 2003
<i>Pinus pinaster</i>	Detection, Stability (Age)	F2S (120)	3 (age)	No	RAPD	IM	MAPMAKER/QTL	Plomion et al. 1996a
	Detection	F2S	1	No	RAPD, AFLP, proteins	CIM	QTLcartographer	Costa 1999
	Detection	F2OB (186)	1	No	AFLP	CIM	MULTIQTL	Brendel et al. 2002
	Detection	F2OB (186)	1	No	AFLP, EST	CIM	MULTIQTL	Pot et al. 2005
	Detection	F1OB (80)	1	No	AFLP, SSR, EST	IM	SAS	Markussen et al. 2002
	Detection, stability (age, season)	F2OB (186)	1	No	AFLP, EST	CIM	MULTIQTL	Pot 2004
	Detection	F2S (120)	1	No	RAPD	IM	MAPMAKER/QTL	Plomion et al. 1996b

^a F1OB: two-generation outbred design, F2OB: three-generation outbred design, F2S: three-generation inbred design, F2HS: Two-generation half-sib design, CD: complex design (in this case more information is provided)

^b When the traits were measured on one site and at one given time, 1 is indicated, otherwise more details are provided

^c SM: single-marker analysis, MM: multiple-marker analysis (without map information), IM: interval mapping, CIM: composite interval mapping, SG: selective genotyping, BSA: bulk segregant analysis

Table 4. (continued)

Species	Objective	Population (size) ^a	Replications ^b	Clonal replicates	Genetic markers	QTL analysis (analytical model) ^c	Software	Reference
<i>Pinus radiata</i>	Detection, stability (age)	F1OB (174)	1	No	RAPD	SM, BSA (9 vs 9)	-	Emebiri et al. 1997
	Detection, stability (age)	F1OB (80-93)	3 times (age)	No	RAPD, AFLP, SSR	IM, MTM	-	Kumar et al. 2000
	Detection, stability (age)	F1OB (93)	2 times (age)	No	RAPD, AFLP, SSR	Bayesian approach	Splus	Ball 2001
	Detection	CD	1	No	SSR	-	-	Kumar et al. 2004
	Detection, validation, stability (genetic background)	3 unrelated F1OB (400 each)	2 sites	No	RFLP and SSR	SG (Juvenile wood density : 50 vs 50; Diameter at breast height : 100 vs 100), SM	GENSTAT	Devey et al. 2004a
	Detection, validation	Detection: 6 related F2OB (202) Verification: 1 F2OB (400)	1 site for the detection population 2 sites for the verification population	No Yes	RFLP + SSR	SM within family + SM among families	GENSTAT	Devey et al. 2004b
<i>Pinus taeda</i>	Detection	F2OB (Detection population : 172)	6 sites (19 to 35 trees per sites)	No	RFLP	SG, SM	Home made	Groover et al. 1994
	Detection	F2OB (Detection population : 172)	6 sites (19 to 35 trees per sites)	No	RFLP	IM	Home made	Knott et al. 1997
	Detection, stability (age, genetic background)	2 F2OB (Base : 95, Detection : 172)	2 sites for BASE (48+47), 6 sites for Detection (19 to 35 trees per sites)	No	RFLP, RAPD, Isozymes	IM	Home made	Kaya et al. 1999
	Detection, stability (age), development of statistical methods	F2OB (91)	5 times (age)	No	SSR	IM	Home made	Gwaze et al. 2003
	Detection, stability (age, season)	F2OB (Detection population : 172)	6 sites (19 to 35 trees per sites)	No	RFLP	IM	Home made	Sewell et al. 2000
	Detection, stability (age, season)	F2OB (Detection population : 172)	6 sites (19 to 35 trees per sites)	No	RFLP	IM	Home made	Sewell et al. 2002
	Detection, validation, stability (genetic background)	1 F2OB detection (D) : 172, validation (V) : 457	D : 6 sites (19 to 35 trees per sites)	No	RFLP + EST	IM	QTL express	Brown et al. 2003

method suffers from several limitations. First, it does not provide any information regarding the location of the QTLs in the genome, and furthermore the definition of the QTL effect is largely inaccurate given the inability to separate small-effect QTLs with tight linkage from QTLs of large effect but more distant linkage.

In order to improve the efficiency of QTL mapping through the use of genetic map information, Lander and Botstein (1989) developed the IM method. This approach allows QTLs to be detected in the intervals defined by the markers. The IM has been widely used in pines (Groover 1994; Plomion et al. 1996a, b; Knott et al. 1997; Kaya et al. 1999; Costa and Plomion 1999; Costa et al. 2000; Sewell et al. 2000, 2002; Brown et al. 2003). This method is, however, rather limited as QTL detection is made in a linear way, i.e., the same test is applied at each point of the interval without taking into account the results of successive tests.

More recently, statistical approaches have been developed to increase the statistical power of QTL detection and have received increasing attention in pines (Costa 1999; Brendel et al. 2002; Pot et al. 2005b). One of these approaches is CIM, developed by Zeng (1993a, b) and Jansen (1993), which combines IM with multiple regression. Like IM, this method evaluates the presence of a QTL at multiple analysis points across each interlocus interval. However, at each point it also includes in the analysis the effect of one or more markers elsewhere in the genome. However, although IM and CIM brought significant improvements, both methods lack dimensionality: i.e., only a single QTL is being searched at a time.

The method proposed by Kao et al. (1999) and Zeng et al. (2000) called MIM differed from the previous methods through the implementation of multi-QTL models. The selection of the model (i.e., QTL number, position, effect, and interactions) that best fits the data follows an iterative process. After identification of QTL number and position by CIM, the MIM strategy consists of looking for additional QTLs through forward-backward selection cycles while integrating interaction information between the different QTLs. Within each iterative cycle, QTL position and effect are reevaluated. MIM also allows the evaluation of QTL epistasis. However, it is important to note that, although this strategy provides significant improvements over CIM, it does not allow the detection of nonsignificant QTLs at the individual level. Therefore, even if the concept aimed at detecting multiple QTL effects is simple, its implementation is relatively complex given the number of potential QTLs and the

resulting number of possible interactions. Thus, the problem is no longer genetic but resides in the ability to test all the possible genetic models and select the one that best fits the observed data (*model selection*). Until now, the MIM algorithm implemented in QTL Cartographer developed by Kao et al. (1999) and Zeng et al. (2000) has not been directly used in pines, but Bayesian approaches, also based on model selection, have been applied in these species (Hurme et al. 2000; Ball 2001).

The simultaneous analysis of multiple correlated traits has also been incorporated into algorithms in order to improve QTL detection efficiency (Jiang and Zeng 1995; Korol et al. 1995, 1998). Although these algorithms have not been specifically used up till now, there is much interest in such algorithms, especially for such traits as annual growth or wood density. Additionally, application of specific genotyping strategies has further maximized the efficiency of QTL mapping. One such strategy, called selective genotyping, has been extensively used in pines (Groover et al. 1994; Hurme et al. 2000; Kubisiak et al. 2000; Devey et al. 2004a). Another strategy, termed bulked segregant analysis (BSA), commonly used to study qualitative traits, has also been applied to analyze quantitative traits (Emebiri et al. 1997).

Besides statistical procedures, it has been clearly shown that the power of QTL detection largely depends upon the quality of the phenotypic assessment. Poor phenotypic assessments, i.e., imprecise measurements, generally result in QTLs with true effects being left undetected, but in some circumstances (but to a lesser extent) might even lead to the detection of false QTLs. Without the possibility of developing F_3 populations or recombinant inbred lines to precisely estimate the value of the traits, clonally propagated material has become the material of choice for forest tree geneticists (e.g., Scotti-Saintagne et al. 2004). However, this type of material has only been rarely used in pines (Devey et al. 2004b). This is most likely due to the large capital investments and technical expertise needed to clonally propagate pines and the wide variability noted among specific genotypes in their ability to produce rooted cuttings.

2.3.2

QTL Discovery in Single Family Pedigree Designs

Twenty-six QTL studies aimed at detecting associations between molecular markers and trait variation have been performed in seven pine species. However,

most of the efforts have been concentrated in three species, *P. pinaster*, *P. taeda*, and *P. radiata* (Tables 4 and 5). In addition, studies on inbreeding depression have been carried out in *P. radiata* and *P. taeda* (Kuang et al. 1999a, b; Remington and O'Malley 2000a, b; Williams et al. 2001).

Qualitative Traits

Associations between qualitative traits and molecular markers have led to the identification of markers linked to resistance to different rust diseases: pine needle gall midge in *P. thunbergii* (Hayashi et al. 2004), white pine blister rust in *P. lambertiana* Dougl (Devey et al. 1995; Harkins et al. 1998), and fusiform rust disease in *P. taeda* (Wilcox et al. 1996). A major gene controlling the biosynthesis of δ -3 carene has also been mapped in *P. pinaster* (Plomion et al. 1996b). Most of these studies have been based on intraspecific mapping populations. Additionally, two interspecific crosses were used to increase the level of polymorphism and the range of variation for branch architecture in the mapping population (Shepherd et al. 2002b).

Quantitative Traits

Growth has been the most studied trait in QTL mapping studies in pines. To date, a comparable number of studies have been achieved for wood and end-use properties. These two classes of traits have been studied either globally (e.g., height growth, specific gravity) or after decomposition into simpler components (e.g., growth unit, ring density). Traits involving adaptation to the environment have also been studied, including tree response to heavy metal, drought, and cold stress, as well as bud phenology (Costa 1999; Hurme et al. 2000; Kubisiak et al. 2000; Lerceteau et al. 2000; Brendel et al. 2002; Yazdani et al. 2003).

QTL Results

The diversity of population types, population sizes, marker types, QTL detection methods, detection thresholds, and variation of phenotypic trait measurements have made the comparison between QTL experiments a difficult task. However, general observations can be made regarding QTL number, position, phenotypic effect, and stability.

QTL Number and Effect Most QTLs have been detected at a single maturation stage and in a single environment. Therefore, although all the analyzed traits

show continuous variation, suggesting polygenic control of the traits, only a limited number of QTLs per trait (Table 4: between 0 and 8) have been detected. This number is in general smaller than in annual crops and has led to a smaller proportion of the phenotypic variation being explained (Table 5). As stated in the chapter on Eucalypts, "the limited power to detect QTL in forest trees compared to crop species may be due to the high environmental and developmental variation in tree plantations, as well as to the small size of the analyzed populations" (mainly between 91 and 200 genotypes, Table 4). However, QTL analyses carried out by Brown et al. (2003) and Devey et al. (2004a) using large mapping populations (>400 individuals) do not support the latest hypothesis. Brown et al. (2003) reported QTL effects that were two- to threefold smaller than those reported by Sewell et al. (2000, 2002) for the same traits. Such divergence likely represents more accurate estimates of the QTL effects owing to the larger segregating population analyzed. These results suggest that most QTL studies performed in pines, with the exception of the analyses carried out by Devey et al. (2004a, b) and Brown et al. (2003), have yielded an overestimation of the QTL effects, which was also suggested by Beavis et al. (1994). Consequently, as underlined by Wilcox et al. (1997), the genetic determinism of most target traits for pine breeding is likely to be explained by small-effect genes, rather than any moderate- to large-effect genes. However, a larger number of QTL experiments with larger pedigree sizes will be required to validate this hypothesis.

Compared to the small number of QTLs detected at any one maturation stage, the analysis of QTL stability along a cambial age or seasonal gradient (Table 4) revealed a significant increase in the number of QTLs detected. For instance, the simultaneous analysis of different maturation stages combined with seasonal variation allowed for the detection of 7 to 23 QTLs for wood-specific gravity, the percentage of latewood, and microfibrillar angle (Sewell et al. 2002). Similar results were reported by Pot (2004), who observed a total of 30 QTLs for wood density, 42 for wood heterogeneity, and 33 for radial growth when measured over several cambial ages.

QTL Stability In their review, Sewell and Neale (2000) pointed out that "before a commitment to marker-aided selection or breeding (MAS/MAB) can be made in tree breeding, QTL that have been detected must be verified in different experiments, as well as in differ-

Table 5. QTLs and marker-trait associations detected in pines: number of QTLs and phenotypic variance explained

Species	Type of trait	Trait (abbreviation used in the article)	#QTLs ^a	%Variance explained by each QTL ^b	Note	Reference
<i>Pinus elliottii</i>	Adaptative traits	Aluminium tolerance	3*	15.61 ^{all}		Kubisiak et al. 2000
<i>Pinus elliottii</i>	Wood-quality traits	Average branch angle (AVBRA)	0	–		Sheperd et al. 2002b
<i>var elliottii</i> × <i>Pinus caribaea</i>	and growth traits	Average number of branch per whorl (AVBRN)	3***	12–18		
<i>var hondurensis</i>		Average branch diameter (AVBRD)	2***	16–17		
		Average whorl spacing (AVWS)	2***	15–19		
		Regularity of whorl spacing (CVWS)	1***	17		
		Stem class (SC)	0	–		
		Number of distinctively large, steep angled branches observed per tree (RAM)	0	–		
		Number of "leaders" (DL)	0	–		
		Bark thickness (AVBT)	2***	11–12		
		Relative bark thickness (RBT)	0	–		
		Trunk height (HT)	3***	13–21		
		Overbark diameter at breast height (OBDBH)	1***	16		
		Underbark diameter at breast height (UBDBH)	1***	17		
		Basic density (BD)	1***	14		
<i>(Pinus palustris</i> Mill × <i>Pinus elliottii</i> Engl) × <i>Pinus elliottii</i> Engl	Growth traits	Total height (month 7,16,29,41)	5 ^{lod2}	3.6–11		Weng et al. 2002
		Stem diameter	6 ^{lod2}	4–10		
		Height increments	5 ^{lod2}	4.3–8.5		
		Diameter increments	5 ^{lod2}	4.2–11		
<i>Pinus sylvestris</i>	Growth, wood-quality, and adaptive traits	Tree height (TH)	3 ^{lod2}	11.5–12.2		Lerceteau et al. 2000
		Trunk diameter at breast height (DBH)	2 ^{lod2}	9.3–15		
		Trunk diameter 0.5m from ground (D0.5)	2 ^{lod2}	12.6–13.3		

^a * significant at chromosomeswide level ($p < 0.05$),

*** significant at genomewide level ($p < 0.05$),

^{lodx} lod threshold value provided by authors (IM),

^{pV=x} significance level used for single-marker analysis,

⚙: unique QTLs, defined as subset of QTLs (suggestive or significant) that map within ca. 15 cM of one to another and have the same general profile for their parental and interaction effects (magnitude and direction of effects)

^b when the % of phenotypic variance explained by each QTL was not provided, the phenotypic variance explained by all QTLs is indicated ^{all}

Table 5. (continued)

Species	Type of trait	Trait (abbreviation used in the article)	#QTLs ^a	%Variance explained by each QTL ^b	Note	Reference
<i>Pinus pinaster</i>		Branch diameter of average branch at fourth branch level from terminal bud (BDA)	0	–		
		Branch angle (BA)	1 ^{lod2}	15.9		
		Basic wood density measured with Pilodyn (PIL)	0	–		
		Frost hardness, critical temperature giving mean injury (CT _m)	2 ^{lod2}	12.1–21.1		
		Frost hardness, estimated temperature causing slight injury to 50% of needles (CT ₅₀)	2 ^{lod2}	11.3–22.7		
		Tree volume calculated with DBH (VOL)	2 ^{lod2}	10.7–16.7		
		Tree volume calculated with D0.5 (VOL0.5)	2 ^{lod2}	11–14.6		
	Adaptive traits	Bud set	3	2.1–12.7		Hurme et al. 2000
		Frost hardness	7	3–11.1		
	Adaptive traits	Cold acclimation	16 ^{p=0.01}	9–19.5 ^{all}		Yazdani et al. 2003
	Growth traits	Height growth	11 ^{p=0.01}	10.8–30.8 ^{all}		
	Growth traits	Total height week 15	2 ^{**}	7–12		Plomion et al. 1996a
		Total height week 38	3 ^{**}	6.2–11.5		
		Total height week 92	1 ^{**}	10		
		Fertile zone length (LF)_ cycle 5	1 ^{**}	19.6		
		Fertile zone length (LF) adjusted to NSU_ cycle 5	1 ^{**}	11.8		
		Number of stem units in the fertile zone (NSU)_ cycle 5	1 ^{**}	20.4		
		Mean stem unit length (MSUL)_ cycle 5	1 ^{**}	6		
		Germination date (GERMD)	2 ^{**}	15–17		
		Hypocotyl length (Lhypo)	2 ^{**}	9.5–12.7		
		Megagametophyte weight (MW)	2 ^{**}	6–8.6		
	Adaptive and growth traits	Delta 13C	3 ^{***} + 5 ^{**}	4.7–12.4	pop : id to	Brendel et al. 2002
		Ring width	1 ^{***} + 5 ^{**}	5.9–18.1	Pot et al 2006 and Pot 2004	
	Wood-quality and growth traits	Total height	0	–	pop : id to	Pot et al. 2006
		Mean density of all the rings	2 ^{**}	4.6–4.8	Brendel et al 2002	
		Density estimated by Pilodyn penetration	2 ^{**} + 1 ^{***}	3.7–8.8	and Pot 2004	
		Density heterogeneity	3 ^{**}	5.1–8		
		Lignin content	7 ^{**}	5–9		

Table 5. (continued)

Species	Type of trait	Trait (abbreviation used in the article)	#QTLs ^a	%Variance explained by each QTL ^b	Note	Reference
<i>Pinus radiata</i>	Wood-quality and growth traits	Water extractives content	2** + 1***	5.8–8.8		
		Acetone extractive content	0	–		
		Alpha cellulose content	4**	3.7–7.8		
		Hemicellulose content	3** + 1***	4.5–8.1		
		Lignin composition	3**	5.1–9.4		
		Kraft pulping yield adjusted to Kappa number	1***	8.8		
		Kappa index	3**	4.7–6.4		
		Arithmetic fiber length	2** + 1****	4.9–12.3		
		Weighted fiber length	2** + 1***	6.1–11.9		
		Fibre width	2** + 1***	5.4–8.2		
		Coarseness	2** + 1***	4.4–11.9		
		Curl	1** + 1***	6–10.2		
		Zero span tensile value	5** + 2****	4.9–7.3		
		Alpha cellulose content	5**	5.98–12.24		Markussen et al. 2002
		Lignin content	6**	5.98–13.17		
		Pulp yield	7**	9.65–18.43		
		Extractives content	2**	4.26–13.36		
		CIE brightness	3**	7.76–13.74		
		Mean wood density	8**	6.17–12.95		
		Minimum wood density	3**	5.09–8.58		
		Maximum wood density	6**	7.50–14.51		
		Diameter	7**	5.04–14.51		
		Height growth	3**	4.93–9.78		
	Secondary metabolism	δ3Carene content	1***	26.5		Plomion et al. 1996b
	Growth traits	Stem growth index	2 ^{p=0.01}	9.23–10.56		Emebiri et al. 1997
	Wood-quality trait	Wood density 1–5 years old	1***	–	Pop identical	Kumar et al. 2000
		Wood density 6–10 years old	0	–	to Ball 2001,	
		Wood density 14 years old	0	–	LG3 only, multitrait	
	Wood-quality trait	Wood density 1–5 year-old	1***	–	model analysis Pop identical	Ball 2001

Table 5. (continued)

Species	Type of trait	Trait (abbreviation used in the article)	#QTLs ^a	%Variance explained by each QTL ^b	Note	Reference
<i>Pinus taeda</i>		Wood density 14 year-old	0	–	to Kumar et al. 2000, LG1 and 3 only	
	Growth and wood-quality traits	Diameter at breast height	2***_0	–		Kumar et al. 2004
		Stem straightness	2***_0	–		
		Branching cluster frequency	2***_0	–		
		Wood density	1***_0	–		
	Growth and wood-quality traits	Diameter at breast height	Verification: 2 ^{p=0.05}	Verification: 2.2 ^{all}		Devey et al. 2004a
			Bridging pop : 0			
		Juvenile wood density	Verification : 8 ^{p=0.05}	Verification: 14.1 ^{all}		
	Biotic stress tolerance		Bridging pop : 4 ^{p=0.05}			Devey et al. 2004b
		Resistance to Dothistroma needle blight	4 ^{p=0.05} in the detection population, 4 ^{p<0.05} in the validation population	detection: not estimated, validation: 1.76–4.8		
	Wood-quality trait	Wood specific gravity	5 ^{p=0.05}	23 ^{all}		Groover et al 1994
	Wood-quality trait	Wood specific gravity	4**, 1***	–	re-analysis of Groover et al. 1994	Knott et al. 1997
	Growth traits	Height increment	Base: 6***, detection: 7***	Base: 23.1–30.5 ^{all} , detection: 7.3–11.7 ^{all}	Detection population: same as Groover et al 1994,	Kaya et al. 1999
		Diameter increment	Detection : 8	Detection : 12.5–59.5 ^{all}	Knott 1997, Sewell 2000, 2002, Brown et al 2003.	
	Growth traits	Total height 2 years	2**	7.9–10.8		Gwaze et al. 2003
		Total height 3 years	1***	10.3		
		Total height 4 years	1***	12.2		
		Total height 5 years	1***	12.2		

Table 5. (continued)

Species	Type of trait	Trait (abbreviation used in the article)	#QTLs ^a	%Variance explained by each QTL ^b	Note	Reference
	Wood-quality trait	Total height 10 years	1 ^{***}	10.5		
		Growth rate	1 ^{***}	11.3		
		Wood specific gravity (individual rings and composite rings, measured for early and latewood)	23 [♣]	5.4–15.7	Same population as Groover et al. 1994 and Knott et al. 1997	Sewell et al. 2000
		Percentage of latewood (individual rings and composite rings)	16 [♣]	5.5–12.3		
		Average microfibrilar angle (individual rings and composite rings, measured for early and latewood)	7 [♣]	5.4–11.9		
	Wood-quality trait	Cell wall content	8 [♣]	5.3–12.7	Same population as Groover et al. 1994 and Knott et al. 1997	Sewell et al. 2002
	Wood-quality traits	Wood-specific gravity	V : 18 [♣] , U : 5 [♣]	V : 1.7–5.7, U : 1.8–4.4	Detection population: same as	Brown et al. 2003
		Percentage of latewood (individual rings and composite rings)	V : 12 [♣] , U : 5 [♣]	V : 1.7–5.7, U : 1.8–4.4	Groover et al 1994, Knott et al. 1997,	
		Average microfibrilar angle (individual rings and composite rings, measured for early and latewood)	V : 4 [♣] , U : 2 [♣]	V : 1.7–5.7, U : 1.8–4.4	Sewell et al. 2000, 2002	
		Cell wall content	V : 10 [♣]	V : 1.7–5.7		

ent genetic and environmental backgrounds.” Indeed, estimation of QTL stability (position and phenotypic variation explained) is one of the most critical factors if QTL analysis is to be performed for application in MAS. This question has been addressed at different levels, including the stability of QTLs across different developmental stages (ontogenic or cambial age effect), time points during the growing season (seasonal effect), environments, and among diverse genetic backgrounds.

