

NATIVE WILDFLOWER SEED PRODUCTION WITH LIMITED SUBSURFACE DRIP IRRIGATION

Clinton C. Shock, Erik B. G. Feibert, and Lamont D. Saunders
Malheur Experiment Station
Oregon State University
Ontario, OR

Nancy Shaw
U.S. Forest Service
Rocky Mountain Research Station
Boise, ID

Introduction

Native wildflower seed is needed to restore rangelands of the Intermountain West. Commercial seed production is necessary to provide the quantity of seed needed for restoration efforts. A major limitation to economically viable commercial production of native wildflower (forb) seed is stable and consistent seed productivity over years. Variations in spring rainfall and soil moisture result in highly unpredictable water stress at flowering, seed set, and seed development, which for other seed crops is known to compromise seed yield and quality.

Native wildflower plants are often not competitive with crop weeds in cultivated fields, which also limits seed production. Both sprinkler and furrow irrigation could provide supplemental water for seed production, but these irrigation systems risk further encouraging weeds. Also, sprinkler and furrow irrigation can lead to the loss of plant stand and seed production due to fungal pathogens. By burying drip tapes at 12-inch depth and avoiding wetting of the soil surface, we hope to assure flowering and seed set without encouraging weeds or opportunistic diseases. This trial tested the effects of three irrigation intensities on the seed yield of 13 native forb species.

Materials and Methods

Plant Establishment

Seed of the seven Intermountain West forb species (the first seven species in Table 1) was received in late November in 2004 from the Rocky Mountain Research Station (Boise, ID). The plan was to plant the seed in the fall of 2004, but due to excessive rainfall in October, the ground preparation was not completed and planting was postponed to early 2005. To ensure germination the seed was submitted to a cold stratification treatment. The seed was soaked overnight in distilled water on January 26, 2004, after which the water was drained and the seed soaked for 20 min in a 10 percent by volume solution of 13 percent bleach in distilled water. The water was drained and the seed placed in a thin layer in plastic containers. The plastic containers

had lids with holes drilled to allow air movement. These containers were placed in a cooler set at approximately 34°F. Every few days the seed was mixed and, if necessary, distilled water added to maintain seed moist. In late February, seed of *Lomatium grayi* and *L. triternatum* had started sprouting.

In late February, 2005 drip tape (T-Tape TSX 515-16-340) was buried at 12-inch depth between two rows (30-inch rows) of a Nyssa silt loam with a pH of 8.3 and 1.1 percent organic matter. The drip tape was buried in alternating inter-row spaces (5 ft apart). The flow rate for the drip tape was 0.34 gal/min/100 ft at 8 psi with emitters spaced 16 inches apart, resulting in a water application rate of 0.066 inch/hour.

On March 3, seed of all species (Table 1) was planted in 30-inch rows using a custom-made plot grain drill with disk openers. All seed was planted at 20-30 seeds/ft of row. The *Eriogonum umbellatum* and the *Penstemon* spp. were planted at 0.25-inch depth and the *Lomatium* spp. at 0.5-inch depth. The trial was irrigated with a minisprinkler system (R10 Turbo Rotator, Nelson Irrigation Corp., Walla Walla, WA) for even stand establishment from March 4 to April 29. Risers were spaced 25 ft apart along the flexible polyethylene hose laterals that were spaced 30 ft apart and the water application rate was 0.10 inch/hour. A total of 1.72 inches of water was applied with the minisprinkler system. *Eriogonum umbellatum*, *Lomatium triternatum*, and *L. grayi* started emerging on March 29. All other species except *L. dissectum* emerged by late April. Starting June 24, the field was irrigated with the drip system. A total of 3.73 inches of water was applied with the drip system from June 24 to July 7. Thereafter the field was not irrigated.

Plant stands for *Eriogonum umbellatum*, *Penstemon* spp., *Lomatium triternatum*, and *L. grayi* were uneven. *Lomatium dissectum* did not emerge. None of the species flowered in 2005. In early October, 2005 more seed was received from the Rocky Mountain Research Station for replanting. The blank lengths of row were replanted by hand in the *Eriogonum umbellatum* and *Penstemon* plots. Entire row lengths were replanted using the planter in the *Lomatium* plots. The seed was replanted on October 26, 2005. In the spring of 2006, plant stand of the replanted species was excellent, except for *P. deustus*.

On April 11, 2006 seed of three globemallow species (*Sphaeralcea parvifolia*, *S. grossulariifolia*, *S. coccinea*), two prairie clover species (*Dalea searlsiae*, *D. ornata*), and basalt milkvetch (*Astragalus filipes*) was planted at 30 seeds/ft of row. The field was sprinkler irrigated until emergence. Emergence was poor. In late August of 2006 seed of the three globemallow species was harvested by hand. On November 9, 2006 the six forbs were flailed. On November 10, 2006 the six forbs were replanted. On November 11, the *Penstemon deustus* plots were replanted at 30 seeds/ft of row.

Table 1. Forb species planted at the Malheur Experiment Station, Oregon State University, Ontario, OR (U.S. Department of Agriculture, Natural Resources Conservation Service. 2009).

Species	Common names
<i>Eriogonum umbellatum</i>	Sulfur-flower buckwheat
<i>Penstemon acuminatus</i>	Sharpleaf penstemon, sand-dune penstemon
<i>Penstemon deustus</i>	Scabland penstemon, hot-rock penstemon
<i>Penstemon speciosus</i>	Royal penstemon, sagebrush penstemon
<i>Lomatium dissectum</i>	Fernleaf biscuitroot
<i>Lomatium triternatum</i>	Nineleaf biscuitroot, nineleaf desert parsley
<i>Lomatium grayi</i>	Gray's biscuitroot, 'Gray's lomatium
<i>Sphaeralcea parvifolia</i>	Smallflower globemallow
<i>Sphaeralcea grossulariifolia</i>	Gooseberryleaf globemallow
<i>Sphaeralcea coccinea</i>	Scarlet globemallow, red globemallow
<i>Dalea searlsiae</i>	Searls' prairie clover
<i>Dalea ornata</i>	Western prairie clover
<i>Astragalus filipes</i>	Basalt milkvetch

Irrigation for Seed Production

In April, 2006 the field was divided into 30-ft-long plots. Each plot contained four rows each of *Eriogonum umbellatum*, *P. acuminatus*, *P. speciosus*, *P. deustus*, *L. dissectum*, *L. triternatum*, and *L. grayi*. The experimental design was a randomized complete block with four replicates. The three irrigation treatments were a nonirrigated check, 1 inch per irrigation, and 2 inches per irrigation. Each treatment received four irrigations that were applied approximately every 2 weeks starting with flowering of the forbs. The amount of water applied to each treatment was measured by a water meter and recorded after each irrigation to ensure correct water applications.

In March, 2007 the drip-irrigation system was modified to allow separate irrigation of the species due to different growth habits. The three *Lomatium* spp. were irrigated together and *Penstemon deustus* and *P. speciosus* were irrigated together, but separately from the others. *Penstemon acuminatus* and *Eriogonum umbellatum* were irrigated individually. In early April, 2007 the three globemallow species, two prairie clover species, and basalt milkvetch were divided into plots with a drip-irrigation system to allow the same irrigation treatments that were received by the other forbs.

Irrigation dates can be found in Table 2(a and b). In 2007, irrigation treatments were inadvertently continued after the four irrigations were applied, as in 2006. Irrigation treatments for all species were continued until the last irrigation on June 24 in 2007.

Soil volumetric water content was measured by neutron probe. The neutron probe was calibrated by taking soil samples and probe readings at 8-, 20-, and 32-inch depths during installation of the access tubes. The soil water content was determined volumetrically from the soil samples and regressed against the neutron probe readings, separately for each soil depth. The regression equations were then used to transform the neutron probe readings during the season into volumetric soil water content.