Ontogenic effect: Given the long-lived characteristic of forest trees, it has been questioned whether or not the same genomic regions would control quantitative traits (e.g., annual growth, density) as trees mature. Several experiments have been conducted to answer this question (Plomion et al. 1996a; Emebiri et al. 1997; Kaya et al. 1999; Sewell et al. 2000, 2002; Weng et al. 2002; Brown et al. 2003; Gwaze et al. 2003; Pot 2004). In general, a rather low QTL stability has been noted across different maturation stages for growth (Brown et al. 2003; Pot 2004). In only one case did the authors report on the detection of the same QTL regions at different maturation stages (Gwaze et al. 2003). Conversely, Kaya et al. (1999) did not find a single “common” QTL controlling growth rate through tree development. The same trend – low QTL stability across maturation stages – was observed for wood density, a trait that presents a higher juvenile-mature correlation than growth (e.g., Williams and Megraw 1993, Hannrup and Ekberg 1998). This unexpected result suggests that some of the wood-density QTLs probably reflect the genotypic response to annual climatic variation. Recently, Rozenberg et al. (2002) reported on the alteration of wood-density profiles in response to drought. More interestingly, they showed that the alteration of annual wood-density profiles (the presence of a false late wood ring in the early wood zone) in response to drought was genetically controlled.

Seasonal effect: Sewell et al. (2000, 2002) and Pot (2004) analyzed the seasonal stability of growth and wood-quality QTLs for traits measured in spring (early wood) or summer (late wood). Overall, half of the detected QTLs were specific to one type of wood (early vs. late wood). These results agree with recent transcriptome studies that reveal that different sets of genes are regulated throughout the growing season (Le Provost et al. 2003; Egertsdotter et al. 2004).

Environmental effect: although several studies have been performed in multisite trials (Groover et al.

1994; Knott et al. 1997; Kaya et al. 1999; Sewell et al. 2000, 2002; Shepherd et al. 2002b; Weng et al. 2002; Brown et al. 2003; Devey et al. 2004a, b), in only one study did the authors analyze QTL stability at different sites (Groover et al. 1994).

Genetic effect: a complete understanding of the genetic variability of the traits of interest will rely on the analysis of multiple populations as all the major genes involved in the genetic control of a given trait are unlikely to be polymorphic in a single family. On a more practical side, QTL stability across different genetic backgrounds is a prerequisite to marker-assisted breeding (MAB) in multiparental tree breeding programs. As underlined by Brown et al. (2003), MAB will reveal its full potential under two scenarios: (1) if a genetic marker in full LD with molecular polymorphism causing trait variation at the population level is discovered or (2) if the gene (polymorphism) underlying a QTL is identified. Given their allogamous reproductive system and their recent domestication, pines are characterized by high levels of genetic diversity and low levels of LD (reviewed in González-Martínez et al. 2006b). The combination of these two factors (high diversity and low LD), together with their perennial characteristics (maturation, environmental heterogeneity), is likely to contribute to QTL instability across genotypes. Yet few studies have addressed this important issue in pines. Kaya et al. (1999) did not find any QTL shared between pedigrees, while Brown et al. (2003) and Devey et al. (2004a, b) found only a small fraction of the detected QTLs to be common across different genetic backgrounds.

Multiple-Trait QTL Analysis As stated earlier, multiple-trait QTL analysis will likely become very important for breeding purposes since pine breeding is a multitrait process. In several QTL experiments, more than one trait (e.g., growth, wood quality) was studied. In most cases, colocalizations between QTLs for different traits were observed, which might be expected for highly correlated traits. Possibly more important was the occurrence of multiple colocalizations in the genome, suggesting the effect of pleiotropic genes rather than the existence of physically linked genes controlling different traits. It should be noted that QTL clusters were also observed for traits that were not phenotypically correlated (Brown et al. 2003; Pot et al. 2005b), suggesting strong environmental/developmental effects masking genetic correlation.

2.3.3

Future Direction on QTL Mapping

As underlined in the previous section, the most reliable QTLs – from a breeding perspective – are those that have been consistently detected at different developmental stages, in different environments, and in diverse genetic backgrounds. However, it is important to remind the reader that most of the studies performed so far in pines have been based on single pedigree analysis, and that only a handful of experiments have attempted to validate QTLs across different genetic backgrounds (Brown et al. 2003; Devey et al. 2004a, b). Considering that pine improvement involves the deployment of many families/clones, the genetic stability of marker-trait association is a prerequisite before any extended use of molecular markers is considered in operational breeding programs. There have been two major attempts to identify diagnostic markers for MAB either using neutral markers spanning the genome or selecting candidate genes based on their coincidence with QTLs.

Complex Designs for Detecting Marker-Trait Associations: LD Mapping

As underlined in Guevara et al. (2005b), while QTL identification is based on physical LD generated in one or a few generations of crossing, association mapping or LD mapping takes advantage of events that created associations in the past to find a statistical association between molecular markers and a phenotype on a much finer scale. As presented in Sect. 2.1.6., five studies have provided valuable information regarding the extent of LD in pines. In all cases, it was found that LD extended only over short distances. In *P. taeda*, Brown et al. (2004) and González-Martínez et al. (2006a) revealed a rapid decay of LD within 800 to 2,000 bp in candidate genes for wood quality and drought-stress response. In *P. sylvestris* a rapid decay of LD was also detected by Dvornyk et al. (2003) and García-Gil et al. (2003). The genotyping of a breeding population of *P. radiata* with microsatellites (Kumar et al. 2004) also yielded the same result, i.e., no significant LD was observed between pairs of genetically linked markers, suggesting that LD decreases rapidly with physical distance.

LD mapping is only in its infancy in pines. To our knowledge, only one study has reported the use of molecular markers (SSRs) in unrelated trees (Kumar et al. 2004). Marker-trait associations were analyzed in a *P. radiata* trial consisting of 45 parents (40 males

and 5 females). Parental trees were genotyped and the association between parental genotypes and the performance of 200 full-sib-generated families were analyzed according to the strategy used in larch and eucalyptus by Arcade et al. (1996) and Verhaegen et al. (1998), respectively. This analysis allowed the identification of several significant associations between markers and traits. However, it is important to note that none of the marker-trait associations found using full-sib family performance were identified when the parental general combining ability was regressed on the allelic frequencies of the marker. This result suggests that the first associations detected were probably biased due to population structure (only five females), as in larch and eucalyptus.

Pan-genomic LD mapping in pines will require an extremely high marker density, given the low extent of LD as estimated so far within the few genes analyzed. Alternatively, marker-trait association can be performed on selected candidate genes.

The Candidate-Gene Approach

Candidate genes (CGs) can be proposed based on the coincidence between QTLs and known functional genes putatively involved in the genetic control of the trait. Such positional CGs have been described in *P. taeda* (Brown et al. 2003) and *P. pinaster* (Chagné et al. 2003; Pot et al. 2005b) for wood-quality-related traits. In *P. taeda*, colocalizations between genes involved in monolignol biosynthesis (4CL, C4H, C3H, and CcOAOMT) and QTLs for wood density were observed (Brown et al. 2003). In *P. pinaster*, a single candidate gene-QTL colocalization was found between KORRIGAN, a gene involved in the hemicellulose/cellulose biosynthesis, and QTLs for hemicellulose and fiber characteristics.

Other types of analysis can be used to select CGs related to plant adaptation. As underlined by González-Martínez et al. (2006b), “standard neutrality tests applied to DNA sequence variation data can be used to select candidate genes or amino acid sites that are putatively under selection for association mapping.” Unusual patterns of nucleotide diversity and/or population differentiation have been detected in *P. taeda* (González-Martínez et al. 2006a), *P. pinaster*, and *P. radiata* (Pot et al. 2005a). Pot et al. (2005a) observed singular patterns of nucleotide diversity in three genes: a glycin-rich protein homolog that was found to be up-regulated in late wood-forming tissue (Le Provost et al. 2003); *CesA3*, a cellulose synthase gene; and *KORRI-*

GAN, a membrane-bound endo-1,4-beta-glucanase involved in cellulose/cellulose biosynthesis. As mentioned before, colocalization between *KORRIGAN* and wood-quality QTLs has been reported (Pot et al. 2005b). In *P. taeda*, although the action of neutral processes cannot be completely ruled out to explain the patterns of nucleotide diversity observed for *CcoA-OMT1*, several characteristics of its nucleotide diversity seem to indicate the action of natural selection on this gene. The colocalization of this gene with a QTL of water use efficiency (Brendel et al. 2002; Pot et al. 2005b) and the differential expression of this protein under different watering regimes (Costa et al. 1998) clearly emphasize the putative role of this gene in wood-trait variation. Association studies should now be used to validate this hypothesis.

2.4

Marker-Assisted Breeding

This section discusses the application of information from DNA polymorphisms in conifer-type tree-improvement programs. Note here that such information includes not only purposefully designed markers but also information obtained directly from (re)sequencing, for which specific markers may not have been designed. Applications of information from DNA polymorphisms fall into four generic areas: audit and quality control, elucidation of genetic phenomena, population management, and selection and breeding. We discuss each of these aspects below, covering both existing and potential applications. Furthermore, we provide examples where appropriate, as well as comments regarding the current status of each of these applications.

2.4.1

Quality Control and Audit

Development of an array of DNA marker systems over the past two decades has provided tree breeding programs with a range of tools to achieve basic aspects of quality control that hitherto could only be addressed with difficulty, if at all. Seedlot and/or clonal fidelity have been the key concerns for both commercial and research applications. Indeed, this area was the earliest commercial application of DNA markers in tree breeding programs and is still the most widespread, at least in coniferous species.

From a commercial perspective, the key objective is assurance of genetic gain by ensuring that seedlots and/or clones are true to intention. The applicability and efficacy of various DNA marker systems is dependent, therefore, on the way in which genetic gain is delivered. In conifers, such delivery can come in a number of different forms (and costs), even within the same breeding program.

Open-Pollinated Seed

Open-pollinated seed from a mixture of selected seed orchard material generally consists of a few dozen maternal parents, with little if any control of the pollen source. This is a common means of seed production, particularly in commercial *Pinus* species in the south-eastern United States. In such cases, there may be a need to ensure no contributions from unwanted maternal parents, which can be achieved by genotyping megagametophytes with sufficient markers, assuming maternal parentage information is maintained for all seed. Furthermore, markers could be used to quantify the relative contributions of each maternal parent to individual seedlots if seed counts are not available. A variation on this method of seed production is used in mass pollination techniques, such as supplemental mass pollination or liquid pollination. In these approaches, receptive conelets are pollinated using pollens from selected genotypes by various means, but the conelets themselves are not actually covered to prevent fertilization by unwanted pollens.

Control-Pollinated Seed

Control-pollinated seed, where pollens from selected parents are used to fertilize selected seed parents, in the process excluding pollen from other sources using bags to cover receptive cones and/or undertaking pollinations in contained greenhouse facilities. This is usually undertaken to produce full-sib families or individual half-sib families with known selected pollen parents. Such methods are used in both breeding and commercial production for species such as *P. radiata*. Vegetative propagation, sometimes involving in vitro technologies, can be used to amplify genotypes, particularly for commercial production, largely because of shortages of seed and/or cost of seed production. DNA markers – particularly codominant multiallelic marker systems such as microsatellites – have been developed for such purposes (e.g., Devey et al. 2003). In addition, paternal parentage can sometimes be evaluated via paternally inherited chloroplast markers. Such mark-

ers have been developed and applied in *P. radiata* for checking paternal inheritance (Kent and Richardson 1997), although judiciously chosen nuclear DNA markers may also suffice. In general, markers have shown that misidentification is common in breeding programs; for example, Bell et al. (2004) estimated that 2.6% of parents were misidentified in a sample of an Australian breeding population of *P. radiata* and that 8.4% of offspring of ten families were not consistent with expected parentage. These results also indicate that a proportion of misidentification of open-pollinated seedlots is possible. Furthermore, our experience at Scion with putative full-sib families used for gene mapping experiments has revealed very few such families – produced either commercially or by research groups – are completely consistent with expected parentage (unpublished data). DNA markers, particularly codominant marker systems, have therefore been useful in checking and assuring parentage.

Clonal Production

Clonal production is used either for experimental purposes (such as clonal tests) or for mass-propagation of tested clones for clonal forestry. In these cases, genotyping is undertaken to ensure that ramets do represent the desired genotype(s). Similarly, ramets deployed in seed orchards raise the same issue, although sometimes misidentification of parents is detected via parentage testing of the seed obtained via methods described above. For applications where genotype fidelity is needed, “profiling” marker systems such as RAPDs and AFLPs is useful in that they are generally cheaper to both develop and use for this specific application, particularly as the high level of polymorphism revealed overcomes issues associated with dominance. However, in more recent years, fingerprinting kits have tended to utilize codominant markers (see references above), as these have been developed for other purposes (above) and are generally adequate for clonal fingerprinting, particularly if enough marker loci are used (e.g., Kirst et al. 2005b).

DNA markers have, therefore, been developed and utilized extensively in tree breeding programs, for both verifying commercially deployed materials and for ensuring that experimental materials meet the requirements. Even so, some programs still do not universally implement or rely on such genotyping, largely due to the expenses involved.

2.4.2

Elucidation of Genetic Phenomena

All genetic gain ultimately depends on DNA polymorphisms. Knowledge of the nature and effects of the polymorphisms has the potential to generate far more gain than the use of purely phenotypic data – on the parents and/or their progeny. Information from DNA polymorphisms enables the elucidation of phenomena such as understanding of the genetic architecture of trait variation, revelation of population structure and history, and detection of selection fingerprints, all of which can have direct or indirect applications in tree breeding.

The genetic architecture of trait variation can be defined as the frequencies, location, magnitude, and mode(s) of action of quantitative trait loci/nucleotide (QTL/N) effects underpinning quantitative traits. While QTL mapping has been very informative in this regard, the results are relevant only to the pedigree(s) used, not to whole populations. Association genetics may, therefore, be more relevant for understanding the genetic landscape of trait variation in forest trees.

Estimation of Genetic Variance Structures and Heritabilities

DNA markers offer new ways of obtaining some key knowledge that is fundamental to tree improvement regarding genetic parameters, in particular genetic variances, heritabilities, and correlations among traits. Such knowledge informs the breeder not only about the feasibility of breeding for certain traits or combinations thereof, but also how it might be efficiently undertaken.

For obtaining genetic gain, a trait must be both variable and heritable. Genetic correlations between economic traits can be a major constraint if they are adverse, but they can provide the breeder with great opportunities if they are favorable. Even genetic correlations between noneconomic indicator traits and economic ones can be used to great advantage if the correlations are strong and the indicator traits highly heritable. For genetic variances and heritabilities, if they have to be estimated from seed collected in natural stands, then the coefficients of relationship within seed-parent families need to be known. Indeed, the relative contributions of inbreeding as such and finite effective numbers of (unrelated) pollen parents per seed parent represent important information. While in the past isozymes were a valuable

tool for providing such information, DNA markers can be much more powerful for the purpose, and with a greater range of applications. This issue of nonrandom components in the mating system can be important with native stands of conifers (Burdon et al. 1992), despite their wind pollination, even if it is less acute than in insect-pollinated species like eucalypts (Hodge et al. 1996). The wind pollination of conifers tends to reduce the nonrandomness of mating, while the conifers' mechanism of archegonial polyembryony probably reduces the inbreeding component still further. For estimating genetic correlations between traits, knowledge of coefficients of relationship within such families is likely to be much less crucial. As we will mention later, *a priori* family information as such may not be essential for this purpose, but it can be supplemented with information from markers.

Population Structuring and History

The structuring and history of populations influence both the availability of genetic variation for the breeder to exploit and the potential for association genetics to contribute. While large, essentially panmictic populations cannot be expected to have appreciable across-family linkage disequilibrium (LD), cryptic structuring may exist that generates significant disequilibrium that could be useful for breeders. For example, localized population bottlenecks, followed by coalescences, could easily cause this. Such LD could provide valuable clues to "metapopulation" history. Despite wind pollination, various factors can generate population structure in conifers (Mitton 1992). Interesting possibilities of structure exist in populations derived from recent admixtures. In *P. radiata*, the exotic, domesticated "landraces" still have large elements of the wild state. Interestingly, they evidently represent a genetically recent fusion of two of the native populations, Año Nuevo and Monterey (Burdon et al. 1998), which may provide a basis for some admixture disequilibrium.

Polymorphisms revealed by DNA sequence data derived from both genic and nongenic regions can reveal much about the genetic history of those regions. Departures from Hardy-Weinberg equilibrium could reveal the presence of previously undetected genetic phenomena such as the presence/absence of inbreeding. Indeed, genetic variance (and gain) estimates are based on assumptions regarding the relatedness of parents used in genetic tests. Such data can be used to

check these assumptions and provide empirical data for more accurate estimates. Similarly, sequence data from genic regions can reveal evidence of selection: for example, balancing selection was detected by Cato et al. (2006b) in a gene associated with wood density and growth rate in *P. radiata*. Krutovsky and Neale (2005) found evidence for selection in three of 18 genes in Douglas-fir. Such evidence – which can be generated on a relatively small subset of genotypes – could be an effective prescreen for genes more likely to be associated with trait variations, although some caveats apply regarding the power to detect the effects of selection (Wright and Gaut 2005).

Assignment of Gene Function

Knowledge of gene function, if acquired, offers the greatest long-term opportunities to capture genetic gain, using either endogenous variation or genes introduced by conventional breeding or genetic engineering. LD mapping and association genetics can provide clues in the search for quantitative trait nucleotides (QTNs) regarding which genes have functional roles in trait variation. While relatively short stretches of disequilibrium (usually hundreds to low thousands of base pairs) represent constraints for breeding applications, a key advantage is the potential for assignment of function – even identification of individual QTNs. However, because of the size of conifer genomes and the lack of genomic sequence, identifying candidate genes will be crucial. Possible approaches are described in the following section and in more detail by Wilcox et al. (2006).

In carefully selected cases, genetic transformation can be used to verify the role of a QTN in generating phenotypic variation. The costs of achieving a transformation, and often the regulatory issues, will demand a highly selective application of this approach. On the other hand, knowledge of gene function may be useful for identifying genes to target for genetic transformation, to create new variations of use for breeders, and to provide commercial cultivars. There are no reports yet of cloning QTLs from tree species, partly due to the large number of candidates within QTL confidence intervals, but also because of the time required for trait expression of transformants arising from complementation studies. Nonetheless, association genetics will be a key step in increasing resolution and, in some cases, identifying putative QTN for further analyses.

2.4.3

Population Management

Pedigree Reconstruction and Detection of Genetic Contamination

Breeding populations represent the “engine room” for capturing the additive gene effects that allow cumulative genetic gain over successive generations, through recurrent cycles of selection, intermating, evaluation, selection, and so on. Maintaining full pedigree has been favored on the grounds that it helps preclude inbreeding and maintains effective population size. However, the expense of maintaining full pedigree can limit the size of the breeding population that can be handled, raising the question of whether larger populations can be handled if pair crossing is not mandatory. Moreover, there may be situations where the breeder has reason to resort to material, e.g., commercial stands, in which pedigree has been sacrificed but which has the advantage of huge numbers (Burdon 1997).

Modern marker technology has major potential for pedigree reconstruction (Lambeth et al. 2001; Kumar et al. 2006), at least in open-pollinated families of known seed parentage. Complete reconstruction of pedigree will be more challenging, especially with the wind pollination that characterizes conifers. In maintaining gene resources that underpin breeding populations, controlled crossing is typically far too expensive, yet there may be a call for quantifying and even detecting individual cases of contamination (Burdon and Kumar 2003). Different types of markers may be required for this purpose, and the task may be challenging, but it appears inherently feasible. Pedigree reconstruction is discussed later, in connection with selection.

Tracking and Maintenance of Genetic Diversity

A key element of population management is maintenance of genetic diversity, to give the breeder flexibility in both the short and long terms and to safeguard continuing long-term genetic gain. While pedigree information is an indicator of genetic diversity, it does not provide definitive information in itself. Achieving that poses significant challenges.

The “gold standard” for functional genetic diversity will usually be performance in well-designed and properly located common-garden genetic experiments. However, such experiments are expensive and often slow to deliver results. The use of DNA polymorphisms is clearly much quicker, but such marker

diversity will need to be cross-referenced with the functional diversity since the two classes of diversity are not necessarily closely coupled, at least among species (Morgante and Salamini 2003; Paran and Zamir 2003). Components of DNA diversity obviously include percentage of polymorphic genes (in either the coding or regulatory regions), percentage of base pairs that are polymorphic, particularly for QTN that must exist in coding or regulatory regions), and allele frequencies for the polymorphisms. Such information, in conjunction with knowledge of magnitudes of QTN effects, provides a benchmark for monitoring changes in diversity, for any forest trees.

Loss of low-frequency alleles is an obvious manifestation of a decrease in genetic diversity. Paradoxically, abrupt increases in the frequencies of such alleles, as can occur through genetic drift, can be a manifestation of the same phenomenon. While the inherently outbreeding genetic systems of forest trees may be able to cope with significant inbreeding in the wild, through selection for balanced heterozygotes, such mechanisms may be impeded under conditions of artificial breeding. In principle, almost any sort of genetic marker should be able to manifest the losses of alleles or sharp fluctuations in their frequencies. Nevertheless, it seems preferable to know what genes are of particular current or contingent importance and monitor their frequencies. This, however, will depend on knowing the functions of the genes.

Provision for Biotic Crises

While it appears QTNs typically exert minor individual effects in conifers, disease-resistance genes can represent a notable exception (Burdon 2001). Such genes can both have large effects (despite outward appearances of classical quantitative inheritance) (e.g., Kinloch et al. 1970; Wilcox et al. 1996) and be present at low frequencies (R.D. Burdon and P.L. Wilcox personal communication). These genes can also have the feature of gene-for-gene specificities between host genotypes and fungal strains (pathotypes), which has important implications for ensuring durability of disease resistance against pathogen mutations (Burdon 2001). Establishing the nature of such genes can require carefully planned mating between parents and inoculation studies based on single-spore isolates, preferably backed up with genomic studies. Here, as in other areas, comparative genomics can have a major role, at least in studying resistance genes of lower specificity.

The implications of such patterns of genetic variation within both hosts and pathogens, which are called

pathosystems, can be major for both population management and selection. Disease resistance can be very important in plantation forestry, especially with exotics that are being grown in the absence of natural or other pathogens. Conserving low-frequency genes of large effect, which are sometimes important in conifers, can pose a threefold challenge. A large population may be needed to find trees with such resistance. The requisite population size can be increased by the desirability of obtaining resistance genes in unrelated pedigrees. It may be further increased by the desirability of combining ("pyramiding") genes in the genetic material that represent diverse mechanisms of resistance to the pathogen, toward ensuring durability of resistance against mutation or genetic shifts in the pathogen population. In implementing this approach, identification of resistance genes will be of enormous help.

The information on forest-tree pathosystems, in general, is still very sketchy, although there are resistance genes identified for some coevolved forest pathosystems (e.g., Wilcox et al. 1996). Nevertheless, what is known indicates that preparations for biotic crises in the form of new fungal diseases should include having very large population resources available. In at least some cases, it is very doubtful whether breeding programs effectively contain such a provision (Burdon and Gea 2006).

2.4.4

Selection and Breeding

Pedigree Reconstruction as an Alternative to Maintaining Pedigree Records

A proposed application (Lambeth et al. 2001) is to use markers in genetic tests to reconstruct pedigrees retrospectively, as opposed to maintaining pedigree information throughout the life of a genetic test. This involves applying pollen mixes of known composition to a range of seed parents, planting the resultant offspring in designed experiments with limited or no maintenance of family information (Kumar et al. 2006). Upon subsequent measurement, pedigree identity is ascertained via DNA markers. Benefits of such an approach include logistical simplicity, cost reductions for breeding and testing, potentially better estimates of genetic parameters and increased gains, and assurance of parentage (see above). Furthermore, fewer financial costs are incurred if experiments are abandoned or lost prior to remeasurement – which does happen in tree-improvement programs. Disad-

vantages include underrepresentation of some specific families due to either chance or factors affecting fertilization, prevalence of inbreeding in some families distorting breeding-value estimates of those families, difficulties in detecting full-sib-specific combining ability in full-sib families, and genotyping costs. Most of these disadvantages could be overcome, although further research is needed to fully evaluate various possible application scenarios. To date, this approach is currently being evaluated in the context of operational breeding in New Zealand with *Eucalyptus* and *P. radiata*.