Flowering, Harvesting, and Seed Cleaning

Flowering dates for each species are found in Table 2(a and b). The *Eriogonum umbellatum* and *Penstemon* spp. plots produced seed in 2006, probably because they had emerged in the spring of 2005. The plant stand for *P. deustus* was too poor to result in reliable seed yield estimates. Replanting of *P. deustus* in the fall of 2006 did not result in adequate plant stand in the spring of 2007. Each year, the middle two rows of each plot were harvested when seed of each species was mature (Table 2(a and b)) using the methods listed in Table 3.

Eriogonum umbellatum seeds did not separate from the flowering structures in the combine; the unthreshed seed was taken to the U.S. Forest Service Lucky Peak Nursery (Boise, ID) and run through a dewinger to separate seed. The seed was further cleaned in a small clipper seed cleaner.

Penstemon deustus seed pods were too hard to be opened in the combine; the unthreshed seed was precleaned in a small clipper seed cleaner and then seed pods were broken manually by rubbing the pods on a ribbed rubber mat. The seed was then cleaned again in the small clipper seed cleaner.

Penstemon acuminatus and *P. speciosus* were threshed in the combine and the seed was further cleaned using a small clipper seed cleaner.

Cultural Practices in 2006

On October 27, 2006, 50 lb phosphorus (P)/acre and 2 lb zinc (Zn)/acre were injected through the drip tape to all plots of *Eriogonum umbellatum*, *Penstemon* spp., and *Lomatium* spp. On November 11, 100 lb nitrogen (N)/acre as urea was broadcast to all *Lomatium* plots. On November 17, all plots of *Eriogonum umbellatum*, *Penstemon* spp. (except *P. deustus*), and *Lomatium* spp. had Prowl[®] at 1 lb ai/acre broadcast on the soil surface. Irrigations for all species were initiated on May 19 and terminated on June 30. Harvesting and seed cleaning methods for each species are listed in Table 3.

Cultural Practices in 2007

Penstemon acuminatus and *P. speciosus* were sprayed with Aza-Direct[®] at 0.0062 lb ai/acre on May 14 and May 29 for lygus bug control. All plots of *Sphaeralcea* were flailed on November 8, 2007. Irrigations for each species were initiated and terminated on different dates (Table 2a). Harvesting and seed cleaning methods for each species are listed in Table 3.

Cultural Practices in 2008

On November 9, 2007 and on April 15, 2008 Prowl at 1 lb ai/acre was broadcast on all plots for weed control.

Capture[®] 2EC at 0.1lb ai/acre was sprayed on all plots of *Penstemon acuminatus* and *P. speciosus* on May 20 for lygus bug control.

Irrigations for each species were initiated and terminated on different dates (Table 2). Harvesting and seed cleaning methods for each species are listed in Table 3.

Cultural Practices in 2009

On March 18, Prowl at 1 lb ai/acre and Volunteer® at 8 oz/acre were broadcast on all plots for weed control. On April 9, 50 lb N/acre and 10 lb P/acre were applied through the drip irrigation system to the three *Lomatium* species.

Irrigations for each species were initiated and terminated on different dates (Table 2b). Harvesting and seed cleaning methods for each species are listed in Table 3.

Table 2a. Native forb flowering, irrigation, and seed harvest dates by species in 2006, 2007, and 2008, Malheur Experiment Station, Oregon State University, Ontario, OR.

2007, and 2008, Manned Experiment Station, Oregon State University, Corvallis, OR.