Estimation of Heritability Based on Marker-Ascertained Relatedness

Because molecular markers have the potential to estimate relatedness between genotypes, methods to estimate the heritability of phenotypic characteristics have been proposed and evaluated (Andrew et al. 2005; Kumar and Richardson 2005) without the need to have any prior knowledge of genetic relationships (see above). Such methods have the potential benefit of allowing one to obtain information from existing forests (e.g., natural forests or commercial plantations), thereby obviating the need for genetic testing involving progenies, thus speeding up the generation of information, particularly for species where little a priori information is available. Some disadvantages involve the reliability of marker-based estimates, particularly as assumptions need to be either made or checked regarding genetic structure and prior levels of relatedness, as well as the reliability of phenotypic information, particularly if sourced from forests rather than purpose-designed common-garden tests with appropriate controls. These approaches have not been extensively investigated but may have a role. In part, this may be due to the fact that existing approaches are well established and successful, as well as to the existence of infrastructure such as extensive testing involving progenies. Kumar and Richardson (2005) compared phenotype-based with marker-based heritability estimates for wood density in *P. radiata* and found very little difference. Similarly, Andrew et al. (2005) reported nonzero estimates of heritability for a number of foliar defence-related chemicals in *Eucalyptus melliodora*, some of which were consistent with independent estimates based on phenotype (see Andrew et al. 2005 and references therein). Such approaches may be of use also for association genetics, particularly in revealing cryptic population structuring in natural forests, as well as

for selecting maximally unrelated individuals for estimating LD.

Within-Family Selection Based on Marker-Trait Associations Derived from Pedigreed QTL Mapping Populations

Such selection is based upon selecting individuals within known full- or half-sib families where marker-trait associations have been previously determined. Genotypes with the desired multilocus genotypic composition are selected from within families. In theory, this can be applied within families in breeding populations and/or production populations. Generic benefits include earlier selection, increased selection intensity, and potentially cheaper selection (Stromberg et al. 1994). In conifer improvement, breeding populations are usually based on a few hundred parent genotypes; thus it is largely impractical and very expensive to detect associations for each genotype independently (Strauss et al. 1992; Johnson et al. 2000). Rather, application of markers is more likely to be restricted to populations where specific families are advanced, as for example in elite populations or in small nucleus- or mainline breeding populations.

A number of situations have been evaluated either for species-specific scenarios or from a wider theoretical perspective. One of the earliest such studies was that by Strauss et al. (1992), who concluded that within-family marker-assisted selection (MAS) was of limited or no value unless a high proportion of additive genetic variation could be explained by markers for traits of low heritability but high value and where selection intensities within families were high compared to that among families. They also concluded that difficult- or expensive-to-measure traits such as wood quality or resistance to certain diseases showed the most potential for MAS. Williams and Neale (1992) evaluated the relative efficiency (RE) of MAS and also concluded RE was greatest where (a) traits were not expressed sufficiently early to enable a reduction in generation length as well as (b) with lower-heritability traits where markers explain a high proportion of additive genetic variability for the traits. However, results from many QTL mapping experiments have indicated that the genetic architecture of quantitatively inherited traits is dominated by genes of small effect (Wilcox et al. 1997; Sewell and Neale 2000; Brown et al. 2003; Devey et al. 2004a, b), thus limiting the opportunity to explain sufficient variation with markers.

Wu (2002) evaluated the tradeoff between proportion of variance explained by markers and shortening of selection interval. For traits of moderate heritability (0.2 to 0.4) marker-assisted early selection (MAES) was more efficient than phenotypic selection, if they explained more than 5 to 10% of additive genetic variance and allowed the selection interval to be reduced by half. For later-expressed traits, even less variance explained by markers was necessary for markers to be more efficient than phenotypic selection. Wu (2002) also evaluated a number of other scenarios, including combining phenotypic selection with markers, and found that MAES is only marginally more efficient than phenotypic selection except in cases of relatively low juvenile-mature phenotypic correlations. Given genetic architectures of QTLs explaining a few percent each (Wilcox et al. 1997; Brown et al. 2003; Devey et al. 2004a, b), and limitations for the selection of up to ten unlinked markers per family linked to such QTLs, the effectiveness of early selection is limited for traits with low-moderate age-age phenotypic correlations, particularly once costs of QTL detection are taken into account. Time savings for traits of delayed expression or low juvenile-mature correlation will, however, depend on the prior establishment of QTL/phenotype associations.

All the above studies investigating the application of markers for breeding population advancement have generically shown that while there are scenarios where markers could be used to generate genetic gains, the financial gains could be quite limited. This is in part due to the cost of genotyping as well as the length of time, meaning that additional costs will be incurred. A further limitation stems from the fact that most breeding objectives involve multiple quantitatively inherited traits whose genetic architectures are likely to involve predominantly small-effect genes (see references above). This means limited genetic gains from individual markers. Furthermore, relatively few markers can be used to select within any particular family: for a simple two-genotype-per-marker scenario, it would be necessary to generate and genotype thousands of offspring per family to have sufficient power to generate the optimal ten-marker-locus genotype. If such selection were restricted to a limited range of traits, there would also be the need to augment marker information with phenotypic records to effectively address the breeding objective; therefore the opportunity for earlier selection could be lost. On the other hand, few studies have investigated the efficacy of marker-based or marker-assisted selection

over multiple generations. Application over generations is likely to reduce costs, although not in a manner linearly proportional to gains. This is owing to the fact that additional revenues in future generations are discounted relative to cost, as costs are effectively incurred early (e.g., Johnson et al. 2000).

An exception to the limitations outlined above is the selection for genes of large effect such as major-gene disease resistance (see references above). In breeding populations, frequencies of resistance genes could be increased much faster via marker-based selection than phenotypic selection; markers could obviate most of the need for field and/or greenhouse screening by allowing only those individuals to be deployed in field tests (for screening for other traits) that have the favorable marker phenotypes. Where gene pyramiding is crucial for durable resistance, this marker-based approach could be especially valuable. As with all marker-assisted (or based) selection scenarios, such genes need to be detected, which incurs costs early in the breeding cycle. However, such costs are likely to be lower in that such large-effect genes are less financially burdensome to detect, and in cases where resistance is valuable, financial gains can be considerable, particularly if undertaking development of disease-resistant (or tolerant) breeds.

Opportunities for MAS have also been evaluated for production populations in species such as Douglas-fir (*Pseudotsuga menziesii*) (Johnson et al. 2000) and *Pinus radiata* (Wilcox et al. 2001b), and for clonal deployment in *P. radiata* (Kumar and Garrick 2001). Results in general indicate that gain from MAS is possible in *P. radiata* for a range of options but marginal for Douglas fir, implying rotation length and product value are both important. For example, Wilcox et al. (2001b) showed that even modest gains in physical traits of 3.0 to 3.4% resulted in product value gains in excess of 9% and internal rates of return ranging from 9.1 to 21%. However, a key condition here was the need to achieve adequate multiplication rates via vegetative propagation. Propagation technologies are key to MAS being cost-effective. Johnson et al. (2000) showed that modest genetic and financial gains were possible for MAS in Douglas fir for production population applications, although results were highly dependent upon assumed genetic architectures and relatively large areas of plantations were needed to justify the extra costs associated with MAS.

Given all of the above, it is not surprising that relatively few breeding programs are actively pursuing MAS, particularly for quantitatively inherited

characteristics. Some private companies in the USA and New Zealand have undertaken or are still undertaking MAS on a limited scale, but there is as yet no widespread uptake either for breeding-population advancement or as a tool for more immediate genetic gains. Nonetheless, there is still some potential, as some of the above studies have indicated, as well as other possible areas of application yet to be explored. It may be, however, that other technologies, e.g., GAS (see below) or genetic modification, may supersede MAS.

Combined Among- and Within-family Selection Based upon Results from Association Genetics Experiments

Advances in genomics technologies over the past 5 to 10 years have made possible almost unrestricted access to any region of tree genomes, particularly sequences within and associated with expressed genes. Variation in genic regions and the associated regulatory regions form the basis of phenotypic variation. LD mapping and association genetics are key tools in correlating such sequence variation with observed trait variation. Unlike marker-trait associations from pedigreed QTL detection populations (i.e., families), associations derived from association genetics have applicability for both within- and among-family selection. The term gene-assisted selection (GAS) has been used to describe this method of selection (Wilcox et al. 2003, 2006; Wilcox and Burdon 2006); it differentiates the manner in which markers are found (i.e., via association genetics) and utilized from MAS (above).

Because association genetics is relatively new to forest trees (Neale and Savolainen 2004; Wilcox and Burdon 2006; Wilcox et al. 2006) – and many other plant species (Flint-Garcia et al. 2003) – very few quantitative analyses have been undertaken to date that involve detailed specific strategies for incorporation into tree breeding programs. Wilcox et al. (2006) described a range of applications within tree breeding programs, which are summarized below.

Conifer breeding programs can be generically characterized as consisting of highly heterozygous genotypes in hierarchically arranged (and managed) populations, with multitrait breeding objectives. To be effective, marker-trait associations derived from association genetics must integrate within this scheme, as with MAS. A key generic benefit of GAS in this regard is application to *both* within- and among-family selection, in contrast to MAS, which is re-

stricted to those families in which the associations have been detected. Association genetics, therefore, has application to all levels of population hierarchies, from essentially unimproved germplasm, through to advanced-generation lines developed for clonal deployment.

A further benefit would be selection at the seedling stage, much sooner than full trait expression – as with MAS. For most commercially important pine species, phenotypic selection is applied at 6 to 12 years of age, with the onset of reproductive competency occurring slightly earlier. Opportunity exists for early selection to either increase selection intensities via a multistage approach as with MAS (Wu 2002) or undertake an additional round of selection prior to finalizing genotypes for deployment (Wilcox et al. 2001b). Where onset of reproductive maturity precedes trait expression, there is also opportunity to reduce breeding-cycle length, although this would require known marker-trait associations for at least the majority of traits in the breeding objective(s). The opportunity for selection among as well as within families provides even greater opportunity to shorten breeding cycles compared to MAS.

An additional benefit is cost reduction relative to phenotypic selection. Once marker-trait associations are detected, genotyping costs are generally somewhat lower as compared to phenotypic evaluation. Such evaluations, however, are considerably more expensive than obtaining sufficient markers for genome coverage for linkage and QTL mapping as well as for MAS purposes, as virtually all polymorphisms within and associated with expressed genes need to be detected – which, on a whole-transcriptome basis, entails major effort. Such information nonetheless has other potential applications (e.g., prescreening genes for genetic modification experiments), so the costs could be spread over several funding sources and applications.

Because of the breadth of potential selection applications within any one breeding program, as well as the fact that conifer breeding programs all differ in some regard from one another, the application of association genetics will need case-by-case evaluation – as has been the case with MAS – ultimately needing numerical simulation to evaluate different scenarios. However, although a number of breeding programs are involved in some of the basic research required for detecting marker-trait associations, as yet there are no published reports of evaluations of strategies that incorporate association genetics within conifer

breeding programs. We expect this will change as results from association genetics and LD experiments become available.

Indeed, the outlook from results to date is cautiously favorable, as marker-trait associations have been reported for a number of tree species, including *P. taeda* (Brown et al. 2004), *P. radiata* (Cato et al. 2006b), and *Eucalyptus nitens* (Thumma et al. 2005), although all of these need to be independently verified. Results from a range of other plant species are also emerging (see Gupta et al. 2005 for a review), also with encouraging messages.

Given the size of gymnosperm genomes and the lack of widespread genomic DNA sequence for most hardwood species, it appears that sequences within and associated with expressed genes are likely to be investigated. But how should such sequences be chosen? Possible approaches are as follows.

- Identifying gene sequences using EST libraries from both angiosperms and conifers, and complete genomic sequences from model angiosperm species to select numerous genes for high-throughput (re)sequencing. This approach is being used in *P. taeda* (see <http://dendrome.ucdavis.edu/adept2>).
- Expression information at the mRNA level, via *in silico* expression profiling or application of a number of gene expression profiling technologies. Protein-expression information is also available for some species.
- Selecting genes on the basis of biochemical role(s) and/or known sequence motifs.
- Information from colocalization of expressed genes and QTL from pedigreed QTL mapping populations, within either the species of interest or related species.
- Combinations of the above, for example gene- or protein-expression information combined with QTL mapping (Kirst et al. 2004).
- Genes shown to be involved in trait variation via genetic modification experiments.

Technical advances in “omics” technologies may also offer a number of other approaches.

The undoubted importance of comparative genomics, especially for establishing gene function, means that genomic information is needed on other species, in addition to the classical model species like *Arabidopsis*. This need is actually being met, to some degree at least. In conifers, parallel research is proceeding in *P. radiata* in Australasia, *P. taeda* in the

southeastern USA, and *P. pinaster* in Europe. The evidently close synteny among these species promises great opportunities to apply genomic information on any one species to the others. Using these as model species within conifers should allow one to capitalize on the most favorable model features of all the species at once. Similarly, whole-genome sequences of *Populus* and *Eucalyptus* will allow for even more rapid advances in these species.

Should association genetics be utilized by tree breeders, then a number of other requirements need to be addressed. These include:

- Populations and analytical methods for adequate detection of marker-trait association (see Ball 2005),
- Suitable laboratory and bioinformatics capabilities functionally integrated into tree-improvement programs, and
- Development of strategic alliances among appropriate entities to effectively manage resource association genetics programs in a manner that allows operational implementation.

The above issues are discussed in more detail by Wilcox et al. (2006).

2.4.5 Summary of Applications of DNA Polymorphisms in Conifer Breeding

Information from DNA polymorphisms has a wide range of applications for tree improvement, including quality control, pedigree reconstruction, elucidation of genetic phenomenon, monitoring and maintenance of genetic diversity, and selection and breeding based upon polymorphisms associated with trait variation. To date, only some of these applications have been implemented in operational breeding programs – largely those associated with quality control. Some potential applications, such as association genetics and pedigree reconstruction as part of operational testing, are largely in the research and development phase, while others such as within-family MAS have been sufficiently evaluated to identify at least some strategies for implementation. The lack of uptake across the spectrum of potential applications is likely due to cost, which is particularly important in tree-improvement programs, which usually take years to recoup such costs. Nonetheless, technological advances will increase the scope of applications for tree improvement.

2.5 Genomics Resources for the Genus *Pinus*

2.5.1 Efforts Toward Complete Genome Sequencing

BAC Libraries

Large-insert genomic DNA libraries in which each DNA clone is stored and archived individually (i.e., ordered libraries) are a fundamental tool in modern genomics research. The most popular large-insert vector is the bacterial artificial chromosome (BAC), which, despite its name, is not a chromosome but rather a modified bacterial plasmid (F-factor) that can stably carry large inserts (ca. 50 to 400 kb) (Zhang and Wu 2001). BAC libraries have been made for a host of taxa and employed in a variety of applications (reviewed by Zhang and Wu 2001).

Despite the importance of pine and other conifers and the value of BAC libraries to genomic research, BAC resources are extremely limited for gymnosperms. To our knowledge, there are three gymnosperm BAC libraries in existence – all of them for pine. The first pine BAC library, constructed by Islam-Faridi et al. (1998), affords $0.05\times$ coverage of the *P. taeda* genome. A second library, developed by Claros et al. (2004) for *P. pinaster*, provides considerably higher genome coverage ($0.32\times$), although the probability of finding a gene of interest is still relatively low (27%). However, the *P. pinaster* library is reportedly being expanded to $3\times$ (Claros et al. 2004), which would provide 95% probability of finding a given genome sequence (Fig. 3). In September 2004, the US National Science Foundation funded a 3-year project that included construction of a $10\times$ BAC library from the *P. taeda* genotype “7-56” (Peterson et al. 2005) as a primary objective (Fig. 4). When completed, this library will be, to our knowledge, the single largest BAC library ever made with roughly 1.7 to 2 million individual clones. As of this writing, the 7-56 BAC library affords $2\times$ coverage of the pine genome. When the 7-56 library reaches $3\times$ coverage (ca. June 2006), it will be gridded onto BAC macroarrays and screened with STS and molecular genetic markers including those associated with genes of economic importance. The resulting information will be utilized to isolate and sequence intact pine genes. Additionally, marker localization will represent the start of STS-based

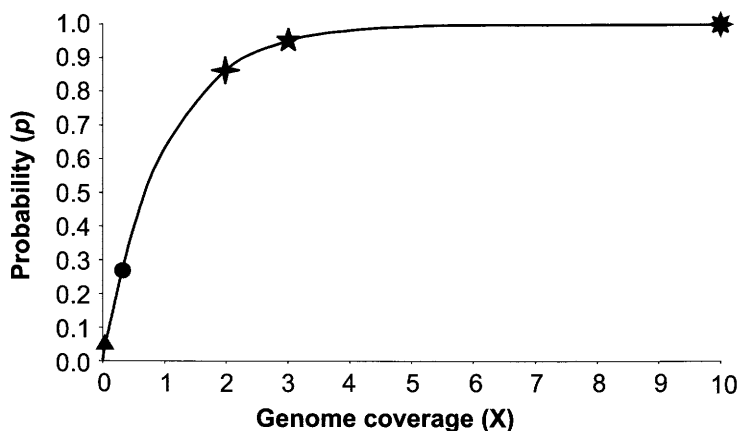


Fig. 3. The relationship between genome coverage ($\times$) provided by a BAC library and the probability (p) of finding a particular sequence of interest in that library. A $0.05\times$ BAC library, like the *P. taeda* BAC library generated by Islam-Faridi et al. (1998), affords only a 4.8% chance ($p = 0.048$) of finding a sequence of interest (triangle), while a $0.32\times$ library, like the current *P. pinaster* library (Claros et al. 2004), affords a considerably better chance ($p = 0.27$; circle). The *P. taeda* 7-56 library (Peterson et al. 2006) currently has a size of $2\times$ ($p = 0.86$; four-point star) and will be used to isolate genes of interest once it reaches $3\times$ coverage ($p = 0.95$; five-point star). At $10\times$ coverage ($p = 0.9999$; eight-point star), the *P. taeda* 7-56 BAC library will be of adequate size for essentially any use including genome-wide physical mapping and genome sequencing

physical mapping in pine. Comparison between the *P. pinaster* and *P. taeda* libraries should provide considerable insight into genome evolution within *Pinus*.

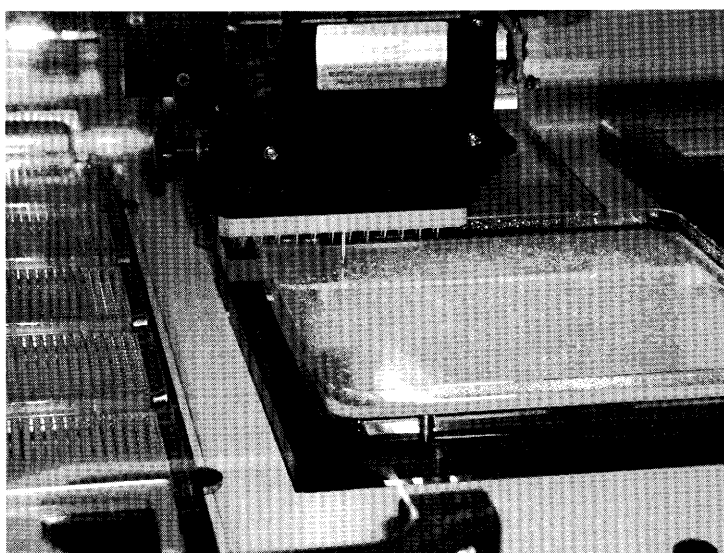
Need for Pine Genomic Sequence

Although EST sequencing in pine is relatively advanced (Sect. 2.5.2), there are few genomic sequence data available for pine or any other gymnosperm. As of 6 March 2006, the longest continuous gymnosperm/conifer genomic sequence in GenBank was only 6,884 bp. While ESTs provide the coding regions of many expressed genes, an understanding of gene function and regulation requires knowledge of those noncoding sequences that coordinate gene expression in response to biotic and abiotic cues (e.g. promoters, enhancers, silencers). Such information necessarily comes from sequencing genomic DNA.

As the pine genome is ten times larger than that of maize (the largest plant genome to be the target of a full-scale genome sequencing effort, NSF 2005), it may be a while before whole-genome sequencing of pine becomes a reality. Nonetheless, pine genomics can be greatly advanced by even relatively modest sequencing of genomic DNA, especially sequencing of large continuous pieces of DNA such as BAC inserts. The following are just a few areas of pine genome research that will benefit from more extensive sequencing of genomic DNA.

- *Genome structure:* Very little is known about the structure of any conifer genome, and relationships between genes, repeats, and pseudogenes have only been tangentially explored (e.g., Elsik and Williams 2000; Rabinowicz et al. 2005; Peterson et al. 2006). BAC and shotgun sequencing of pine DNA will provide a more detailed understanding of the sequence structure of the pine genome. Such information will be essential in the development of an efficient strategy for sequencing the *gene space* of pine.
- *Differential gene expression:* The lack of genomic sequence for pine has severely limited the study of the noncoding regulatory regions of its genes. Information on these regions is essential if we are to understand differential gene expression in pine and manipulate conifer genes in a useful and controllable manner (No et al. 2000). To determine the general tissue/development/environment specificity of the regulatory sequences for a given gene, the expression profiles of that gene can be examined; since there is already a considerable amount of expression profile data available (e.g., Egertsdotter et al. 2004; Lorenz et al. 2006), associating regulatory sequences with specific tissues and/or developmental events could be initiated immediately after sequencing. Comparison of regulatory sequences from genes with similar expression patterns can be used to gain insight into the ac-

Fig. 4. Constructing the *Pinus taeda* 7-56 BAC library. A Genetix QPixII robot picks 7-56 BAC colonies off of agar in a “Q-Tray.” Picked clones are used to inoculate media in 384-well microtiter plates (left side of image)



tivation/repression mechanisms underlying major developmental events.

- *Comparative genomics of conifers:* Pine BAC/shotgun sequences and comparative genomic approaches can be utilized to advance the study of other gymnosperm genomes, which will afford considerable insight into the evolution of this important group of organisms. Comparison of orthologous regions of conifer genomes (e.g., through sequencing of BACs and/or BAC contigs recognized by common markers) will provide high-resolution means of investigating conifer sequence evolution (see Stirling et al. 2003 and Yan et al. 2004 for angiosperm examples).
- *Gymnosperm/angiosperm comparisons:* Information garnered from pine genomic sequence should provide nearly limitless opportunities to explore comparative genome evolution between gymnosperms and angiosperms. BAC sequences from pines can be compared with existing plant genome sequences as a means of evaluating macro- and microsynteny as well as sequence conservation/divergence between pines and angiosperms. As pine has been widely used as an outgroup in angiosperm comparative research, pine genomic sequences should enable investigations by many angiosperm research groups throughout the world.
- *Polymorphism discovery and characterization:* EST sequencing has afforded tremendous insight into polymorphisms within the coding sequences of

pine genes (e.g., Krutovsky et al. 2004; Le Dantec et al. 2004). However, sequence changes in the non-coding regulatory regions of pine genes may provide as much, if not more, information on phenotypic diversity than coding sequence.

- *Association mapping:* Relatively long DNA stretches produced by BAC sequencing will facilitate genomewide studies of LD, which, in turn, will fuel association mapping projects (e.g., Brown et al. 2004; Krutovsky and Neale 2005).
- *Sequencing adaptive and economic genes of pine:* There is considerable interest in sequencing intact genes of economic importance and/or particular value to conifer/gymnosperm research. Potential target genes include those associated with outcrossing (Williams et al. 2001), embryogenesis (Ciavatta et al. 2001), disease resistance/susceptibility (e.g., Devey et al. 1995; Morse et al. 2004; Kayihan et al. 2005), abiotic stress resistance (e.g., Chang et al. 1996; Dubos and Plomion 2003; Dubos et al. 2003; Krutovsky and Neale 2005; Lorenz et al. 2006), and wood properties (Brown et al. 2003; Devey et al. 2004a; Krutovsky and Neale 2005; Pot et al. 2005b). Complete gene sequences can be utilized to explore pine/conifer evolution and facilitate pine improvement through traditional breeding, marker-aided selection, and genetic engineering. Of note is the fact that a BAC containing a target gene may well contain other genes of interest. The study of these “bonus” genes will likely be as important as studying target genes themselves.

- *Exploring the evolution of repeat sequences:* Recently a 454 Life Sciences Genome Sequencer 20 (Margulies et al. 2005) was used to generate 100 Mb of sequence from the *P. taeda* genotype 7-56 (Peterson et al. 2006). This sequence should provide considerable insight into pine genome structure, although the short length of 454 reads (ca. 100 bp each) limits the utility of the sequence. While pseudomolecule contigs have been assembled, 100 Mb affords only $0.046\times$ coverage of the pine genome, and consequently it is likely that any 454 contig of significant length is a conglomeration of sequences in a repeat sequence family rather than an individual element found in native DNA. However, repeat sequences obtained from sequenced BACs and/or contigs assembled from Sanger/capillary shotgun reads can presumably be used as scaffolds on which 454 reads can be aligned. The depth and shape of the resulting alignments should permit detailed analysis of the evolution of major repeat-sequence families in pine.
- *Genetic engineering:* Regulatory sequences obtained from genomic sequences can be attached to reporter genes and used to study promoter/regulator specificities (e.g., No et al. 2000). Such testing will provide insight into pine gene regulation, which will eventually enable insertion of constructs containing pine promoters and genes/alleles of value into conifers. In addition, the genomic sequences can serve as sources of molecular markers that can be utilized in tree improvement through MAS.

Physical Mapping

Given the enormous size of the pine genome and the time and financial resources required to conduct physical mapping research, it is likely that complete physical mapping of the pine genome will be too costly to pursue, at least with present technology. Because DNA sequencing technologies are advancing rapidly (e.g., Margulies et al. 2005), it is probable that complete shotgun sequencing of the pine genome will become affordable before complete physical mapping does. However, physical mapping of gene-rich genomic regions has been utilized to advance understanding of important chromosomal regions in many species (e.g., Folkertsma et al. 1999; Sanchez et al. 1999; Dilbirligi et al. 2004; Barker et al. 2005), and indeed, physical mapping of gene-rich regions is a goal of the research group constructing the *P. taeda* 7-56 BAC library.

Sequencing the Pine Genome

C. Plomion: Will the pine genome be sequenced?

D.G. Peterson: Yes.

C. Plomion: When?

D.G. Peterson: Not this week.

The complete sequencing and assembly of the *Arabidopsis* and rice genomes have afforded considerable insight into the evolution of higher plants including pines. For example, Kirst et al. (2003) compared the genome sequence of *Arabidopsis* with pine ESTs and showed that gymnosperms and angiosperms possess highly similar gene complements. Other plant genomes, most notably poplar, sorghum (*Sorghum bicolor* L.), and maize (*Zea mays* L.), are current subjects of full-scale genome sequence efforts, and the genome sequences of these plants will further facilitate understanding of plant genome evolution and function (Paterson et al. 2005). The poplar sequence, in particular, may provide information that will help advance understanding of wood formation in trees (Tuskan et al. 2004). However, angiosperms and gymnosperms diverged from a common ancestor more than 300 million years ago (Bowe et al. 2000), and consequently the utility of angiosperm sequences in the study of pine and other gymnosperms will be limited. Ultimately, the best means of advancing pine and conifer genomics is complete sequencing of a conifer genome. While it is now theoretically feasible to sequence the genome of any organism, the large, repetitive nature of conifer genomes will likely prevent them from being targets of full-scale genome sequencing for at least a few years. However, the likelihood that the pine genome will be sequenced is high based upon past and present investments made by the NSF (including construction of the *P. taeda* 7-56 BAC library) and other granting agencies and growing worldwide interest in pine as a biofuel/carbon sequestration crop (Jackson and Schlesinger 2004; Perlack et al. 2005).

While whole-genome sequencing may have to wait, sequencing of gene-rich BACs is likely to begin in relatively short order. Additionally, reduced-representation sequencing (RRS) methods (see Peterson 2005 for review) are being used to investigate sequence subsets of the pine genome and are likely to play a role in eventual whole-genome sequencing as well. Two RRS techniques that have been successfully utilized in maize genome exploration are methylation filtration (MF) and Cot

filtration (CF)¹ (Whitelaw et al. 2003; Springer et al. 2004).

- MF is based on the preferential hypermethylation of retroelements and hypomethylation of genes observed in some plants (Rabinowicz et al. 1999). In MF, genomic DNA fragments are ligated into a plasmid containing an antibiotic-resistance gene and the resulting recombinant molecules are used to transform a bacterial strain containing enzymes that preferentially cleave hypermethylated DNA. Linearization of a hypermethylated recombinant molecule results in its loss and makes its host cell susceptible to an antibiotic in the growth medium. Consequently, only hypomethylated sequences are successfully cloned. While a useful gene-enrichment tool in maize and sorghum (Rabinowicz et al. 1999; Bedell et al. 2005), MF has proven considerably less effective when applied to the "mega genomes" of pine and wheat (Rabinowicz et al. 2005).
- CF is rooted in the principles of Cot analysis, the study of DNA reassociation in solution (Peterson et al. 2002). When sheared DNA is heated to 100 °C, the two complementary strands of each double helix come apart in a process known as denaturation. If the denatured DNA is slowly cooled, complementary DNA strands find each other and form double helices (duplexes). The rate at which a particular DNA sequence finds a complementary sequence with which to pair is proportional to the number of times that sequence (and hence its complementary sequence) is found within the genome. In other words, repetitive sequences renature more quickly than low-copy sequences. In CF, sheared genomic DNA is denatured and allowed to reassociate for a period of time in which only repetitive DNA sequences are likely to form duplexes. The double-stranded repetitive DNA is then separated from single-stranded low-copy DNA using hydroxyapatite chromatography. The low-copy DNA and/or the repetitive DNA can then be cloned and sequenced (Peterson et al. 2002; Yuan et al. 2003;

Lamoureux et al. 2005) or used directly as a pyrosequencing substrate. With regard to the latter option, Peterson et al. (2006) recently sequenced Cot-filtered *P. taeda* 7-56 DNA including isolated highly repetitive, moderately repetitive, and low-copy sequences using the new 454 Life Sciences Genome Sequencer 20 (see Margulies et al. 2005 for details on this instrument). These data are currently being evaluated, although initial results look promising. CF has proven highly effective in wheat, where it provides 19.5-fold enrichment for low-copy DNA (Lamoureux et al. 2005).

2.5.2

Genomic Tools to Identify Genes of Economic and Ecological Interest

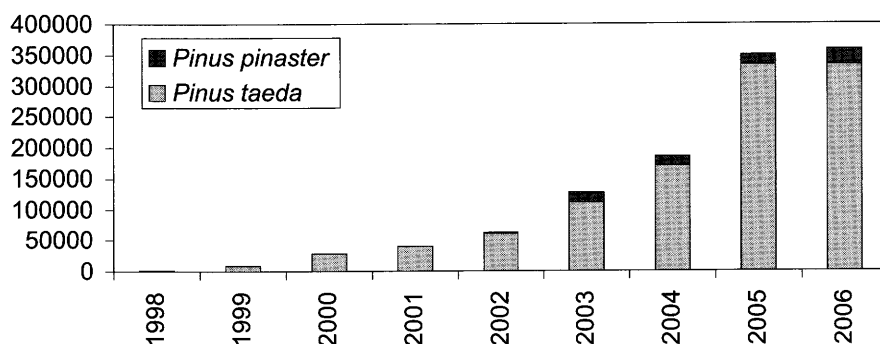
Characterization of the Pine Gene Space

A large-scale expressed sequence tag (EST) sequencing project can generate a partial sequence for a large proportion of genes from a given organism (Adams et al. 1991). As only genic sequences are analyzed regardless of genome size, this approach is considered the most cost-efficient strategy of gene discovery for organisms whose genome sequences are not yet available (Rudd 2003), especially for species, such as pines, with exceedingly large genomes primarily composed of nongenic repetitive elements. For this reason, such projects have been initiated for many organisms including the numerous agronomically important plant species (Rounsley et al. 1998). To rapidly scan for all the protein coding genes and to provide a sequence tag for each gene of the pine genome, large collections of ESTs has been developed in *P. taeda* and, to a lesser extent, in *P. pinaster*.

Loblolly pine ESTs Three major EST sequencing projects, mainly funded by the NSF, have been conducted so far in *P. taeda*. A first project, "Genomics of wood formation in Loblolly pine," was concerned with gene discovery in wood formation (<http://pinetree.ccgb.umn.edu/>). It resulted in ca. 60,000 ESTs placed in the public domain. A second project, entitled "Transcriptome responses to environmental conditions in Loblolly pine roots," aimed at completing this information by targeting especially roots undergoing a variety of biotic and abiotic stresses (<http://funken.org/Projects/Pine/Pine.htm>). It resulted in 140,000 additional ESTs. A third project, "Genomics of Loblolly pine embryogenesis," seeks to identify and characterize

¹ Cot filtration was originally called "Cot-Based Cloning and Sequencing" (CBCS; Peterson et al. 2002) and later "high Cot" (HC; Yuan et al. 2003) sequencing. However, the term "Cot filtration" and the acronym CF are becoming more widely used because (a) the acronym "CBCS" can also stand for "clone-by-clone sequencing," (b) "high Cot" is frequently confused with "high copy," and (c) Cot components may be sequenced without prior cloning (Peterson et al. 2006).

Fig. 5. Pine sequence resources available at EMBL (www.ebi.ac.uk/srs), on 28 February 2006



an additional 85,000 ESTs derived from somatic and zygotic embryos (<http://www.tigr.org/tdb/e2k1/pine/index.shtml>).

Maritime pine ESTs Early molecular biology studies paved the way for the construction of cDNA libraries from different woody tissues (Cantón et al. 1993; García-Gutiérrez et al. 1995), roots (Dubos and Plomion 2003) and buds (C. Collada, M.A. Guevara, and M.T. Cervera, unpublished results) of maritime pine. Funded by the European Union (GEMINI project), INRA (lignome project), and CNS (ForEST project), 28,000 high-quality sequences have been generated in about equal number from wood-forming tissues, roots, and buds. These ESTs have been made available at the EMBL database and at the following URL: <http://cbi.labri.fr/outils/SAM/COMPLETE/>. The *P. pinaster* cDNA clones are available for the scientific community, at the Platform for Integrated Clone Management PICME Web site (<http://www.picme.at/>). A Web-based environment for assembly and annotation of collections of ESTs (Le Provost et al. in prep.) was used to analyze the *P. pinaster* ESTs.

A comprehensive unigene assembled for the user community The number of pine ESTs in public databases has increased dramatically during the past decade (Fig. 5), especially for *P. taeda*, ranking at the 19th position in dbEST with 329,469 ESTs, and to a lesser extent for *P. pinaster* (about 28,000 ESTs). The other pine species, *P. elliottii*, *P. banksiana*, *P. patula*, and *P. sylvestris*, represent less than 2,000 sequences. In 2005, the *Pinus* genus was introduced in the TIGR gene index (Quackenbush et al. 2001), where a comprehensive collection of all publicly available pine ESTs has been clustered into a tentative

unigene set of 45,500 genes (http://www.tigr.org/tigr-scripts/tgi/T_index.cgi?species=pinus), representing probably a major fraction of the coding genome of pine. This resource has laid the foundation for future identification, characterization, and cloning of economically important genes in pines and should provide breeders with a wealth of information to significantly enhance and speed up the breeding process.

Besides this resource, a significant number of ESTs have not been released yet in the public domain and are mainly held by private companies. For instance, Strabala (2004) reported that in August 2001, 344,279 ESTs from *P. radiata* were sequenced by AgriGenesis Biosciences Ltd. based in New Zealand.

Microarrays: New Nools to Study the Functioning of the Pine Genome

ESTs represent snapshots of the genes expressed in a given tissue or at a given developmental stage and provide a good representation of the expressed regions of the genome if a sufficient number of different libraries from a range of tissues and developmental stages and after various environmental challenges are analyzed. To this end, several cDNA libraries have been produced from pine roots, needles, differentiating xylem, and buds (see previous section). These genes have enabled the development of microarrays to identify genes whose expression varies in response to various environmental and developmental cues (mainly drought-stress response and wood formation). These studies, which will likely increase in the future, are listed in Table 6 with others that have used other transcriptomic (SAGE, SSH, cDNA-AFLP) and proteomic (2D PAGE MS/MS) approaches to study the functioning of the pine genome.

Table 6. Transcriptome and proteome analysis in pines

Species	Methodology	Study	Tissue	Authors
<i>P. contorta</i>	Microarrays	Adventitious root development	Roots	Brinker et al. 2004
<i>P. halepensis</i> , <i>P. taeda</i>	cDNA library, Northern blot, RT-PCR	Drought response in two pine species	Roots, needles	Sathyan 2005
<i>P. pinaster</i>	cDNA-AFLP, reverse Northern	Drought response	Roots	Dubos and Plomion 2003
<i>P. pinaster</i>	cDNA-AFLP, reverse Northern	Drought response	Needles	Dubos et al. 2003
<i>P. pinaster</i>	cDNA library	Bud phenology	Buds at different developmental stages	Collada, Guevara, and Cerera, unpublished results
<i>P. pinaster</i>	Six SSH cDNA libraries	Wood formation	Early vs. late, juvenile vs. mature wood-forming tissues.	Pacheco et al. 2005
<i>P. pinaster</i>	cDNA-AFLP; reverse Northern; RT-PCR	Wood formation	Early vs. late, opposite vs. compression wood-forming tissues	Le Provost et al. 2003
<i>P. pinaster</i>	cDNA library	Wood formation	Composite xylem tissues	Cantón et al. 2003
<i>P. pinaster</i>	EST sequencing, microarrays, qPCR	Wood formation	Early vs. late, juvenile vs. mature wood-forming tissues and 8 different organs	Paiva 2006
<i>P. pinaster</i>	EST sequencing, macroarrays, qPCR, 2DE MS/MS	Drought response in two ecotypes	Roots	Chaumeil 2006
<i>P. pinaster</i>	2DE, tandem MS	Wood formation	Wood-forming tissues	Gion et al. 2005
<i>P. pinaster</i>	2DE, micro-sequencing	Drought response	Needles	Costa et al. 1998
<i>P. pinaster</i>	2DE, micro-sequencing	Wood formation	compression and opposite wood forming tissues	Plomion et al. 2000
<i>P. pinea</i>	Two SSH cDNA libraries	Cytokinin induction of adventitious buds	Cotyledons	Alonso, Cantón, Ordás, unpublished results
<i>P. sylvestris</i>	cDNA library	Nitrogen and carbon assimilation	Cotyledons	Avila et al. 2000
<i>P. radiata</i>	CAGE	Wood formation	Wood-forming tissues	Cato et al. 2006a
<i>P. radiata</i>	Three SSH cDNA libraries	Adventitious rooting capacity	Rooting competent tissues	Sánchez et al. 2005
<i>P. taeda</i>	Differential screening, northern blot	Drought response	Needles, stems, roots	Chang et al. 1996
<i>P. taeda</i>	Microarrays	Wood formation	Wood-forming tissues	Yang and Loopstra 2005
<i>P. taeda</i>	Microarrays	Drought response and recovery	Needles	Watkinson et al. 2003
<i>P. taeda</i>	Microarrays	Drought response	Needles, rooted cuttings, 2 unrelated genotypes	Heath et al. 2002

Table 7. (continued)

Species	Methodology	Study	Tissue	Authors
<i>P. taeda</i>	Differential display	Response to galled tissues	Healthy and galled stem	Warren and Covert 2004
<i>P. taeda</i>	0.35k microarrays; qRT-PCR	Wood formation	Unlignified early and latewood xylem: cambial region and developing secondary nonlignified xylem	Egertsdotter et al. 2004
<i>P. taeda</i>	cDNA and subtractive libraries; EST sequencing; Northern blot	Wood formation	Compression and opposite wood	Allona et al. 1998
<i>P. taeda</i>	EST sequencing, microarray	Wood formation	Different tissues and organs, including several types of differentiating xylem samples	Whetten et al. 2001
<i>P. taeda</i>	SAGE profiling	Wood formation	Juvenile and mature wood formatting tissues	Lorenz and Dean 2002
<i>P. taeda</i>	cDNA library; EST sequencing, comparative sequence analysis	Wood formation	Six types of wood forming tissues	Kirst et al. 2003
<i>P. taeda</i>	Microarray, qRT-PCR	Identification of genes preferentially expressed in <i>P. taeda</i>	Differentiating xylem, needles, megagametophytes, embryo (growing radicle)	Yang et al. 2004
<i>P. taeda</i>	Comparative sequence analysis, digital profiling	Computational analysis of transcript accumulation in xylem.	Different types of wood-forming tissues	Pavy et al. 2005
<i>P. taeda</i>	EST sequencing, in silico detection of differentially expressed genes	Drought response	Roots	Lorenz et al. 2005
<i>P. taeda</i> , <i>P. sylvestris</i> , <i>P. abies</i>	Macroarrays	Heterologous arrays	Needles	van Zyl et al. 2002
<i>P. taeda</i>	Microarrays	Disease resistance	Stem (infection court)	Myburg et al. 2006
<i>P. elliotii</i> var. <i>elliottii</i>	Microarrays	Disease resistance	Stem (infection court)	Morse et al. 2004

2.6

Future Perspective: Challenges for Molecular Breeding of Pines

Pines offer a great opportunity for molecular breeding given their large size and long generation interval. The large size causes inefficiencies in progeny testing and concomitant decreases in heritability of traits. Molecular-marker information can reduce these inefficiencies and effectively increase heritability, leading to more accurate selections and commensurate increases in genetic gain (Fernando and Grossman 1989; Lande and Thompson 1990; Hospital et al. 1997). The long generation intervals cause delays in obtaining genetic gain. Savings of many years in the breeding cycle can easily be envisioned if selections can be based on DNA marker information collected at a very early age as opposed to phenotypes measured much later (Williams and Neale 1992). However, these gains can only be achieved if breeding can commence very soon after the selections are made. This has traditionally been known as accelerated breeding, and it is a critical, complementary technology (e.g., Bramlett and Burris 1995).

While the opportunity for increased genetic gain per year is great, major challenges to implement molecular breeding include pines' highly heterozygous, outbreeding nature and the presence of low levels of LD in natural populations (Strauss et al. 1992; Brown et al. 2004). Each of these factors limits the opportunity for finding QTLs by association (Kruglyak 1999; Hirschhorn and Daly 2005) or candidate-gene (Pflieger et al. 2001; Neale and Savolainen 2004) mapping approaches, and for extending QTL results from one pedigree to another within a breeding population. However, given very dense genetic maps or good leads about possible candidate genes, these low LD levels allow for the possibility of discovering tight linkages to important genes. These close linkages should be generally useful across the population for predicting genetic value and implementing molecular breeding.

The challenge in moving forward lies in the development of powerful genetic markers and maps and populations providing genetic information on traits of interest (e.g., Georges et al. 1995; Farnir et al. 2002; Laurie et al. 2004). These populations can include both pedigreed and nonpedigreed populations, and both are valid for the application of molecular breeding. Pedigreed populations place and maintain large re-

gions of the genome in LD, allowing for the tracking of most regions with mapped genetic markers (Lander and Botstein 1989). Once certain regions are identified as causing (or at least being predictive of) significant variation in the phenotype, marker alleles for these regions can be selected for two or more successive generations, thereby increasing the frequency of favorable alleles in the population (Edwards and Page 1994; Hospital et al. 2000). One such view of this for pine breeding is termed marker-directed population improvement (MDPI) (Nelson and Echt 2003, 2004).

Nonpedigreed populations provide an opportunity for tracking very small regions across the genome (Hirschhorn and Daly 2005). These are the regions that have remained in LD for a very long time, possibly since the creation of the DNA polymorphism used as the genetic marker or the gene mutation itself. Extremely dense genetic maps are required for mapping QTLs in nonpedigreed populations through a genome-scan analysis (Kruglyak 1999). Testing specific genes known as candidate genes for association to phenotype does not require a genetic map (Pflieger et al. 2001; Neale and Savolainen 2004). Instead, a hypothesis about what gene(s) might affect a trait's phenotype is needed as well as markers that allow detection of alternative alleles of the candidate gene(s) in the population (Krutovsky and Neale 2005).

The greatest opportunities for implementing molecular breeding exist in breeding programs that utilize best testing practices, including good experimental design, careful field site selection and maintenance, and high-quality measurements on all important traits (e.g., Stuber et al. 1999; Tanksley and Nelson 1996). Most of these programs will contain populations with pedigree structure, allowing for moderately dense genetic maps (10 to 20 cM spacing) to be used in various linkage-mapping approaches (Darvasi et al. 1993) including MDPI. In addition, all trees are drawn from a source population where tight linkages will be maintained over short recombination intervals, allowing for mapping by association through a genome-scan or candidate-gene approach (Wu et al. 2002; Lund et al. 2003).

Where no breeding program exists, large random mating populations could be established specifically for association mapping and subsequent marker-assisted selection and breeding. These would most likely use candidate-gene methods, as candidate genes could be selected from first principles or knowledge of related species and then tested for association with phenotype in the target species and population. It

would seem that genome-scan approaches will only be viable in very intensively studied species where dense genetic maps will be developed. In all cases, highly efficient DNA isolation and genotyping methods (e.g., Darvasi and Soller 1994; Mosig et al. 2001) and effective database management and bioinformatic tools will be required for cost-effective implementation (Nelson 1997). Expected results include increased genetic gain per generation due to increased effective heritability and with accelerated breeding, increasing the number of generations per unit of time. In addition, traits that are difficult to measure can be improved more readily assuming they are measured and mapped in an early generation.