Species	Flowering			Irrigation		Harvest
	start	peak	end	start	end	
2006						
<i>Eriogonum umbellatum</i>	19-May		20-Jul	19-May	30-Jun	3-Aug
<i>Penstemon acuminatus</i>	2-May	10-May	19-May	19-May	30-Jun	7-Jul
<i>Penstemon deustus</i>	10-May	19-May	30-May	19-May	30-Jun	4-Aug
<i>Penstemon speciosus</i>	10-May	19-May	30-May	19-May	30-Jun	13-Jul
<i>Lomatium dissectum</i>				19-May	30-Jun	
<i>Lomatium triternatum</i>				19-May	30-Jun	
<i>Lomatium grayi</i>				19-May	30-Jun	
<i>Sphaeralcea parvifolia</i>						
<i>S. grossulariifolia</i>						
<i>Sphaeralcea coccinea</i>						
<i>Dalea searlsiae</i>						
<i>Dalea ornata</i>						
2007						
<i>Eriogonum umbellatum</i>	25-May		25-Jul	2-May	24-Jun	31-Jul
<i>Penstemon acuminatus</i>	19-Apr		25-May	19-Apr	24-Jun	9-Jul
<i>Penstemon deustus</i>	5-May	25-May	25-Jun	19-Apr	24-Jun	
<i>Penstemon speciosus</i>	5-May	25-May	25-Jun	19-Apr	24-Jun	23-Jul
<i>Lomatium dissectum</i>				5-Apr	24-Jun	
<i>Lomatium triternatum</i>	25-Apr		1-Jun	5-Apr	24-Jun	29-Jun, 16-Jul
<i>Lomatium grayi</i>	5-Apr		10-May	5-Apr	24-Jun	30-May, 29-Jun
<i>Sphaeralcea parvifolia</i>	5-May	25-May		16-May	24-Jun	20-Jun, 10-Jul, 13-Aug
<i>S. grossulariifolia</i>	5-May	25-May		16-May	24-Jun	20-Jun, 10-Jul, 13-Aug
<i>Sphaeralcea coccinea</i>	5-May	25-May		16-May	24-Jun	20-Jun, 10-Jul, 13-Aug
<i>Dalea searlsiae</i>						20-Jun, 10-Jul
<i>Dalea ornata</i>						20-Jun, 10-Jul
2008						
<i>Eriogonum umbellatum</i>	5-Jun	19-Jun	20-Jul	15-May	24-Jun	24-Jul
<i>Penstemon acuminatus</i>	29-Apr		5-Jun	29-Apr	11-Jun	11-Jul
<i>Penstemon deustus</i>	5-May		20-Jun	29-Apr	11-Jun	
<i>Penstemon speciosus</i>	5-May		20-Jun	29-Apr	11-Jun	17-Jul
<i>Lomatium dissectum</i>				10-Apr	29-May	
<i>Lomatium triternatum</i>	25-Apr		5-Jun	10-Apr	29-May	3-Jul
<i>Lomatium grayi</i>	25-Mar		15-May	10-Apr	29-May	30-May, 19-Jun
<i>Sphaeralcea parvifolia</i>	5-May		15-Jun	15-May	24-Jun	21-Jul
<i>S. grossulariifolia</i>	5-May		15-Jun	15-May	24-Jun	21-Jul
<i>Sphaeralcea coccinea</i>	5-May		15-Jun	15-May	24-Jun	21-Jul
<i>Dalea searlsiae</i>		19-Jun				
<i>Dalea ornata</i>		19-Jun				

Results and Discussion

The soil volumetric water content responded to the irrigation treatments (Figs. 1-6).

Flowering and Seed Set

Emergence for the two prairie clover (*Dalea*) species in the spring of 2007 was again poor. Emergence for *Penstemon deustus* and *Astragalus filipes* was extremely poor; *A. filipes* produced negligible amounts of seed in 2007.

Penstemon acuminatus and *P. speciosus* had poor seed set in 2007, partly due to a heavy lygus bug infestation that was not adequately controlled by the applied insecticides. In the Treasure Valley, the first hatch of lygus bugs occurs when 250 degree days (52°F base) are accumulated. Data collected by an AgriMet weather station adjacent to the field indicate that the first lygus bug hatch occurred on May 14 in 2006, on May 1 in 2007, and on May 18 in 2008. *Penstemon acuminatus* and *P. speciosus* start flowering in early May. The earlier lygus bug hatch in 2007 probably resulted in harmful levels of lygus bugs present during a larger part of the *Penstemon* flowering period than in 2006 or 2008. Poor seed set for *P. acuminatus* and *P. speciosus* in 2007 also was related to poor vegetative growth compared to 2006 and 2008. In 2009, all plots of *P. acuminatus* and *P. speciosus* again showed poor vegetative growth and seed set. Root rot affected all plots of *P. acuminatus* in 2009, killing all plants in two of the four plots of the wettest treatment (2 inches per irrigation).

The three *Sphaeralcea* species (globemallow) showed a long flowering period (early May through September) in 2007. Multiple harvests were necessary because the seed falls out of the capsules once they are mature. The flailing of the three *Sphaeralcea* species in the fall of 2007 and 2008 resulted in a more concentrated flowering in 2008 and 2009, which allowed one mechanical harvest. Precipitation in June of 2009 (2.27 inches) was substantially higher than average (0.76 inches). A rust infected all plots of the *Sphaeralcea* species in June, causing substantial leaf loss and reduced vegetative growth.

Seed Yields

In 2006, seed yield of *Eriogonum umbellatum* increased with increasing water application, up to 8 inches, the highest amount tested (Table 4, Fig. 7). In 2007-2009 seed yield showed a quadratic response to irrigation rate. Seed yields were maximized by 8.1 inches, 7.2 inches, and 6.9 inches of water applied in 2007, 2008, and 2009, respectively. Averaged over 4 years, seed yield of *E. umbellatum* increased with increasing water applied up to 8 inches, the highest amount tested (Fig. 7).

There was no significant difference in seed yield between irrigation treatments for *Penstemon acuminatus* in 2006 (Table 4). Precipitation from March through June was 6.4 inches in 2006. The 64-year average precipitation from March through June is 3.6 inches. The wet weather in 2006 could have attenuated the effects of the irrigation treatments. In 2007 and 2008, seed yield showed a quadratic response to irrigation

Table 4. Native forb seed yield response to irrigation rate (inches/season) in 2006-2009. Malheur Experiment Station, Oregon State University, Ontario, OR, 2009.

Species	2006				2007				2008				2009			
	0 inches	4 inches	8 inches	LSD (0.05)	0 inches	4 inches	8 inches	LSD (0.05)	0 inches	4 inches	8 inches	LSD (0.05)	0 inches	4 inches	8 inches	LSD (0.05)
	----- lb/acre -----															
<i>Eriogonum umbellatum</i> ^a	155.3	214.4	371.6	92.9	79.6	164.8	193.8	79.8	121.3	221.5	245.2	51.7	132.3	223.0	240.1	67.4
<i>Penstemon acuminatus</i> ^a	538.4	611.1	544.0	NS	19.3	50.1	19.1	25.5 ^b	56.2	150.7	187.1	79.0	20.7	12.5	11.6	NS
<i>Penstemon deustus</i> ^c	1246 ^d	1201 ^d	1069 ^d	NS	120.3	187.7	148.3	NS	---- very poor stand ----				---- very poor stand ----			
<i>Penstemon speciosus</i> ^a	163.5	346.2	213.6	134.3	2.5	9.3	5.3	4.7 ^b	94.0	367.0	276.5	179.6	6.8	16.1	9.0	6.0 ^b
<i>Lomatium dissectum</i> ^e	---- no flowering ----				---- no flowering ----				-- very little flowering --				66.0	322.8	431.4	233.7 ^b
<i>Lomatium triternatum</i> ^e	---- no flowering ----				2.3	17.5	26.7	16.9 ^b	195.3	1060.9	1386.9	410.0	181.6	780.1	676.1	177.0
<i>Lomatium grayi</i> ^e	---- no flowering ----				36.1	88.3	131.9	77.7 ^b	393.3	1287.0	1444.9	141.0	359.9	579.8	686.5	208.4
<i>Sphaeralcea parvifolia</i> ^f					1062.6	850.7	957.9	NS	436.2	569.1	544.7	NS	285.9	406.1	433.3	NS
<i>Sphaeralcea grossularifolia</i> ^f					442.6	324.8	351.9	NS	275.3	183.3	178.7	NS	270.7	298.9	327.0	NS
<i>Sphaeralcea coccinea</i> ^f					279.8	262.1	310.3	NS	298.7	304.1	205.2	NS	332.2	172.1	263.3	NS
<i>Dalea searlsiae</i> ^f					11.5	10.2	16.4	NS	---- very poor stand ----				---- very poor stand ----			
<i>Dalea ornata</i> ^f					47.4	27.3	55.6	NS	---- very poor stand ----				---- very poor stand ----			

^a planted March, 2005, areas of low stand replanted by hand in October 2005.