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References

- Adams MD, Kelley JM, Gocayne JD, Dubnick M, Polymeropoulos MH, Xiao H, Merril CR, Wu A, Olde B, Moreno RF, Kerlavage AR, McCombie WR, Venter JC (1991) Complementary DNA sequencing: expressed sequence tags and human genome project. *Science* 252:1651–1656
- Aimers-Halliday J, Shelbourne CJA, Hong SO (1997) Issues in developing clonal forestry with *P. radiata*. In: Burdon RD, Moore JM (eds) "IUFRO '97 Genetics of Radiata Pine", Proc IUFRO Conf 1-4 Dec and Workshop 5 Dec, Rotorua, New Zealand. FRI Bull 203:264–272
- Alazard P (2001) Genetic gain in seeds from first generation seed orchards of maritime pine. *Informations Forests*, Afoel, France, No 628
- Alía R, Gil L, Pardos JA (1995) Performance of 43 *Pinus pinaster* provenances on 5 locations in Central Spain. *Silvae Genet* 44:75–81
- Alía R, Moro J, Denis JB (1997) Performance of *Pinus pinaster* Ait. provenances in Spain: interpretation of the genotype-environment interaction. *Can J For Res* 27:1548–1559
- Allona I, Quinn M, Shoop I, Swope K, ST-Cyr S, Carlis J, Rield J, Retzel E, Campbell M, Sederoff R, Whetten RW (1998) Analysis of xylem formation in pine by cDNA sequencing. *Proc Natl Acad Sci USA* 95:9693–9698
- Andrew RL, Peakall R, Wallis IR, Wood JT, Knight EJ, Foley WJ (2005) Marker-based quantitative genetics in the wild?: The heritability and genetic correlation of chemical defenses in *Eucalyptus*. *Genetics* 171:1989–1998
- Apiolaza LA, Greaves BL (2001) Why are most breeders not using economic breeding objectives. In: IUFRO Symp "Developing the Eucalypt for the Future", 10-14 Sept 2001, Valdivia, Chile
- Arcade A, Faivre-Rampant P, Le Guerroue B, Paques LE, Prat D (1996) Quantitative traits and genetic markers: analysis of a factorial mating design in larch. In: Ahuja MR, Boerjan W, Neale D (eds) *Somatic Cell Genetics and Molecular Genetics of Trees*. Kluwer, Dordrecht, pp 211–216
- Arumuganathan K, Earle ED (1991) Nuclear DNA content of some important plant species. *Plant Mol Biol Rep* 9:208–218
- Atwood RA, White TL, Huber DA (2002) Genetic parameters and gains for growth and wood properties in Florida source loblolly pine in the southeastern United States. *Can J For Res* 32:1025–1038
- Auckland L, Bui T, Zhou Y, Shepherd M, Williams CG (2002) *Conifer Microsatellite Handbook*. Corporate Press, College Station, TX
- Avila C, Muñoz-Chapuli R, Plomion C, Frigerio J-M, Cánovas FM (2000) Two genes encoding distinct cytosolic glutamine synthetases are closely located in the pine genome. *FEBS Lett* 477:237–243
- Bahrman N, Damerval C (1989) Linkage relationships of loci controlling protein amounts in maritime pine (*Pinus pinaster* Ait). *Heredity* 63:267–274
- Ball RD (2001) Bayesian methods for quantitative trait loci mapping based on model selection: approximate analysis using the Bayesian Information Criterion. *Genetics* 159:1351–1364
- Ball RD (2005) Experimental designs for reliable detection of linkage disequilibrium in unstructured random population association studies. *Genetics* 170:859–873
- Barker CL, Donald T, Pauquet J, Ratnaparkhe MB, Bouquet A, Adam-Blondon AE, Thomas MR, Dry I (2005) Genetic and physical mapping of the grapevine powdery mildew resistance gene, *Run1*, using a bacterial artificial chromosome library. *Theor Appl Genet* 111:370–377
- Beaulieu J, Plourde A, Daoust G, Lamontagne L (1996) Genetic variation in juvenile growth of *Pinus strobus* in replicated Quebec provenance-progeny tests. *For Genet* 3:103–112
- Beavis WD (1994) The power and deceit of QTL experiments: lessons from comparative QTL studies. In: Proc 49th Annu Corn and Sorghum Industry Res Conf, ASTA, Washington, DC, pp 250–266
- Bedell JA, Budiman MA, Nunberg A, Citek RW, Robbins D, Jones J, Flick E, Rholing T, Fries J, Bradford K, McMenamy J, Smith M, Holeman H, Roe BA, Wiley G, Korff IE, Rabinowicz PD, Lakey N, McCombie WR, Jeddelloh JA, Martienssen

- RA (2005) Sorghum genome sequencing by methylation filtration. *PLoS Biol* 3:13
- Bell JC, Powell M, Devey ME, Moran GF (2004) DNA profiling, pedigree lineage analysis and monitoring in the Australian breeding program of radiata pine. *Silvae Genet* 53:130–134
- Blada I (1994) Inter-specific hybridization of Swiss Stone Pine *Pinus cembra* L. *Silvae Genet* 43:14–20
- Borzan Z, Papes D (1978) Karyotype analysis in *Pinus*: contribution to the standardization of the karyotype analysis and review of some applied techniques. *Silvae Genet* 27:144–150
- Bowe LM, Coat G, dePamphilis CW (2000) Phylogeny of seed plants based on all three genomic compartments: Extant gymnosperms are monophyletic and Gnetales' closest relatives are conifers. *Proc Natl Acad Sci USA* 97:4092–4097
- Bradshaw HD, Foster GS (1992) Marker-aided selection and propagation systems in trees: advantages of cloning for studying quantitative inheritance. *Can J For Res* 22:1044–1049
- Bramlett DL, Burris LC (1995) Topworking young scions into reproductively-mature loblolly pine. In: Weir RJ, Hatcher AV (eds) *Proc 23rd South For Tree Improv Conf*, Asheville, NC, pp 234–241
- Brendel O, Pot D, Plomion C, Rozenberg P, Guehl JM (2002) Genetic parameters and QTL analysis of $\delta^{13}C$ and ring width in maritime pine. *Plant Cell Environ* 25:945–953
- Brettschneider R (1998) RFLP analysis. In: Karp A, Isaac PG, Ingram DS (eds) *Molecular Tools for Screening Biodiversity Plants and Animals*. Chapman & Hall, London, pp 83–95
- Brinker M, van Zyl L, Liu WB, Craig D, Sederoff RR, Clapham DH, von Arnold S (2004) Microarray analyses of gene expression during adventitious root development in *Pinus contorta*. *Plant Physiol* 135(3):1526–1539
- Brown GR, Kadel III EE, Bassoni DL, Kiehne KL, Temesgen B, van Buijtenen JP, Sewell MM, Marshall KA, Neale DB (2001) Anchored reference loci in loblolly pine (*Pinus taeda* L.) for integrating pine genomics. *Genetics* 159:799–809
- Brown GR, Bassoni DL, Gill GP, Fontana JR, Wheeler NC, Megraw RA, Davis MF, Sewell MM, Tuskan GA, Neale DB (2003) Identification of quantitative trait loci influencing wood property traits in loblolly pine. III. QTL verification and candidate gene mapping. *Genetics* 164:1537–1546
- Brown GR, Gill GP, Kuntz RJ, Langley CH, Neale DB (2004) Nucleotide diversity and linkage disequilibrium in loblolly pine. *Proc Natl Acad Sci USA* 101:15255–15260
- Burban C, Petit RJ (2003) Phylogeography of maritime pine inferred with organelle markers having contrasted inheritance. *Mol Ecol* 12:1487–1495
- Burdon RD (1997) Genetic diversity for the future: conservation or creation and capture? In: Burdon RD, Moore JM (eds) "IUFRO '97 Genetics of Radiata Pine", *Proc IUFRO Conf* 1–4 Dec and workshop 5 Dec, Rotorua, New Zealand. *FRI Bull* 203:263–274
- Burdon RD (2001) Genetic diversity and disease resistance: some considerations for research, breeding and deployment. *Can J For Res* 32:596–606
- Burdon RD (2002) An introduction to pines. In: *Pines of Silvicultural Importance*. CABI, Wallingford, UK pp x–xxi
- Burdon RD (2004) Breeding goals: Issues of goal-setting and applications. In: Walter C, Carson M (eds) *Plantation Forest Biotechnology for the 21st Century*, Research Signpost, Kerala, India, pp 101–118
- Burdon RD, Bannister MH, Low CB (1992) Genetic Survey of *Pinus radiata* 4: variance structures and heritabilities in juvenile clones. *NZ J For Sci* 22:187–210
- Burdon RD, Firth A, Low CB, Miller MA (1998) Multi-site provenance trials of *Pinus radiata* in New Zealand. In: *Forest Genetic Resources* 26:3–8, FAO, Rome
- Burdon RD, Gea LD (2006) Pursuit of genetic gain and biotic risk management for *Pinus radiata* in New Zealand. In: *Proc 13th Australasian Plant Breed Conf*, Christchurch, New Zealand pp 75–85, <http://www.apbc.org.nz>
- Burdon RD, Kumar S (2003) Stochastic modelling of the impact of four generations of pollen contamination in unpedigreed gene resources. *Silvae Genet* 52:1–7
- Burdon RD, Kumar S (2004) Forwards versus backwards selection: trade-offs between expected genetic gain and risk avoidance. *NZ J For Sci* 34:3–21
- Burdon RD, Low CB (1992) Genetic survey of *Pinus radiata*. 6: Wood properties: variation, heritabilities, and interrelationship with other traits. *NZ J For Sci* 22:228–245
- Burdon RD, Miller JT (1992) Introduced forest trees of New Zealand: Recognition, role and seed source. 12. Radiata pine (*Pinus radiata* D. Don). *NZ For Res Inst Bull* 124/12 pp 59
- Byun KO, Kim MZS, Shim SY, Hong SH, Sohn SI (1989) Review of pitch- loblolly hybrid pine (*Pinus rigida* and *P. taeda*) breeding researches in Korea and future strategy. *Res Rep, For Genet Res Inst, Korea* 25:204–211
- Cahalan CM (1981) Provenance and clonal variation in growth, branching and phenology in *Picea sitchensis* and *Pinus contorta*. *Silvae Genet* 30:40–46
- Cai Q, Zhang D, Liu ZL, Wang XR (2006) Chromosomal localization of 5S and 18S rDNA in five species of subgenus *Strobus* and their implications for genome evolution of *Pinus*. *Ann Bot* 97:715–722
- Cánovas FM, Dumas-Gaudot E, Recorbet G, Jorrin J, Hans-Peter Mock HP, Rossignol M (2004) Plant proteome analysis. *Proteomics* 4:285–298
- Cantón FR, García-Gutiérrez A, Gallardo F, de Vicente A, Cánovas FM (1993) Molecular characterization of a cDNA clone encoding glutamine synthetase from a gymnosperm: *Pinus sylvestris*. *Plant Mol Biol* 22:819–828
- Cantón FR, Le Provost G, García V, Barré A, Frigerio J-M, Paiva J, Fevereiro P, Avila C, Mouret J-F, de Daruvar A, Cánovas FM, Plomion C (2003) Transcriptome analysis of wood formation in maritime pine In: Espinel S, Barredo Y, Ritter E (eds) *Sustainable Forestry, Wood Products and Biotechnology*. DFA-AFA Press, Vitoria-Gasteiz, Spain, pp 333–347
- Cato S, Mc Millan L, Donaldson L, Richardson T, Echt C, Gardner R (2006a) Wood formation from the base to the crown in

- Pinus Radiata*: gradients of tracheid wall thickness, wood density, radial growth rate and gene expression. *Plant Mol Biol* 60:565–581
- Cato SA, Pot D, Kumar S, Douglas J, Gardner RC, Wilcox PL (2006b) Balancing selection in a dehydrin gene associated with increased wood density and decreased radial growth in *Pinus radiata* (Abstr). In: *Plant & Animal Genome XIV Conf*, San Diego
- Cato SA, Richardson TE (1996) Inter- and intra-specific polymorphism at chloroplast SSR loci and the inheritance of plastids in *Pinus radiata* D. Don. *Theor Appl Genet* 93:587–592
- Cervera MT, Plomion C, Malpica CA (2000a) Molecular markers and genome mapping in woody plants. In: Jain SM, Minocha SC (eds) *Molecular Biology of Woody Plants*. Kluwer, Dordrecht, pp 375–394
- Cervera MT, Remington D, Frigerio JM, Storme V, Ivens B, Boerjan W, Plomion C (2000b) Improved AFLP analysis of tree species. *Can J For Res* 30:1608–1616
- Chagné D, Brown G, Lalanne C, Madur D, Pot D, Neale DB, Plomion C (2003) Comparative genome and QTL mapping between maritime and loblolly pines. *Mol Breed* 12:185–195
- Chagné D, Chaumeil P, Ramboer A, Collada C, Guevara A, Cervera M-T, Vendramin GG, Garcia V, Frigerio JM, Echt C, Richardson T, Plomion C (2004) Cross species transferability and mapping of genomic and cDNA SSRs in pines. *Theor Appl Genet* 109:1204–12
- Chagné D, Lalanne C, Madur D, Kumar S, Frigerio JM, Krier C, Decroocq S, Savouré A, Bou-Dagher-Kharrat M, Bertocchi E, Brach J, Plomion C (2002) A high density genetic map of Maritime pine based on AFLPs. *Ann For Sci* 59:627–636
- Chambers PGS, Borralho NMG (1999) A simple model to examine the impact of changes in wood traits on the cost of thermomechanical pulping and high-brightness newsprint production with radiata pine. *Can J For Res* 29:1615–1626
- Chang S, Puryear JD, Dias MADL, Funkhouser EA, Newton RJ, Cairney J (1996) Gene expression under water deficit in loblolly pine (*Pinus taeda*): isolation and characterization of cDNA clones. *Physiol Planta* 97:139–148
- Chaumeil P (2006) Plasticité moléculaire de deux écotypes de pin maritime soumis à un stress osmotique. Thèse de Doctorat. Université Henri Poincaré, Nancy I, Nancy
- Ciavatta VT, Morillon R, Pullman GS, Chrispeels MJ, Cairney J (2001) An aquaglyceroporin is abundantly expressed early in the development of the suspensor and the embryo proper of loblolly pine. *Plant Physiol* 127:1556–1567
- Claros MG, Cantón FR, Cánovas FM (2004) Construction of a pine BAC genomic library and bioinformatics platform for automated high-throughput sequence analysis (Abstr). 2nd Annual Meeting of the Spanish Forest Functional Genomics Network. Pontevedra, Spain, 29 Nov 2004. http://www.difo.uah.es/forestgenomics/pontevedra_04/Gonzalo%20CLAROS.pdf
- Colbert T, Till BJ, Tompa R, Reynolds S, Steine MN, Yeung AT, McCallum CM, Comai L, Henikoff S (2001) High-throughput screening for induced point mutations. *Plant Physiol* 126:480–484
- Conkle MT (1981) Isozyme variation and linkage in six conifer species. In: *Proc Isozymes of North American Forest Trees and Insects*, USD Forest Service, General Technical Report PSW-48, pp 11–17
- Costa P (1999) Réponse moléculaire, physiologique et génétique du pin maritime à une contrainte hydrique. PhD thesis. Univ of Nancy
- Costa P, Plomion C (1999) Genetic analysis of needle protein in Maritime pine. 2. Quantitative variation of protein accumulation. *Silvae Genet* 48:146–150
- Costa P, Pot D, Dubos C, Frigerio JM, Pionneau C, Bodenes C, Bertocchi E, Cervera MT, Remington DL, Plomion C (2000) A genetic map of Maritime pine based on AFLP, RAPD and protein markers. *Theor Appl Genet* 100:39–48
- Costa P, Bahrman N, Frigerio J-M, Kremer A, Plomion C (1998) Water-deficit-responsive proteins in maritime pine. *Plant Mol Biol* 38:587–596
- Cornelius J (1994) Heritabilities and additive genetic coefficients of variation in forest trees. *Can J For Res* 24:372–379
- Danjon F (1995) Observed selection effects on height growth, diameter and stem form in maritime pine. *Silvae Genet* 44(1):10–19
- Darvasi A, Soller M (1994) Selective DNA pooling for determination of linkage between a molecular marker and a quantitative trait locus. *Genetics* 138:1365–1373
- Darvasi A, Weinreb A, Minke V, Weller JI, Soller M (1993) Detecting marker-QTL linkage and estimating QTL gene effect and map location using a saturated genetic map. *Genetics* 134:943–951
- Devey ME, Fiddler TA, Liu BH, Knapp SJ, Neale DB (1994) An RFLP linkage map for loblolly pine based on a three-generation outbred pedigree. *Theor Appl Genet* 88:273–278
- Devey ME, Fino-Mix A, Kinloch BB Jr, Neale DB (1995) Random amplified polymorphic DNA markers tightly linked to a gene for resistance to white pine blister rust in sugar pine. *Proc Natl Acad Sci USA* 92:2066–2070
- Devey ME, Bell JC, Smith DN, Neale DB, Moran GF (1996) A genetic linkage map for *Pinus radiata* based on RFLP, RAPD, and microsatellite markers. *Theor Appl Genet* 6:673–679
- Devey ME, Sewell MM, Uren TL, Neale DB (1999) Comparative mapping in loblolly and radiata pine using RFLP and microsatellite markers. *Theor Appl Genet* 99:656–662
- Devey ME, Bell JC, Uren TL, Moran GF (2003) A set of microsatellite markers for fingerprinting and breeding applications in *Pinus radiata*. *Genome* 45:984–989
- Devey ME, Carson SD, Nolan MF, Matheson AC, Te Riini C, Hohepa J (2004a) QTL associations for density and diameter in *Pinus radiata* and the potential for marker-aided selection. *Theor Appl Genet* 108:516–524

- Devey ME, Groom KA, Nolan MF, Bell JC, Dudzinski MJ, Old KM, Matheson AC, Moran GF (2004b) Detection and verification of quantitative trait loci for resistance to Dothistroma needle blight in *Pinus radiata*. *Theor Appl Genet* 108:1056–1063
- Dhakal LP, White TL, Hodge GR (1996) Realized genetic gains from slash pine (*Pinus elliottii*) tree improvement. *Silvae Genet* 45:190–197
- Dilbirli M, Erayman M, Sandhu D, Sidhu D, Gill KS (2004) Identification of wheat chromosomal regions containing expressed resistance genes. *Genetics* 166:461–481
- Doudrick RL, Heslop-Harrison JS, Nelson CD, Schmidt T, Nance WL, Schwarzbacher T (1995) Karyotype of slash pine (*Pinus elliottii* var *elliottii*) using patterns of fluorescence *in situ* hybridization and fluorochrome banding. *J Hered* 86:289–296
- Drewry A (1982) G-banded chromosomes in *Pinus resinosa*. *J Hered* 73:305–306
- Du Toit B, Smith C, Carlson C, Esprey L, Allen R, Little K (1998) Eucalypt and pine plantations in South Africa. In: Nambiar EKS, Cossalter C, Triarks A (eds) Workshop Proceedings. CIFOR, 16–20 Feb 1998, South Africa, pp 23–30
- Dubos C, Plomion C (2003) Identification of water-deficit responsive genes in maritime pine (*Pinus pinaster* Ait.) roots. *Plant Mol Biol* 51(2):249–262
- Dubos C, Provost G, Pot D, Salin F, Lalane C, Madur D, Frigerio JM, Plomion C (2003) Identification and characterization of water-stress-responsive genes in hydroponically grown maritime pine (*Pinus pinaster*) seedlings. *Tree Physiol* 23:169–179
- Dvornyk V, Sirviö A, Mikkonen M, Savolainen O (2002) Low nucleotide diversity at the *pal1* locus in the widely distributed *Pinus sylvestris*. *Mol Biol Evol* 19:179–188
- Echt CS, DeVerno LL, Anzidei M, Vendramin GG (1998) Chloroplast microsatellites reveal population genetic diversity in red pine, *Pinus resinosa* Ait. *Mol Ecol* 7:307–317
- Echt CS, Burns R (1999) PCR primer pair sequences and SSRs for various *P. taeda* ESTs. Available on the dendrome Web site at http://dendrome.ucdavis.edu/Gen_res.htm, accessed 20 March 2006
- Echt CS, Nelson CD (1997) Linkage mapping and genome length in eastern white pine (*Pinus strobus* L.). *Theor Appl Genet* 94:1031–1037
- Echt CS, Saha S, Deemer D, Nelson CD (2006) Evaluation of pine EST-SSR markers. In: Plant & Animal Genome XIV Conf, San Diego, W-117 (http://www.intl-pag.org/14/abstracts/PAG14_W117.html, accessed 20 March 2006)
- Edwards MD, Page NJ (1994) Evaluation of marker-assisted selection through computer simulation. *Theor Appl Genet* 88:376–382
- Edwards MD, Stuber CW, Wendel JF (1987) Molecular-marker-facilitated investigations of quantitative-trait loci in maize. I. Numbers, genomic distribution and types of gene action. *Genetics* 116:113–125
- Egertsdotter U, van Zyl LM, Mackay J, Peter G, Kirst M, Clark C, Whetten R, Sederoff R (2004) Gene expression during formation of earlywood and latewood in loblolly pine: expression profiles of 350 genes. *Plant Biol* 6:654–663
- Elsik CG, Williams CG (2000) Retroelements contribute to the excess low-copy-number DNA in pine. *Mol Gen Genet* 264:47–55
- Elsik CG, Williams CG (2001) Low copy microsatellite recovery from a conifer genome. *Theor Appl Genet* 103:1189–1195
- Emebiri LC, Devey ME, Matheson AC, Slee MU (1997) Linkage of RAPD markers to NESTUR, a stem growth index in radiata pine seedlings. *Theor Appl Genet* 95:119–124
- Emebiri LC, Devey ME, Matheson AC, Slee MU (1998) Age-related changes in the expression of QTLs for growth in radiata pine seedlings. *Theor Appl Genet* 97:1053–1061
- Epperson BK, Allard RW (1989) Spatial autocorrelation analysis of the distribution of genotypes within population of lodgepole pine. *Genetics* 121:369–377
- Falkenhagen E (1991) Provenance variation in *Pinus radiata* at six sites in South Africa. *Silvae Genet* 40:41–50
- Farjon A (1984) Pines. Drawings and description of the genus *Pinus*. Brill, Leiden
- Farjon A (2001) World checklist and bibliography of conifers, 2nd edn. The Royal Botanic Gardens, Kew, UK
- Farjon A, Styles BT (1997) *Pinus* (Pinaceae). Monograph 75, Flora Neotropica. New York Botanical Gardens, New York
- Farnir F, Grisart B, Coppieters W, Riquet J, Berzi P, Cambisano N, Karim L, Mni M, Moisis S, Simon P, Wagenaar D, Vilkkilä J, Georges M (2002) Simultaneous mining of linkage and linkage disequilibrium to fine map quantitative trait loci in outbred half-sib pedigrees: revisiting the location of a quantitative trait locus with major effect on milk production on bovine chromosome 14. *Genetics* 161:275–287
- Fernando RL, Grossman M (1989) Marker-assisted selection using best linear unbiased prediction. *Genet Sel Evol* 21:246–277
- Fisher PJ, Gardner RC, Richardson TE (1996) Single locus microsatellites isolated using 5' anchored PCR. *Nucleic Acids Res* 24:4369–4371
- Fisher PJ, Gardner RC, Richardson TE (1998) Characteristics of single and multi-copy microsatellites from *Pinus radiata*. *Theor Appl Genet* 96:969–979
- Flint-Garcia SA, Thornsberry JM, Buckler ES (2003) Structure of linkage disequilibrium in plants. *Annu Rev Plant Biol* 54:357–374
- Folkertsma RT, Spassova MI, Prins M, Stevens MR, Hille J, Goldbach RW (1999) Construction of a bacterial artificial chromosome (BAC) library of *Lycopersicon esculentum* cv. Stevens and its application to physically map the *Sw-5* locus. *Mol Breed* 5:197–207
- Friesen N, Brandes A, Heslop-Harrison JS (2001) Diversity, origin, and distribution of retrotransposons (gypsy and copia) in conifers. *Mol Biol Evol* 18:1176–1188
- Fulton TM, Van der Hoeven R, Eannetta NT, Tanksley SD (2002) Identification, analysis, and utilization of conserved or-

- tholog set markers for comparative genomics in higher plants. *Plant Cell* 14:1457–1467
- García-Gil MR, Mikkonen M, Savolainen O (2003) Nucleotide diversity at two phytochrome loci along a latitudinal cline in *Pinus sylvestris*. *Mol Ecol* 12:1195–1206
- García-Gutiérrez A, Cantón FR, Gallardo F, Sánchez-Jiménez F, Cánovas FM (1995) Expression of ferredoxin-dependent glutamate synthase in dark-grown pine seedlings. *Plant Mol Biol* 27:115–128
- Gaussen H (1960) Les Gymnospermes actuelles et fossils. Fascicule VI. Généralités, genre *Pinus*; Faculté des Sciences, Toulouse
- Geada López G, Kamiya K, Harada K (2002) Phylogenetic relationships of *Diploxylon* pines (subgenus *Pinus*) based on plastid sequence data. *Int J Plant Sci* 163:737–747
- Geldermann H (1975) Investigations on inheritance of quantitative characters in animals by gene markers. I. Methods. *Theor Appl Genet* 46:319–330
- Georges M, Nielsen D, Mackinnon M, Mishra A, Okimoto R, Pasquino AT, Sargeant LS, Sorensen A, Steele MR, Zhao X, Womack JE, Hoeschele I (1995) Mapping quantitative trait loci controlling milk production in dairy cattle by exploiting progeny testing. *Genetics* 139:907–920
- Gerber S, Rodolphe F (1994) An estimation of the genome length of maritime pine (*Pinus pinaster* Ait.). *Theor Appl Genet* 88:289–292
- Gerber S, Rodolphe F, Bahrman N, Baradat P (1993) Seed-protein variation in maritime pine (*Pinus pinaster* Ait.) revealed by two-dimensional electrophoresis: genetic determinism and construction of a linkage map. *Theor Appl Genet* 85:521–528
- Gernandt DS, Geada Lopez G, Ortiz Garcia S, Liston A (2005) Phylogeny and classification of *Pinus*. *Taxon* 54:29–42
- Gernandt DS, Liston A, Pinero D (2003) Phylogenetics of *Pinus* subsections *Cembroides* and *Nelsoniae* inferred from cpDNA sequences. *Syst Bot* 28:657–673
- Gion JM, Lalanne C, Le Provost G, Ferry-Dumazet H, Paiva J, Chaumeil P, Frigerio JM, Brach J, Barré A, de Daruvar A, Claverol S, Bonneau M, Sommerer N, Negroni L, Plomion C, Gion J-M, Lalanne C, et al. (2005) The proteome of maritime pine wood forming tissue. *Proteomics* 5:3731–3751
- Gómez MA, González-Martínez SC, Collada C, Climent J, Gil L (2003) Complex population genetic structure in the endemic Canary Island pine revealed using chloroplast microsatellite markers. *Theor Appl Genet* 107:1123–1131
- González-Martínez SC, Gerber S, Cervera MT, Martínez-Zapater J, Gil L, Alía R (2002) Seed gene flow and fine-scale structure in a Mediterranean pine (*Pinus pinaster* Ait.) using nuclear microsatellite markers. *Theor Appl Genet* 104:1290–1297
- González-Martínez SC, Robledo-Arnuncio JJ, Collada C, Díaz A, Williams CG, Alía R, Cervera MT (2004) Cross-amplification and sequence variation of microsatellite loci in Eurasian hard pines. *Theor Appl Genet* 109:103–111
- González-Martínez SC, Ersoz E, Brown GR, Wheeler NC, Neale DB (2006a) DNA sequence variation and selection of tag SNPs at candidate genes for drought-stress response in *Pinus taeda*. *Genetics* 172:1915–1926
- González-Martínez SC, Krutovsky KV, Neale DB (2006b) Forest-tree population genomics and adaptive evolution. *New Phytol* 170:227–238
- Govindaraju D, Lewis P, Cullis C (1992) Phylogenetic analysis of pines using ribosomal DNA restriction fragment length polymorphisms. *Plant Syst Evol* 179:141–153
- Grace SL, Hamrick JL, Platt WJ (2004) Estimation of seed dispersal in an old-growth population of Longleaf Pine (*Pinus palustris*) using maternity exclusion analysis. *Castanea* 69:207–215
- Grattapaglia D, Sederoff R (1994) Genetic linkage maps of *Eucalyptus grandis* and *Eucalyptus urophylla* using a pseudotestcross mapping strategy and RAPD markers. *Genetics* 137:1121–1137
- Greaves BL (1999) The value of tree improvement: A case study in radiata pine grown for structural sawn timber and liner-board. In: Nepveu G (ed) Connection between silviculture and wood quality through modelling approaches and simulation software. Proc IUFRO Workshop, 5–12 Dec 1999, France, pp 448–459
- Groover AT, Devey ME, Lee JM, Megraw R, Mitchell-Olds T, Sherman B, Vujcic S, Williams C, Neale DB (1994) Identification of quantitative trait loci influencing wood specific gravity in an outbred pedigree of loblolly pine. *Genetics* 138:1293–1300
- Groover AT, Williams CG, Devey ME, Lee JM, Neale DB (1995) Sex-related differences in meiotic recombination frequency in *Pinus taeda*. *J Hered* 86:157–158
- Grotkopp E, Rejmanek M, Sanderson MJ, Rost TL (2004) Evolution of genome size in pines (*Pinus*) and its life-history correlates: supertree analyses. *Evolution* 58:1705–1729
- Guevara MA, Chagné D, Almeida MH, Byrne M, Collada C, Favre JM, Harvengt L, Jeandroz S, Orazio C, Plomion C, Ramboer A, Rocheta M, Sebastiani F, Soto A, Vendramin GG, Cervera MT (2005a) Isolation and characterization of nuclear microsatellite loci in *P. pinaster*. *Mol Ecol Notes* 5:57–59
- Guevara MA, Soto A, Collada C, Plomion C, Savolainen O, Neale DB, González-Martínez SC, Cervera MT (2005b) Genomics applied to the study of adaptation in pine species. *Invest Agrar Syst Recur For* 14:292–306
- Gugerli F, Senn J, Anzidei M, Madaghiele A, Buchler U, Sperisen C, Vendramin GG (2001) Chloroplast microsatellites and mitochondrial *nad1* intron 2 sequences indicate congruent phylogenetic relationships of Swiss stone pine (*Pinus cembra*), Siberian stone pine (*P. sibirica*) and Siberian dwarf pine (*P. pumila*). *Mol Ecol* 10:1489–1497
- Gupta PK, Rustgi S, Kulwal PL (2005) Linkage disequilibrium and association studies in higher plants: Present status and future prospects. *Plant Mol Biol* 57:461–485

- Gwaze DP, Harding KJ, Purnell RC, Bridgwater FE (2002) Optimum selection age for wood density in loblolly Pine. *Can J For Res* 32:1393–1399
- Gwaze DP, Zhou Y, Reyes-Valdés MH, Al-Rababah MA, Williams CG (2003) Haplotypic QTL mapping in an outbred pedigree. *Genet Res* 81:43–50
- Hamrick JL, Godt JW (1990) Allozyme diversity in plant species. In: Brown A, Clegg MT, Kahler AL, Weir BS (eds) *Plant Population Genetics, Breeding and Genetic Resources*. Sinauer, Sunderland, MA, pp 43–63
- Hamrick JL, Godt MJW (1996) Effects of life history traits on genetic diversity in plant species. *Phil Trans R Soc Lond B Biol Sci* 351:1291–1298
- Hannrup B, Ekberg I (1998) Age-age correlations for tracheid length and wood density in *Pinus sylvestris*. *Can J For Res* 28:1373–1379
- Hannrup B, Ekberg I, Persson A (2000) Genetic correlations among wood, growth capacity and stem traits in *Pinus sylvestris*. *Can J For Res* 15:161–170
- Harkins DM, Johnson GN, Skaggs PA, Mix AD, Dupper GE, Devvey ME, Kinloch Jr BB, Neale DB (1998) Saturation mapping of a major gene for resistance to white pine blister rust in sugar pine. *Theor Appl Genet* 97:1355–1360
- Harry DE, Temesgen B, Neale DB (1998) Codominant PCR-based markers for *Pinus taeda* developed from mapped cDNA clones. *Theor Appl Genet* 97:327–336
- Hartmann HT, Kester DE, Davies FT Jr (1990) *Plant Propagation Principles and Practices*, 5th edn. Prentice Hall, Englewood Cliffs, NJ
- Hayashi E, Kondo T, Terada K, Kuramoto N, Goto Y, Okamura M, Kawasaki H (2001) Linkage map of Japanese black pine based on AFLP and RAPD markers including markers linked to resistance against the pine needle gall midge. *Theor Appl Genet* 102:871–875
- Hayashi E, Kondo T, Terada K, Kuramoto N, Kawasaki S (2004) Identification of AFLP markers linked to a resistance gene against pine needle gall midge in Japanese black pine. *Theor Appl Genet* 108:1177–1181
- Heath LS, Ramakrishnan N, Sederoff RR, Whetten RW, Chevone BI, Struble CA, Jouenne VY, Chen D, van Zyl LM, Grene R (2002) Studying the functional genomics of stress responses in loblolly pine with the “Expresso microarray experiment management system”. *Comp Funct Genom* 3:226–243
- Hirschhorn JN, Daly MJ (2005) Genome-wide association studies for common diseases and complex traits. *Nat Rev Genet* 6:95–108
- Hizume M, Arai M, Tanaka A (1990) Chromosome banding in the genus *Pinus*. III. Fluorescent banding pattern of *P. luchuensis* and its relationships among the Japanese diploxylon pines. *Bot Mag Tokyo* 103:103–111
- Hizume M, Ohgiku A, Tanaka A (1989) Chromosome banding in the genus *Pinus*. II. Interspecific variation of fluorescent banding patterns in *P. densiflora* and *P. thunbergii*. *Bot Mag Tokyo* 102:25–36
- Hizume M, Shibata F, Matsusaki Y, Garajova Z (2002) Chromosome identification and comparative karyotypic analyses of four *Pinus* species. *Theor Appl Genet* 105:491–497
- Hodge GR, Dvorak WS (1999) Genetic parameters and provenance variation of *Pinus tecunumanii* in 78 international trials. *For Genet* 6:157–180
- Hodge GR, Volker PW, Potts BM, Owen JV (1996) A comparison of genetic information from open-pollinated and control-pollinated progeny tests in two Eucalypt species. *Theor Appl Genet* 92:53–63
- Hospital F, Goldringer I, Openshaw S (2000) Efficient marker-based recurrent selection for multiple quantitative trait loci. *Genet Res* 75:357–368
- Hospital F, Moreau L, Lacoudre F, Charcosset A, Gallais A (1997) More on the efficiency of marker-assisted selection. *Theor Appl Genet* 95:1181–1189
- Hurme P, Sillanpää M, Repo T, Arjas E, Savolainen O (2000) Genetic basis of climatic adaptation in Scots pine by Bayesian QTL analysis. *Genetics* 156:1309–1322
- Hyun SK (1976) Inter-specific hybridization in pines with the special reference to *Pinus rigida* and *P. taeda*. *Silvae Genet* 25:188–191
- Ingvarsson PK (2005) Nucleotide polymorphism and linkage disequilibrium within and among natural populations of European Aspen (*Populus tremula* L., Salicaceae). *Genetics* 169:945–953
- Isik F, Li B, Frampton J, Goldfarb B (2004) Efficiency of seedlings and rooted cuttings for testing and selection in *Pinus taeda*. *For Sci* 50:44–53
- Islam-Faridi MN, Chang Y-L, Zhang HB, Kinlaw C, Doudrick RL, Neale DB, Echt CS, Price HJ, Stelly DM (1998) Construction of a pine BAC library. In: *Plant & Animal Genome VI Conf*, San Diego
- Islam-Faridi MN, Mujeeb-Kazi A (1995) Visualization of *Secale cereale* DNA in wheat germplasm by FISH. *Theor Appl Genet* 90:595–600
- Islam-Faridi MN, Nelson CD, Kubisiak TL (2007) Reference karyotype and cytomolecular map for loblolly pine (*Pinus taeda* L.). *Genome* (in press)
- Islam-Faridi MN, Nelson CD, Kubisiak TL, Gullirmo MV, McNamara VH, Price HJ, Stelly DM (2003) Loblolly pine karyotype using FISH and DAPI positive banding. In: *Proc 27th Southern For Tree Improv Conf*, 25–27 June 2003, Stillwater, OK, pp 184–188
- Ivkovic M, Kumar S, Wu H (2006) Adding value to the end-products of radiata pine: A review of breeding for structural timber production. In: *Proc 13th Australasian Plant Breed Conf*, 18–22 April 2006, Christchurch, NZ, pp 273–278
- Jackson RB, Schlesinger WH (2004) Curbing the U.S. carbon deficit. *Proc Natl Acad Sci USA* 101:15827–15829
- Jacobs MD, Gardner RC, Murray BG (2000) Cytological characterization of heterochromatin and rDNA in *Pinus radiata* and *P. taeda*. *Plant Syst Evo* 1 223:71–79
- Jansen RC (1993) Interval mapping of multiple quantitative trait loci. *Genetics* 135:205–211

- Jewell DC, Islam-Faridi MN (1994) Details of a technique for somatic chromosome preparation and C-banding of maize. In: Freeling M, Walbot V (eds) *The Maize Hand Book*. Springer, Berlin Heidelberg New York, pp 484–493
- Jiang C, Zeng ZB (1995) Multiple trait analysis of genetic mapping for quantitative trait loci. *Genetics* 140:1111–1127
- Jiang JM, Liu ZX, Lu BS, Jiang JM, Liu ZX, Lu BS (1999) Provenance genetic variation analysis of loblolly pine and determination of suitable provenances (areas). *For Res Beijing* 12:485–492
- Johnson GR, Wheeler NC, Strauss SH (2000) Financial feasibility of marker-aided selection in Douglas-fir. *Can J For Res* 30:1942–1952
- Kamm A, Doudrick RL, Heslop-Harrison JS, Schmidt T (1996) The genomic and physical organization of *Ty1*-copied-like sequences as a component of large genomes in *Pinus elliottii* var. *elliottii* and other gymnosperms. *Proc Natl Acad Sci USA* 93:2708–2713
- Kao C-H, Zeng J, Teasdale RD (1999) Multiple interval mapping for quantitative trait loci. *Genetics* 152:1203–1216
- Karhu A, Hurme P, Karjalainen M, Karvonen P, Kärkkäinen K, Neal DB, Savolainen O (1996) Do molecular markers reflect patterns of differentiation in adaptive traits of conifers? *Theor Appl Genet* 93:215–221
- Kärkkäinen K, Savolainen O (1993) The degree of early inbreeding depression determines the selfing rate at the seed stage: model and results from *Pinus sylvestris* (Scots pine). *Heredity* 71:160–166
- Kaya Z, Neale DB (1995) Utility of random amplified polymorphic DNA (RAPD) markers for linkage mapping in Turkish red pine (*Pinus brutia* Ten). *Silvae Genet* 44:110–116
- Kaya Z, Sewell MM, Neale DB (1999) Identification of quantitative trait loci influencing annual height- and diameter-increment growth in loblolly pine (*Pinus taeda* L.). *Theor Appl Genet* 98:586–592
- Kayihon GC, Huber DA, Morse AM, White TL, Davis JM (2005) Genetic dissection of fusiform rust and pitch canker disease traits in loblolly pine. *Theor Appl Genet* 110:948–958
- Kent J, Richardson TE (1997) Fluorescently labelled, multiplexed chloroplast microsatellites for high-throughput paternity analysis in *Pinus radiata*. *NZ J For Sci* 27:305–312
- Khoshoo TN (1961) Chromosome numbers in gymnosperms. *Silvae Genet* 10:1–9
- Kinlaw CS, Neale DB (1997) Complex gene families in pine genomes. *Trends Plant Sci* 2:356–359
- Kinloch BB, Parks GK, Flower CW (1970) White pine blister rust: Simply inherited resistance in sugar pine. *Science* 167:193–195
- Kirst M, Johnson AF, Baucom C, Ulrich E, Hubbard K, Staggs R, Paule C, Retzel E, Whetten R, Sederoff R (2003) Apparent homology of expressed genes from wood forming tissues of loblolly pine (*Pinus taeda* L.) with *Arabidopsis thaliana*. *Proc Natl Acad Sci USA* 100:7383–7388
- Kirst M, Myburg AA, De Leon JP, Kirst ME, Scott J, Sederoff R (2004) Coordinated genetic regulation of growth and lignin revealed by quantitative trait locus analysis of cDNA microarray data in an interspecific backcross of *Eucalyptus*. *Plant Physiol* 135:2368–2378
- Kirst M, Cordeiro CM, Rezende GDSP, Grattapaglia D (2005) Power of microsatellite markers for fingerprinting and parentage analysis in *Eucalyptus grandis* breeding populations. *J Hered* 96:161–166
- Knott S, Neale DN, Sewell MM, Haley C (1997) Multiple marker mapping of quantitative trait loci in an outbred pedigree of loblolly pine. *Theor Appl Genet* 94:810–820
- Komulainen P, Brown GR, Mikkonen M, Karhu A, Garcia-Gil MR, O'Malley DM, Lee B, Neale DB, Savolainen O (2003) Comparing EST-based genetic maps between *Pinus sylvestris* and *Pinus taeda*. *Theor Appl Genet* 107:667–678
- Kondo T, Terada K, Hayashi E, Kuramoto N, Okamura M, Kawasaki H (2000) RAPD markers linked to a gene for resistance to pine needle gall midge in Japanese black pine (*Pinus thunbergii*). *Theor Appl Genet* 100:391–395
- Korol AB, Ronin YI, Kirzhner VM (1995) Interval mapping of quantitative trait loci employing correlated traits complexes. *Genetics* 140:1137–1147
- Korol AB, Ronin YI, Nevo E, Hayes PM (1998) Multi-interval mapping of correlated trait complexes. *Heredity* 80:273–284
- Kossack DS, Kinlaw CS (1999) IFG, a gypsy-like retrotransposon in *Pinus* (Pinaceae), has an extensive history in pines. *Plant Mol Biol* 39:417–426
- Kremer A (1992) Predictions of age-age correlations of total height based on serial correlations between height increments in Maritime pine (*Pinus pinaster* Ait.). *Theor Appl Genet* 85:152–158
- Kriebel HB (1985) DNA sequence components of the *Pinus strobus* nuclear genome. *Can J For Res* 15:1–4
- Kruglyak L (1999) Prospects for whole-genome linkage disequilibrium mapping of common disease genes. *Nat Genet* 22:139–144
- Krupkin AB, Liston A, Strauss SH (1996) Phylogenetic analysis of the hard pines (*Pinus* subgenus *Pinus*, Pinaceae) from chloroplast DNA restriction site analysis. *Am J Bot* 83:489–498
- Krutovsky KV, Neale DB (2005) Nucleotide diversity and linkage disequilibrium in cold hardiness and wood quality related candidate genes in Douglas-fir. *Genetics* 171:2029–2041
- Krutovsky KV, Troggio M, Brown GR, Jermstad KD, Neale DB (2004) Comparative mapping in the Pinaceae. *Genetics* 168:447–461
- Kuang H, Richardson TE, Carson SD, Bongarten B (1999b) Genetic analysis of inbreeding depression in plus tree 850.55 of *Pinus radiata* D. Don. II. Genetics of viability genes. *Theor Appl Genet* 99:140–146
- Kuang H, Richardson TE, Carson SD, Wilcox PL, Bongarten B (1999a) Genetic analysis of inbreeding depression

- sion in plus tree 850.055 of *Pinus radiata* D. Don. 1. Genetic map with distorted markers. *Theor Appl Genet* 98:697–703
- Kubisiak TL, Nelson CD, Nance WL, Stine M (1995) RAPD linkage mapping in a longleaf pine $\times$ slash pine F_1 family. *Theor Appl Genet* 90:1119–1127
- Kubisiak TL, Nelson CD, Nance WL, Stine M (1996) Comparison of RAPD linkage maps constructed for a single longleaf pine from both haploid and diploid mapping populations. *For Genet* 3:230–211
- Kubisiak TL, Nelson CD, Nowak J, Friend AL (2000) Genetic linkage mapping of genomic regions conferring tolerance to high aluminum in slash pine. *J Sust For* 10:69–78
- Kubisiak TL, Nelson, CD, Stine M (1997) RAPD mapping of genomic regions influencing early height growth in longleaf pine $\times$ slash pine F_1 hybrids. In: *Proc 24th Southern For Tree Improv Conf*, Orlando, FL, 9–12 June 1997, pp 198–206
- Kumar S (2004) Genetic parameter estimates for wood stiffness, strength, internal checking and resin bleeding for radiata pine. *Can J For Res* 34:2601–2610
- Kumar S, Garrick DJ (2001) Genetic response to within-family selection using molecular markers in some radiata pine breeding schemes. *Can J For Res* 31:779–785
- Kumar S, Richardson TE (2005) Inferring heritability and relatedness using molecular markers in radiata pine. *Mol Breed* 15:55–64
- Kumar S, Spelman R, Garrick D, Richardson TE, Wilcox PL (2000) Multiple marker mapping of quantitative trait loci on chromosome *three* in an outbred pedigree of radiata pine. *Theor Appl Genet* 100:926–933
- Kumar S, Echt C, Wilcox PL, Richardson TE (2004) Testing for linkage disequilibrium in the New Zealand radiata pine breeding population. *Theor Appl Genet* 108:292–298
- Kumar S, Gerber S, Richardson TE (2006) Pedigree reconstruction using SSR markers in a radiata pine breeding programme. In: *Proc 13th Australasian Plant Breed Conf*, 17–21 April, Christchurch, New Zealand pp 578–583, <http://www.apbc.org.nz>
- Kutil BL, Williams CG (2001) Triplet-repeat microsatellites shared among hard and soft pines. *J Hered* 92:327–32
- Lambeth C (2000) Realized genetic gains for first generation improved loblolly pine in 45 tests in coastal North Carolina. *J Appl For* 24:140–144
- Lambeth C, Lee BC, O'Malley D, Wheeler N (2001) Polymix breeding with parental analysis of progeny: an alternative to full-sib breeding and testing. *Theor Appl Genet* 103:930–943
- Lamoureux D, Peterson DG, Li W, Fellers JP, Gill BS (2005) The efficacy of Cot-based gene enrichment in wheat (*Triticum aestivum* L.). *Genome* 48:1120–1126
- Lamprecht H (1990) Silviculture in the Tropics: tropical forest ecosystems and their tree species-possibilities and methods for their long-term utilization. GTZ, Eschborn, Germany
- Lande R, Thompson R (1990) Efficiency of marker-assisted selection in the improvement of quantitative traits. *Genetics* 124:743–756
- Lander ES, Botstein D (1989) Mapping Mendelian factors underlying quantitative traits using RFLP linkage maps. *Genetics* 121:185–199
- Laurie CC, Chasalow SD, LeDeaux JR, McCarroll R, Bush D, Hauge B, Lai C, Clark D, Rocheford TR, Dudley JW (2004) The genetic architecture of response to long-term artificial selection for oil concentration in the maize kernel. *Genetics* 168:2141–2155
- Le Dantec L, Chagné D, Pot D, Cantin O, Garnier-Géré P, Bedon F, Frigerio JM, Chaumeil P, Léger P, García V, Laigret F, de Daruvar A, Plomion C (2004) Automated SNP detection in expressed sequence tags: statistical considerations and application to maritime pine sequences. *Plant Mol Biol* 54:461–470
- Le Provost G, Paiva J, Pot D, Brach J, Plomion C (2003) Seasonal variation in transcript accumulation in wood forming tissues of maritime pine (*Pinus pinaster* Ait.) with emphasis on a cell wall Glycine Rich Protein. *Planta* 217:820–830
- Ledig FT (1998) Genetic variation in *Pinus*. In: Richardson DM (ed) *Ecology and Biogeography of Pinus*. Cambridge University Press, Cambridge, UK, pp 251–280
- Ledig FT, Conkle MT (1983) Gene diversity and genetic structure in a narrow endemic Torrey pine (*Pinus torreyana* Parry ex Carr). *Evolution* 37:79–85
- Leitch IJ, Hanson L, Winfield M, Parker J, Bennett MD (2001) Nuclear DNA C-values complete familial representation in gymnosperms. *Ann Bot* 88:843–849
- Lerceteau EC, Plomion C, Andersson B (2000) AFLP mapping and detection of quantitative trait loci (QTLs) for economically important traits in *Pinus sylvestris*: a preliminary study. *Mol Breed* 6:451–458
- Li C, Yeh FC (2001) Construction of a framework map in *Pinus contorta* subsp. *latifolia* using random amplified polymorphic DNA markers. *Genome* 44:147–153
- Liston A, Robinson WA, Pinero D, Alvarez-Buylla ER (1999) Phylogenetics of *Pinus* (Pinaceae) based on nuclear ribosomal DNA internal transcribed spacer region sequences. *Mol Phylogenet Evol* 11:95–109
- Liu ZL, Zhang D, Hong DY, Wang XR (2003) Chromosomal localization of 5S and 18S–5.8S–25S ribosomal DNA sites in five Asian pines using fluorescence *in situ* hybridization. *Theor Appl Genet* 106:198–204
- Lopez-Upton J, White TL, Huber DA (2000) Species differences in early growth and rust incidence of loblolly and slash pine. *For Ecol Mng* 132:211–222
- Lorenz WW, Dean JFD (2002) SAGE profiling and demonstration of differential gene expression along the axial developmental gradient of lignifying xylem in loblolly pine (*Pinus taeda*). *Tree Physiol* 22:301–310
- Lorenz WW, Sun F, Liang C, Kolychev D, Wang H, Zhao X, Cordonnier-Pratt MM, Pratt LH, Dean JFD (2005) Water stress-responsive genes in loblolly pine (*Pinus taeda*) roots

- identified by analyses of expressed sequence tag libraries. *Tree Physiol* 26:1–16
- Lorenz WW, Sun F, Liang C, Kolychev D, Wang H, Zhao X, Cordonnier-Pratt MM, Pratt LH, Dean JF (2006) Water stress-responsive genes in loblolly pine (*Pinus taeda*) roots identified by analyses of expressed sequence tag libraries. *Tree Physiol* 26:1–16
- Lowe WJ, Byram TD, Bridgwater FE (1999) Selecting loblolly pine parents for seed orchards to minimise the costs of producing pulp. *For Sci* 45:213–216
- Lubaretz O, Fuchs J, Ahne R, Meister A, Schubert I (1996) Karyotyping of three *Pinaceae* species via fluorescent *in situ* hybridization and computer-aided chromosome analysis. *Theor Appl Genet* 92:411–416
- Luikart G, England P, Tallmon D, Jordan S, Taberlet P (2003) The power and promise of population genomics: from genotyping to genome typing. *Nat Rev Genet* 4:981–994
- Lund MS, Sorensen P, Guldbrandsen B, Sorensen DA (2003) Multitrait fine mapping of quantitative trait loci using combined linkage disequilibrium and linkage analysis. *Genetics* 163:405–410
- Ma XF, Szmidt A, Wang WR (2006) Genetic structure and evolutionary history of a diploid hybrid pine *Pinus densata* inferred from the nucleotide variation at seven gene loci. *Mol Biol Evol* 23:807–816
- MacPherson P, Filion WG (1981) Karyotype analysis and the distribution of constitutive heterochromatin in five species of *Pinus*. *J Hered* 72:193–198
- Margulies M, Egholm M, Altman WE, Attiya S, Bader JS, Bemben LA, Berka J, Braverman MS, Chen YJ, Chen Z, Dewell SB, Du L, Fierro JM, Gomes XV, Godwin BC, He W, Helgesen S, Ho CH, Irzyk GP, Jando SC, Alenquer ML, Jarvie TP, Jirage KB, Kim JB, Knight JR, Lanza JR, Leamon JH, Lefkowitz SM, Lei M, Li J, Lohman KL, Lu H, Makhijani VB, McDade KE, McKenna MP, Myers EW, Nickerson E, Nobile JR, Plant R, Puc BP, Ronan MT, Roth GT, Sarkis GJ, Simons JE, Simpson JW, Srinivasan M, Tartaro KR, Tomasz A, Vogt KA, Volkmer GA, Wang SH, Wang Y, Weiner MP, Yu P, Begley RF, Rothberg JM (2005) Genome sequencing in microfabricated high-density picolitre reactors. *Nature* 437:376–380
- Mariette S, Chagné D, Decroocq S, Vendramin GG, Lalanne C, Madur D, Plomion C (2001) Microsatellite markers for *Pinus pinaster* Ait. *Ann For Sci* 58:203–206
- Markussen T, Fladung M, Achere V, Favre JM, Faivre-Rampant P, Aragones A, Da Silva Perez, Havengt L, Ritter E (2003) Identification of QTLs controlling growth, chemical and physical wood property traits in *Pinus pinaster* (Ait.). *Silvae Genet* 52:8–15
- Marquardt PE, Epperson BK (2004) Spatial and population genetic structure of microsatellites in white pine. *Mol Ecol* 13:3305–3315
- Matheson AC, Lindgren D (1985) Gain from the clonal and the clonal seed orchard options compared for tree breeding programs. *Theor Appl Genet* 71:242–249
- Matziris DI (1995) Provenance variation of *Pinus radiata* grown in Greece. *Silvae Genet* 44:88–96
- McKeand S, Mullin T, Byram T, White TL (2003) Deployment of genetically improved loblolly and slash pines in the South USA. *J For* 101:32–37
- Mergen F (1958) Natural polyploidy in slash pine. *For Sci* 4:283–295
- Miksche JP, Hotta Y (1993) DNA base composition and repetitive DNA in several conifers. *Chromosoma* 41:29–36
- Millar CI (1998) Early evolution of pines. In: Richardson DM (ed) *Ecology and Biogeography of Pinus*. Cambridge University Press, Cambridge, UK, pp 69–91
- Mitton JB (1992) The dynamic mating system of conifers. *New For* 6:187–216
- Moore G, Devos KM, Wang Z, Gale MD (1995) Grasses, line up and form a circle. *Curr Biol* 5(7):737–739
- Moran GF, Bell JC, Hilliker AJ (1983) Greater meiotic recombination in male vs. female gametes in *Pinus radiata*. *J Hered* 74:62
- Morgante M, Salamini F (2003) From plant genomics to breeding practice. *Curr Opin Biotechnol* 14:214–219
- Morgante M, Vendramin GG, Rossi P, Olivieri AM (1993) Selection against inbreds in early life-cycle phases in *Pinus leucodermis*. *Heredity* 70:622–627
- Morin PA, Luikart G, Wayne RK, and the SNP workshop group (2004) SNPs in ecology, evolution and conservation. *Trends Ecol Evol* 19:208–216
- Morse AM, Nelson CD, Covert SF, Holliday AG, Smith KE, Davis JM (2004) Pine genes regulated by the necrotrophic pathogen *Fusarium circinatum*. *Theor Appl Genet* 109:922–932
- Morse AM, Nelson CD, Covert SF, Smith KE, Davis JM (2004) Pine genes regulated by the necrotrophic pathogen, *Fusarium circinatum*. *Theoretical and Applied Genetics* 109:922–932
- Morton NE (1991) Parameters of the human genome. *Proc Natl Acad Sci USA* 88:7474–7476
- Mosig MO, Lipkin E, Khutoreskaya G, Tchourzyna E, Soller M, Friedmann A (2001) A whole genome scan for quantitative trait loci affecting milk protein percentage in Israeli-Holstein cattle, by means of selective milk DNA pooling in a Daughter design, using an adjusted false discovery rate criterion. *Genetics* 157:1683–1698
- Moura VPG, Dvorak WS, Hodge GR (1998) Provenance and family variation of *Pinus oocarpa* grown in the Brazilian cerrado. *For Ecol Manage* 109:315–322
- Müller-Starck G (1998) Isozymes. In: Karp A, Isaac PG, Ingram DS (eds) *Molecular Tools for Screening Biodiversity: Plants and Animals*. Chapman and Hall, London, UK, pp 75–81
- Muona O, Harju A (1989) Effective population sizes, genetic variability and mating system in natural stands and seed orchards of *Pinus sylvestris*. *Silvae Genet* 38:221–228
- Muona O, Yazdani R, Rudin R (1987) Genetic change between life stages in *Pinus sylvestris*: allozymes variation in seeds and planted seedlings. *Silvae Genet* 35:39–42

- Murray B (1998) Nuclear DNA amounts in gymnosperms. *Ann Bot* 82(Suppl A):3-15
- Myburg H, Morse AM, Amerson HV, Kubisiak TL, Huber DA, Osborne JA, Garcia SA, Nelson CD, Davis JM, Covert SF, van Zyl LM (2006) Differential gene expression in loblolly pine (*Pinus taeda* L.) challenged with the fusiform rust fungus, *Cronartium quercuum* f. sp. *fusiforme*. *Physiological and Molecular Plant Pathology* 69:79-101
- Myers RM, Maniatis T, Lerman LS (1987) Detection and localization of single base changes by denaturing gradient gel electrophoresis. *Methods Enzymol* 155:501-527
- Neale DB, Savolainen O (2004) Association genetics of complex traits in conifers. *Trends Plant Sci* 9:325-330
- Neale DB, Sederoff RR (1989) Paternal inheritance of chloroplast DNA and maternal inheritance of mitochondrial DNA in loblolly pine. *Theor Appl Genet* 77:212-216
- Neale DB, Williams CG (1991) Restriction fragment length polymorphism mapping in conifers and applications to forest genetics and tree improvement. *Can J For Res* 21:545-554
- Nelson CD, Echt CS (2003) New models for marker-assisted selection in tree breeding. In: McKinley C (ed) *Proc 27th South For Tree Improv Conf*, Stillwater, OK, p 114
- Nelson CD, Echt CS (2004) Marker-directed population improvement. In: Li B, McKeand S (eds) *Proc IUFRO Joint Conf of Div 2, For Genet Tree Breed in the Age of Genomics: Progress and Future*, Charleston, SC, p 255
- Nelson CD, Kubisiak TL, Johnson G, Burdine C, Bridgwater FE (2003) Microsatellite analysis of loblolly pine. In: *Plant & Animal Genome XI Conf*, San Diego, p 560 (http://www.intl-pag.org/11/abstracts/P5i_P560_XI.html, accessed 20 March 2006)
- Nelson CD, Kubisiak TL, Stine M, Nance WL (1994) A genetic linkage map of longleaf pine (*Pinus palustris* Mill.) based on random amplified polymorphic DNAs. *J Hered* 85:433-439
- Nelson CD, Nance WL, Doudrick RL (1993) A partial genetic linkage map of slash pine (*Pinus elliotii* Engelm. var. *elliottii*) based on random amplified polymorphic DNAs. *Theor Appl Genet* 8:145-151
- Nelson JC (1997) QGENE: software for marker-based genomic analysis and breeding. *Mol Breed* 3:239-245
- Nikles DG (2000) Experience with some *Pinus* hybrids in Queensland, Australia. In: Dungey HS, Dieters MJ, Nikles DG (Compl) *Hybrid Breeding and Genetics of Forest Trees*. *Proc of QFRI/CRC-SPF Symp*, 9-14 April 2000, Noosa, Queensland, Australia. Dept of Pri Indus, Brisbane, pp 14-26
- No E, Zhou Y, Loopstra CA (2000) Sequences upstream and downstream of two xylem-specific pine genes influence their expression. *Plant Sci* 160:77-86
- NSF (2005) NSF, USDA and DOE Award \$32 Million to Sequence Corn Genome. Press Release 05-197. National Science Foundation. 11-15-2005. http://www.nsf.gov/news/news_summ.jsp?cntn_id=104608
- O'Farrell PH (1975) High resolution two-dimensional electrophoresis of proteins *J Biol Chem* 250:4007-4021
- O'Malley D, Grattapaglia D, Chaparro JX, Wilcox PL, Amerson HV, Liu B-H, Whetton R, McKeand S, Kuhlman EG, McCord S, Crane B, Sederoff RR (1996) Molecular markers, forest genetics and tree breeding. In: Gustafson JP, Flavell RB (eds) *Genomes of Plants and Animals: 21st Stadler Genet Symp*. Plenum, New York
- Ohri D, Khoshoo TN (1986) Genome size in gymnosperms. *Plant Syst Evol* 153:119-131
- Olsson T, Lindgren D, Li B (2001) Balancing genetic gain and relatedness in seed orchards. *Silvae Genet* 50:222-227
- Orita M, Iwahana H, Kanazawa H, Hayashi K, Sekiya T (1989) Detection of polymorphisms of human DNA by gel electrophoresis as single-strand conformation polymorphisms. *Proc Natl Acad Sci USA* 86:2766-2770
- Ott J (1991) *Analysis of Human Genetic Linkage*. Johns Hopkins University Press, Baltimore, MD
- Pacheco D, Díaz S, Osuna D, Bautista R, Claros MG, Cánovas FM, Cantón FR (2005) Identification of candidate genes conferring specific properties to distinct types of *Pinus pinaster* Ait. wood. *IUFRO Tree Biotechnol*, Pretoria, South Africa
- Paiva J (2006) Phenotypic and molecular plasticity of wood forming tissues in maritime pine (*Pinus pinaster* Ait.). PhD thesis of Universidade Nova de Lisboa and Bordeaux 1
- Paran I, Zamir D (2003) Quantitative traits in plants: beyond the QTL. *Trends Genet* 19:303-306
- Park YS (2002) Implementation of conifer somatic embryogenesis in clonal forestry: technical requirements and deployment considerations. *Ann For Sci* 59:651-656
- Paterson AH, Freeling M, Sasaki T (2005) Grains of knowledge: genomics of model cereals. *Genome Res* 15:1643-1650
- Paul AD, Foster GS, Caldwell T, McRae J (1997) Parameters for height, diameter, and volume in a multilocation clonal study with loblolly pine. *For Sci* 43:87-98
- Pavy N, Laroche J, Bousquet J, MacKay J (2005) Large-scale analysis of secondary xylem ESTs in pine. *Plant Mol Biol* 57:203224
- Perlack RD, Wright LL, Turhollow AF, Graham RL, Stokes BJ, Erbach DC (2005) Biomass as a feedstock for a bioenergy and bioproducts industry: the technical feasibility of a billion-ton annual supply. Report No DOE/GO-102005-2135. US Dept of Energy/US Dept of Agri. http://www1.eere.energy.gov/biomass/pdfs/final_billionton_vision_report2.pdf
- Peterson DG (2005) Reduced representation strategies and their application to plant genomes. In: Meksem K, Kahl G (eds) *The Handbook of Genome Mapping: Genetic and Physical Mapping*. Wiley, Weinheim, pp 307-335
- Peterson DG, Chouvarine P, Thummasuwan S, Saha S, Mukherjee D, Carlson JE (2006) Exploring the pine genome using Cot filtration and 454 Life Sciences massively parallel shotgun sequencing. In: *Plant & Animal Genomes XIV Conf*, San Diego. http://www.intl-pag.org/pag/14/abstracts/PAG14_W376.html

- Peterson DG, Nelson CD, Islam-Faridi MN, Main DS, Tomkins JP (2005) Accelerating pine genomics through development and utilization of molecular and cytogenetic resources. In: Plant & Animal Genomes XIII Conf, San Diego. http://www.intl-pag.org/pag/13/abstracts/PAG13_P515.html
- Peterson DG, Schulze SR, Sciara EB, Lee SA, Bowers JE, Nagel A, Jiang N, Tibbitts DC, Wessler SR, Paterson AH (2002) Integration of Cot analysis, DNA cloning, and high-throughput sequencing facilitates genome characterization and gene discovery. *Genome Res* 12:795–807
- Petit RJ, Duménil J, Fineschi S, Hampe A, Salvini D, Vendramin GG (2005) Comparative organisation of chloroplast, mitochondrial and nuclear diversity in plant populations. *Mol Ecol* 14:689–701
- Petit RJ, Vendramin GG (2007) Phylogeography of organelle DNA in plants: an introduction. In: Weiss S, Ferrand N (eds) *Phylogeography of Southern European Refugia*. Kluwer, Amsterdam, (in press)
- Pflieger S, Lefebvre V, Causse M (2001) The candidate gene approach in plant genetics: a review. *Mol Breed* 7:275–291
- Plomion C, Durel C-E (1996) Estimation of the average effects of specific alleles detected by the pseudo-testcross QTL mapping strategy. *Gen Sel Evol* 28:223–235
- Plomion C, O'Malley DM (1996) Recombination rate differences for pollen parents and seed parents in pine. *Heredity* 77:341–350
- Plomion C, Bahrman N, Durel CE, O'Malley DM (1995a) Genomic analysis in *Pinus pinaster* (Maritime pine) using RAPD and protein markers. *Heredity* 74:661–668
- Plomion C, O'Malley DM, Durel CE (1995b) Genomic analysis in Maritime pine (*Pinus pinaster*). Comparison of two RAPD maps using selfed and open-pollinated seeds of the same individual. *Theor Appl Genet* 90:1028–1034
- Plomion C, Durel C-E, O'Malley D (1996a) Genetic dissection of height in maritime pine seedlings raised under accelerated growth condition. *Theor Appl Genet* 93:849–858
- Plomion C, Yani A, Marpeau A (1996b) Genetic determinism of β -carene in maritime pine using random amplified polymorphic DNA (RAPD) markers. *Genome* 39:1123–1127
- Plomion C, Liu BH, O'Malley DM (1996c) Genetic analysis using trans dominant linked markers in an F₂ family. *Theor Appl Genet* 93:1083–1089
- Plomion C, Costa P, Bahrman N, Frigerio JM (1997) Genetic Analysis of needle proteins in maritime pine. 1. Mapping dominant and codominant protein markers assayed on diploid tissue, in a haploid-based genetic map. *Silvae Genet* 46:161–165
- Plomion C, Hurme P, Frigerio J-M, Ridolphi M, Pot D, Pionneau C, Avila C, Gallardo F, David H, Neutlings G, Campbell M, Canovas FM, Savolainen O, Bodénès C, Kremer A (1999) Developing SSCP markers in two *Pinus* species. *Mol Breed* 5:21–31
- Plomion C, Pionneau C, Brach J, Costa P, Baillères H (2000) Compression wood-responsive proteins in developing xylem of maritime pine (*Pinus pinaster* Ait.). *Plant Physiol* 123:959–969
- Plomion C, LeProvost G, Pot D, Vendramin G, Gerber S, Decroocq S, Brach J, Raffin A, Pastuszka P (2001) Pollen contamination in a maritime pine polycross seed orchard and certification of improved seeds using chloroplast microsatellites. *Can J For Res* 31:1816–1825
- Plomion C, Bahrman N, Costa P, Dubos C, Frigério J-M, Gerber S, Gion J-M, Lalanne C, Madur D, Pionneau C (2004) Proteomics for genetics and physiological studies in forest trees: application in maritime pine. In: Kumar S, Fladung M (eds) *Molecular Genetics and Breeding of Forest Trees*. Haworth, Binghamton, NY, pp 53–80
- Pot D (2004) Déterminisme génétique de la qualité du bois chez le pin maritime: du phénotype aux gènes. PhD thesis, University of Rennes I, ENSAR, France
- Pot D, Chantre G, Rozenberg P, Rodrigues JC, Jones GL, Pereira H, Hannrup B, Cahalan C, Plomion C (2002) Genetic control of pulp and timber properties in maritime pine (*Pinus pinaster* Ait.). *Ann For Sci* 59:563–575
- Pot D, McMillan L, Echt C, Le Provost G, Garnier-Géré P, Cato S, Plomion C (2005a) Nucleotide variation in genes involved in wood formation in two pine species. *New Phytol* 167:101–112
- Pot D, Rodrigues J-C, Rozenberg P, Chantre G, Tibbitts J, Cahalan C, Pichavant F, Plomion C (2005b) QTLs and candidate genes for wood properties in maritime pine (*Pinus pinaster* Ait.). *Tree Genet Genom* 2:10–26
- Price RA, Liston A, Strauss SH (1998) Phylogeny and systematics of *Pinus*. In: Richardson DM (ed) *Ecology and Biogeography of Pinus*. Cambridge University Press, Cambridge, UK, pp 49–68
- Provan J, Soranzo N, Wilson NJ, Goldstein DB, Powell W (1999) A low mutation rate for chloroplast microsatellites. *Genetics* 153:943–947
- Quackenbush J, Cho J, Lee D, Liang F, Holt I, Karamycheva S, Parvizi B, Perteau G, Sultana R, White J (2001) The TIGR Gene Indices: analysis of gene transcript sequences in highly sampled eukaryotic species. *Nucleic Acids Res* 29:159–164
- Quencez C, Bastien C (2001) Genetic variation within and between populations of *Pinus sylvestris* L. (Scots pine) for susceptibility to *Melampsora pinitorqua* Rostr. (pine twist rust). *Heredity* 86:36–44
- Rabinowicz PD, Citek R, Budiman MA, Nunberg A, Bedell JA, Lakey N, O'Shaughnessy AL, Nascimento LU, McCombie WR, Martienssen RA (2005) Differential methylation of genes and repeats in land plants. *Genome Res* 15:1431–1440
- Rabinowicz PD, Schutz K, Dedhia N, Yordan C, Parnell LD, Stein L, McCombie WR, Martienssen RA (1999) Differential methylation of genes and retrotransposons facilitates shotgun sequencing of the maize genome. *Nat Genet* 23:305–308
- Rafalski A (1998) Randomly amplified polymorphic DNA (RAPD) analysis. In: Caetano-Anollés G, Gresshoff

- PM (eds) DNA Markers Protocols, Applications, and Overviews. Wiley-Liss, New York, pp 75–83
- Rake AV, Miksche JP, Hall RB, Hansen KM (1980) DNA reassociation kinetics of four conifers. *Can J Genet Cytol* 22:69–79
- Rat Genome Sequencing Project Consortium (2004) Rat Genome Sequencing Project Consortium, Genome sequence of the Brown Norway rat yields insights into mammalian evolution. *Nature* 428:493–521
- Remington DL, O'Malley DM (2000a) Evaluation of major genetic loci contributing to inbreeding depression for survival and early growth in a selfed family of *Pinus taeda*. *Evolution* 54:1580–1589
- Remington DL, O'Malley DM (2000b) Whole genome characterization of embryonic state inbreeding depression in a selfed loblolly pine family. *Genetics* 155:337–348
- Remington DL, Whetten RW, Liu BH, O'Malley DM (1999) Construction of an AFLP genetic map with nearly complete genome coverage in *Pinus taeda*. *Theor Appl Genet* 98:1279–1292
- Ribeiro MM, Plomion C, Petit RJ, Vendramin GG, Szmidt AE (2001) Variation of chloroplast single-sequence repeats in Portuguese maritime pine (*Pinus pinaster* Ait.). *Theor Appl Genet* 102:97–103
- Richardson BA., Brunsfeld SJ, Klopfenstein NB (2002) DNA from bird-dispersed seed and wind-disseminated pollen provides insights into postglacial colonization of whitebark pine (*Pinus albicaulis*). *Mol Ecol* 11:215–227
- Richardson DM, Rundel PW (1998) Ecology and biogeography of *Pinus*: an introduction. In: Richardson DM (ed) *Ecology and Biogeography of Pinus*. Cambridge University Press, Cambridge, UK, pp 3–46
- Ritland K, Zhuang J, Ralph S, Ritland C, Bohlmann J (2006) Comparative conifer genomics and the new “Treenomix” program. In: *Plant & Animal Genomes XIV Conf*, San Diego, W124
- Ritter E, Aragonés A, Markussen T, Acheré V, Espinel S, Fladung M, Wrobel S, Faivre-Rampant P, Jeandroz S, Favre JM (2002) Toward construction of an ultra high density linkage map for *Pinus pinaster*. *Ann For Sci* 59:637–643
- Robledo-Arnuncio JJ, Collada C, Alfá R, Gil L (2005) Genetic structure of montane isolates of *Pinus sylvestris* L. in a Mediterranean refugial area. *J Biogeogr* 32:595–605
- Rosvall O, Lindgren D, Mullen TJ (1998) Sustainability, robustness and efficiency of a multi-generation breeding strategy based on within-family clonal selection. *Silvae Genet* 47:307–321
- Rounsley S, Xiaoying L, Ketchum KA (1998) Large-scale sequencing of plant genomes. *Curr Opin Plant Biol* 1:136–141
- Rozenberg P, Van Loo J, Hannrup B, Grabner M. 2002. Clonal variation of wood density record of cambium reaction to water deficit in *Picea abies* (L.) Karst. *Ann For Sci* 59:533–540
- Rudd S (2003) Expressed sequence tags: alternative or complement to whole genome sequences? *Trends Plant Sci* 8:321–329
- Rudin D, Ekberg I (1978) Linkage studies in *Pinus sylvestris* L. using macrogametophyte allozymes. *Silvae Genet* 27:1–12
- Sanchez AC, Ilag LL, Yang D, Brar DS, Ausubel F, Khush GS, Yano M, Sasaki T, Li Z, Huang N (1999) Genetic and physical mapping of *xa13*, a recessive bacterial blight resistance gene in rice. *Theor Appl Genet* 98:1022–1028
- Sánchez C, Vielva MJ, Vieitez AM, de Mier B, Abarca D, Díaz-Sala C (2005) Identification of genes related to adventitious rooting capacity in pine and chestnut. *IUFRO Tree Biotechnol*, Pretoria, South Africa
- Sathyan P, Newton RJ, Loopstra CA (2005) Genes induced by WDS are differentially expressed in two populations of aleppo pine (*Pinus halepensis*). *Tree Genet Genom* 1:166–173
- Sax K (1923) The association of size differences with seed-coat pattern and pigmentation in *Phaseolus vulgaris*. *Genetics* 8:552–560
- Sax K, Sax HJ (1933) Chromosome numbers and morphology in the conifers. *J Arnold Arbor* 14:356–375
- Saylor LC (1961) A karyotype analysis of selected species of *Pinus*. *Silvae Genet* 10:77–85
- Saylor LC (1964) Karyotype analysis of *Pinus* –group *Lariciones*. *Silvae Genet* 13:165–170
- Saylor LC (1972) Karyotype analysis of *Pinus* –subgenus *Pinus*. *Silvae Genet* 21:155–163
- Saylor LC (1983) Karyotype analysis of the genus *Pinus* –subgenus *Strobus*. *Silvae Genet* 32:119–121
- Schmidt A, Doudrick RL, Heslop-Harrison JS, Schmidt T (2000) The contribution of short repeats of low sequence complexity to large conifer genomes. *Theor Appl Genet* 101:7–14
- Schmidt R (2002) Plant genome evolution: lessons from comparative genomics at the DNA level. *Plant Mol Biol* 48:21–37
- Scotti I, Burelli A, Cattonaro F, Chagné D, Fuller J, Hedley PE, Jansson G, Lalanne C, Madur D, Neale D, Plomion C, Powell W, Troggio M, Morgante M (2005) Analysis of the distribution of marker classes in a genetic linkage map: a case study in Norway spruce (*P. abies* karst). *Tree Genet Genom* 1:93–102
- Scotti-Saintagne C, Bodénès C, Barreneche T, Bertocchi E, Plomion C, Kremer A (2004) Detection of quantitative trait loci controlling bud burst and height growth in *Quercus robur* L. *Theor Appl Genet* 109:1648–1659
- Sewell MM, Bassoni DL, Megraw RA, Wheeler NC, Neale DB (2000) Identification of QTLs influencing wood property traits in loblolly pine (*Pinus taeda* L.). I. Physical wood properties. *Theor Appl Genet* 101:1273–1281
- Sewell MM, Davis MF, Tuskan GA, Wheeler NC, Elam CC, Bassoni DL, Neale DB (2002). Identification of QTLs influencing wood property traits in loblolly pine (*Pinus taeda* L.). II. Chemical wood properties. *Theor Appl Genet* 104:214–222
- Sewell MM, Neale DB (2000) Mapping quantitative traits in forest trees. In: Jain SM, Minocha SC (eds) *Molecular Biology of Woody Plants*. Kluwer, Dordrecht, pp 407–433
- Sewell MM, Sherman BK, Neale DB (1999) A consensus map for loblolly pine (*Pinus taeda* L.). I. Construction and in-

- egration if individual linkage maps from two outbred three-generation pedigrees. *Genetics* 151:321–330
- Shelbourne CJA (1992) Genetic gains from different kinds of breeding population and seed or plant production populations. *S Afr For J* 160:49–65
- Shelbourne CJA (2000) Some insights on hybrids in forest tree improvement. In: Dungey HS, Dieters MJ, Nikles DG (Compl) *Hybrid Breeding and Genetics of Forest Trees*. Proc QFRI/CRC-SPF Symp, 9–14 April 2000, Noosa, Queensland, Australia. Dept of Pri Indus, Brisbane, pp 53–62
- Shelbourne CJA, Apiolaza LA, Jayawickrama KJS, Sorensson CT (1997) Developing breeding objectives for radiata pine in New Zealand. In: Burdon R, Moore JM (eds) "IUFRO '97 Genetics of Radiata Pine", Proc IUFRO Conf 1–4 Dec and workshop 5 Dec, Rotorua, New Zealand. FRI Bull 203:60–168
- Shepherd M, Cross M, Dieters MJ, Henry R (2003) Genetic maps for *Pinus elliottii* var. *elliottii* and *P. caribaea* var. *hondurensis* using AFLP and microsatellite markers. *Theor Appl Genet* 106:1409–1419
- Shepherd M, Cross M, Dieters MJ, Henry R (2002b) Branch architecture QTL for *Pinus elliottii* var. *elliottii* × *Pinus caribaea* var. *hondurensis* hybrids. *Ann For Sci* 59:617–625
- Shepherd M, Cross M, Maguire TL, Dieters MJ, Williams CG, Henry RJ (2002a) Transpecific microsatellites for hard pines. *Theor Appl Genet* 104:819–827
- Shoulders E (1984) The case for planting longleaf pine. Proc of the Southern Silvicultural Res Conf, 7–8 Nov 1984, Atlanta, GA, pp 255–260
- Smith DN, Devey M (1994) Occurrence and inheritance of microsatellites in *Pinus radiata*. *Genome* 37:977–983
- Soranzo N, Alia R, Provan J, Powell W (2000) Patterns of variation at a mitochondrial sequence-tagged-site locus provides new insights into the postglacial history of European *Pinus sylvestris* populations. *Mol Ecol* 9:1205–1211
- Sorensson CT, Shelbourne CJA (2005) Clonal forestry. In: Colley M (ed) *Forestry Handbook*. NZ Inst of Foresters, pp 92–96
- Springer NM, Xu X, Barbazuk WB (2004) Utility of different gene enrichment approaches toward identifying and sequencing the maize gene space. *Plant Physiol* 136:3023–3033
- Stelzer HE, Goldfarb B (1997) Implementing clonal forestry in the southeastern United States: SRIEG satellite workshop summary. *Can J For Res* 27:442–446
- Stirling B, Yang ZK, Gunter LE, Tuskan GA, Bradshaw HD (2003) Comparative sequence analysis between orthologous regions of the *Arabidopsis* and *Populus* genomes reveals substantial synteny and microcollinearity. *Can J For Res* 33:2245–2251
- Strabala TJ (2004) Expressed sequence tag databases from forestry tree species. In: Kumar S, Fladung M (eds) *Molecular Genetics and Breeding of Forest Trees*. Haworth, Binghamton, NY, pp 19–52
- Strauss SH, Doerksen AH (1990) Restriction fragment analysis of pine phylogeny. *Evolution* 44:1081–1096
- Strauss SH, Lande R, Namkoong G (1992) Limitations of molecular marker-aided selection in forest tree breeding. *Can J For Res* 22:1050–1061
- Stromberg LD, Dudley JD, Rufener GK (1994) Comparing conventional early generation selection with molecular marker assisted selection in maize. *Crop Sci* 34:1221–1225
- Stuber CW, Polacco M, Senior ML (1999) Synergy of empirical breeding, marker-assisted selection, and genomics to increase crop yield. *Crop Sci* 39:1571–1583
- Syring J, Willyard A, Cronn R, Liston A (2005) Evolutionary relationships among *Pinus* (Pinaceae) subsections inferred from multiple low-copy nuclear loci. *Am J Bot* 92:2086–2100
- Syvanen AC (2001) Accessing genetic variation. Genotyping single nucleotide polymorphisms. *Nat Rev Genet* 2:930–942
- Szmidt AE, Muona O (1989) Linkage relationships of allozyme loci in *Pinus sylvestris*. *Heredity* 111:91–97
- Tanksley SD, Nelson JC (1996) Advanced backcross QTL analysis: a method for the simultaneous discovery and transfer of valuable QTLs from unadapted germplasm into elite breeding lines. *Theor Appl Genet* 92:191–203
- Telfer EJ, Echt CS, Nelson CD, Wilcox PL (2006) Comparative mapping in *Pinus radiata* and *P. taeda* reveals co-location of wood density-related QTL. In: *Plant & Animal Genome XIV Conf*, San Diego
- Temesgen B, Brown GR, Harry DE, Kinlaw CS, Sewell MM, Neale DB (2001) Genetic mapping of expressed sequence tag polymorphism (ESTP) markers in loblolly pine (*Pinus taeda* L.). *Theor Appl Genet* 102:664–675
- The *Arabidopsis* Genome Initiative (2000) Analysis of the genome sequence of the flowering plant *Arabidopsis thaliana*. *Nature* 408:796–815
- Thiellment H, Plomion C, Zivy M (2001) Proteomics as a tool for plant genetics and breeding. In: Dunn MJ, Pennington S (eds) *Proteomics: from protein sequence to function*. BIOS, Oxford, pp 289–309
- Thoday JM (1961) Location of polygenes. *Nature* 191:368–370
- Thumma BR, Nolan ME, Evans R, Moran GF (2005) Polymorphisms in *Cinnamoyl CoA Reductase* (CCR) are associated with variation in microfibril angle in *Eucalyptus* spp. *Genetics* 171:1257–1265
- Travis SE, Ritland K, Whitham TG, Keim P (1998) A genetic linkage map of pinyon pine (*Pinus edulis*) based on amplified fragment length polymorphisms. *Theor Appl Genet* 97:871–880
- Tulsieram LK, Glaubitz JC, Kiss G, Carlson JE (1992) Single tree genetic linkage mapping in conifers using haploid DNA from megagametophytes. *Bio/Technology* 10:686–690
- Tuskan GA, DiFazio SP, Teichmann T (2004) Poplar genomics is getting popular: the impact of the poplar genome project on tree research. *Plant Biol (Stuttg)* 6:2–4
- Van der Burgh J (1973) Hölzer der niederrheinischen Braunkohlenformation. 2. Hölzer der Braunkohlengruben

- ,Maria Theresia' zu Herzogenrath, 'Zukunft West' zu Eschweiler und 'Victor' (Zülpich Mitte) zu Zülpich. Nebst einer systematisch-anatomischen Bearbeitung der Gattung *Pinus* L. Rev Palaeobot Palynol 15:73–275
- van Tienderen PH, de Haan AA, van der Linden CG, Vosman B (2002) Biodiversity assessment using markers for ecologically important traits. Trends Ecol Evol 17:577–582
- van Zyl L, von Arnold S, Bozhkov P, Chen Y, Egertsdotter U, MacKay J, Sederoff R, Weir B, Shen J, Sun Y-H, Whetten R, Zelena L, Clapham D (2002) Heterologous array analysis in Pinaceae: hybridization of *Pinus taeda* cDNA arrays with cDNA from needles and embryogenic cultures of *P. taeda*, *P. sylvestris* or *Picea abies*. Comp Funct Genom 3:306–318
- Verhaegen D, Plomion C, Poitel M, Costa P, Kremer A (1998) Quantitative trait dissection analysis in *Eucalyptus* using RAPD markers: 2. linkage disequilibrium in a factorial design between *E. urophylla* and *E. Grandis*. For Genet 5:61–69
- Vos P, Hogers R, Bleeker M, Reijans M, van de Lee T, Hornes M, Friters A, Pot J, Paleman J, Kuiper M, Zabeau M (1995) AFLP: a new technique for DNA fingerprinting. Nucleic Acids Res 23:4407–4414
- Wahlenberg WG (1946) Longleaf Pine. Charles Lathrop Pack Forestry Foundation, Washington, DC
- Wakamiya I, Price HJ, Messina MG, Newton RJ (1996) Pine genome size diversity and water relations. Physiol Planta 96:13–20
- Walker S, Haines R, Dieters M (1996) Beyond 2000. In: Dieters MJ, Matheson AC, Nikles DG, Harwood CE (eds) Clonal Forestry in Queensland. Queensland For Res Inst. Tree Improvement for Sustainable Tropical Forestry. QFRI-IUFRO Conf, Caloundra, Queensland, Australia, 27 Oct–1 Nov 1996, vol 2, pp 351–354
- Walter R, Epperson BK (2001) Geographic pattern of genetic variation in *Pinus resinosa*: area of greatest diversity is not the origin of postglacial populations. Mol Ecol 10:103–111
- Wang XR, Szmidt AE, Nguyen HN (2000) The phylogenetic position of the endemic flat-needle pine *Pinus krempffii* (Pinaceae) from Vietnam based on PCR-RFLP analysis of chloroplast DNA. Plant Syst Evol 220:21–36
- Wang XR, Szmidt AE, Savolainen O (2001) Genetic composition and diploid hybrid speciation of a high mountain pine, *Pinus densata*, native to the Tibetan plateau. Genetics 159:337–346
- Wang XT, Tsumara Y, Yoshimaru H, Nagasaka K, Szmidt A (1999) Phylogenetic relationships of Eurasian pines (*Pinus*, Pinaceae) based on chloroplast *rbcl*, *matK*, *rpl20-rps18* spacer, and *trnV* intron sequences. Am J Bot 86:1742–1753
- Ware D, Jaiswal P, Ni J, Pan X, Chang K, Clark K, Teytelman L, Schmidt S, Zhao W, Cartinhour S, McCouch S, Stein L (2002) Gramene: a resource for comparative grass genomics. Nucleic Acids Res 30:103–105
- Watkinson JI, Sioson AA, Vasquez-Robinet C, Shukla M, Kumar D, Ellis M, Heath LS, Ramakrishnan N, Chevone B, Watson IT, van Zyl L, Egertsdotter U, Sederoff RR, Grene R, et al (2003) Photosynthetic acclimation is reflected in specific patterns of gene expression in drought-stressed loblolly pine. Plant Physiol 133:1702–1716
- Wei RP, Lindgren D, Yeh FC (1997) Expected gain and status number following restricted individual and combined-index selection. Genome 40:1–8
- Weng C, Kubisiak TL, Nelson CD, Stine M (2002) Mapping quantitative trait loci controlling early growth in a (longleaf pine × slash pine) × slash pine BC1 family. Theor Appl Genet 104:852–859
- Weng C, Kubisiak TL, Stine M (1998) SCAR markers in a longleaf pine × slash pine F1 family. For Genet 5:239–247
- Whetten R, Sun Y, Zhang Y, Sederoff RR (2001) Functional genomics and cell wall biosynthesis in loblolly pine. Plant Mol Biol 47:275–291
- Whitelaw CA, Barbazuk WB, Perteu G, Chan AP, Cheung F, Lee Y, van Heeringen S, Karamycheva S, Bennetzen JL, SanMiguel P, Lakey N, Bedford J, Yuan Y, Budiman MA, Resnick A, van Aken S, Utterback T, Riedmuller S, Williams SM, Feldblyum T, Schubert K, Beachy R, Fraser CM, Quackenbush J (2003) Enrichment of gene-coding sequences in maize by genome filtration. Science 302:2118–2120
- Wilcox PL (1995) Genetic dissection of fusiform rust resistance. PhD Thesis, Dept of Forestry, North Carolina State University, Raleigh, NC
- Wilcox PL (1997) Linkage groups, map length, and recombination in *Pinus radiata*. In: Burdon RD, Moore JM (eds) "IUFRO '97 Genetics of Radiata Pine", Proc IUFRO Conf 1–4 Dec and workshop 5 Dec, Rotorua, New Zealand. FRI Bull 203
- Wilcox PL, Burdon RD (2006) Application of association genetics to coniferous forest trees. In: Proc 13th Australasian Plant Breed Conf, Christchurch, New Zealand pp 651–659, <http://www.apbc.org.nz>
- Wilcox PL, Amerson HV, Kuhlman EG, Liu B-H, O'Malley DM, Sederoff RR (1996) Detection of a major gene for resistance to fusiform rust disease in loblolly pine by genomic mapping. Proc Natl Acad Sci USA 93:3859–3864
- Wilcox PL, Richardson TE, Carson SD (1997) Nature of quantitative trait variation in *Pinus radiata*: insights from QTL detection experiments. In: Burdon RD, Moore JM (eds) "IUFRO '97 Genetics of Radiata Pine", Proc IUFRO Conf 1–4 Dec and workshop 5 Dec, Rotorua, New Zealand. FRI Bull 203:304–312
- Wilcox PL, Richardson TE, Corbet GE, Ball RD, Lee JR, Djorovic A, Carson SD (2001a) Framework Linkage Maps of *Pinus radiata* D. Don based on pseudotestcross markers. For Genet 8:109–117
- Wilcox PL, Carson SD, Richardson TE, Ball RD, Horgan GP, Carter P (2001b) Benefit-cost analysis of DNA marker-based selection in progenies of *Pinus radiata* seed orchard parents. Can J For Res 31:2213–2224
- Wilcox PL, Echt CS, Cato SA, McMillan LK, Kumar S, Ball RD, Burdon RD, Pot D (2003) Gene assisted selection – a new paradigm for selection in forest tree species? In: Plant & Animal Genomes XI Conf, San Diego

- Wilcox PL, Cato S, McMillan L, Power M, Ball RD, Burdon RD, Echt CS (2004) Patterns of linkage disequilibrium in *Pinus radiata*. In: Plant & Animal Genome XII Conf, San Diego, W89. http://www.intl-pag.org/12/abstracts/W22_PAG12_89.html
- Wilcox PL, Echt CS, Burdon RD (2007) Gene-assisted selection: applications of association genetics for forest tree breeding. In: Oraguzie N, Rikkerink EHA, Gardiner SA, Nihal De Silva H (eds) Association Mapping in Plants. Springer, Berlin Heidelberg New York, pp 211–247
- Williams CG, Neale DB (1992) Conifer wood quality and marker-aided selection: a case study. *Can J For Res* 22:1009–1017
- Williams C, Megraw RA (1993) Juvenile-mature relationships for wood density in *Pinus taeda*. *Can J For Res* 24:714–722
- Williams JGK, Kubelik AR, Livak KJ, Rafalski JA (1990) DNA polymorphisms amplified by arbitrary primers are useful as genetic markers. *Nucleic Acids Res* 18:6531–6535
- Williams CG, Zhou Y, Hall SE (2001) A chromosomal region promoting outcrossing in a conifer. *Genetics* 159:1283–1289
- Wright SI, Gaut BS (2005) Molecular population genetics and the search for adaptive evolution in plants. *Mol Biol Evol* 22:506–519
- Wu H (2002) Study of early selection in tree breeding. 4. Efficiency of marker aided early selection (MAES). *Silvae Genet* 51:261–269
- Wu J, Krutovskii KV, Strauss SH (1999) Nuclear DNA diversity, population differentiation, and phylogenetic relationship in the Californian closed-cone pines based on RAPD and allozyme markers. *Genome* 42:893–908
- Wu R, Ma C-X, Casella G (2002) Joint linkage and linkage disequilibrium mapping of quantitative trait loci in natural populations. *Genetics* 160:779–792.
- Yan HH, Mudge J, Kim DJ, Shoemaker RC, Cook DR, Young ND (2004) Comparative physical mapping reveals features of microsynteny between *Glycine max*, *Medicago truncatula*, and *Arabidopsis thaliana*. *Genome* 47:141–155
- Yang SH, Loopstra CA (2005) Seasonal variation in gene expression for loblolly pines (*Pinus taeda*) from different geographical regions. *Tree Physiol* 25:1063–107325(8):
- Yazdani R, Nilsson JE, Plomion C, Mathur G (2003) Marker trait association for autumn cold acclimation and growth rhythm in *Pinus sylvestris*. *Scand J For Res* 18:29–38
- Yazdani R, Yeh FC, Rimsha J (1995) Genomic mapping of *Pinus sylvestris* (L.) using random amplified polymorphic DNA markers. *For Genet* 2:109–116
- Yin TM, Huang MR, Wang MX, Zhu LH, Zhai WX (1997) Construction of molecular linkage map in masson pine using RAPD markers and megagametophytes from a single tree. *Acta Bot Sin* 39:607–612
- Yin TM, Wang XR, Andersson B, Lerceteau-Köhler E (2003) Nearly complete maps of *Pinus sylvestris* L. (Scots pine) constructed by AFLP marker analysis in a full-sib family. *Theor Appl Genet* 106:1075–1083
- Yuan Y, SanMiguel PJ, Bennetzen JL (2003) High-Cot sequence analysis of the maize genome. *Plant J* 34:249–255
- Zeng ZB (1993a) Theoretical basis for separation of multiple linked gene effects in mapping quantitative trait loci. *Proc Natl Acad Sci USA* 90:10972–10976
- Zeng ZB (1993b) Precision mapping of quantitative trait loci. *Genetics* 136:1457–1468
- Zeng ZB, Liu J, Stam LF, Kao CH, Mercer JM, Cathy CL (2000) Genetic architecture of a morphological shape difference between two *Drosophila* species. *Genetics* 154:299–310
- Zhang H-B, Wu C (2001) BACs as tools for genome sequencing. *Plant Physiol Biochem* 39:195–209
- Zheng Y, Ennos R, Wang HR, Zheng YQ, Wang HR (1994) Provenance variation and genetic parameters in a trial of *Pinus caribaea* Morelet var. *bahamensis* Barr and Golf. *For Genet* 1:165–174
- Zhou Y, Bui T, Auckland L, Williams CG (2002) Undermethylation as a Source of microsatellites in large plant genomes. *Genome* 34:91–99
- Zhou Y, Gwaze DP, Reyes-Valdes MH, Bui T, Williams CG (2003) No clustering for linkage map based on low-copy and undermethylated microsatellites. *Genome* 46:809–816