^b LSD (0.10).

^c planted March, 2005, areas of low stand replanted by hand in October 2005 and whole area replanted in October 2006. Yields in 2006 are based on small areas with adequate stand. Yields in 2007 are based on whole area of very poor and uneven stand.

^d based on small areas with good plant stands in 2006.

^e planted March, 2005, whole area replanted in October 2005.

^f planted spring 2006, whole area replanted in November 2006.

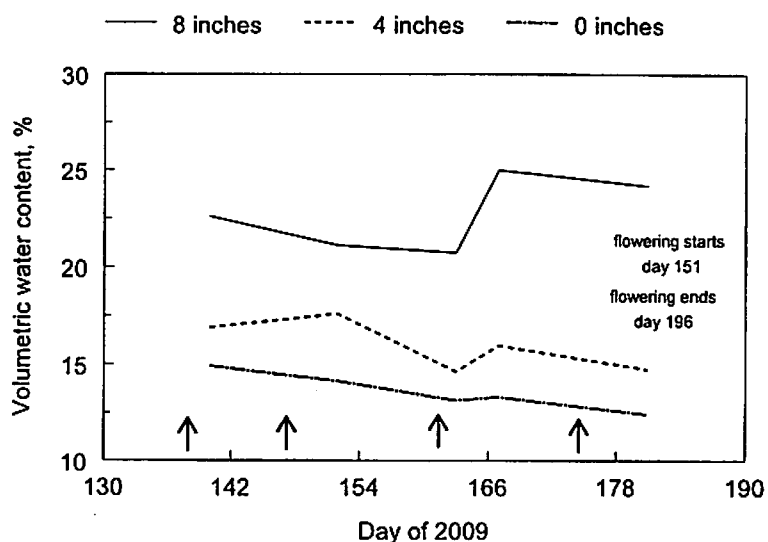


Figure 1. Soil volumetric water content for *Eriogonum umbellatum* over time in 2009. Soil volumetric water content is the combined average at the 8-, 20-, and 32-inch depths. Irrigations started on May 19 and ended on June 24. Arrows denote irrigations. *E. umbellatum* was harvested on July 28 (day 209). Malheur Experiment Station, Oregon State University, Ontario, OR, 2009.

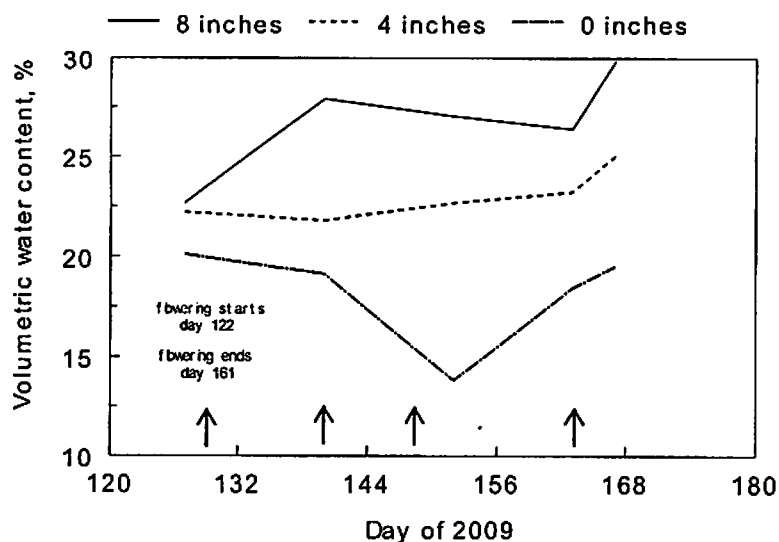


Figure 2. Soil volumetric water content for *Penstemon acuminatus* over time in 2009. Soil volumetric water content is the combined average at the 8-, 20-, and 32-inch depths. Irrigations started on May 8 and ended on June 12. Arrows denote irrigations. *P. acuminatus* was harvested on July 10 (day 191). Malheur Experiment Station, Oregon State University, Ontario, OR, 2009.

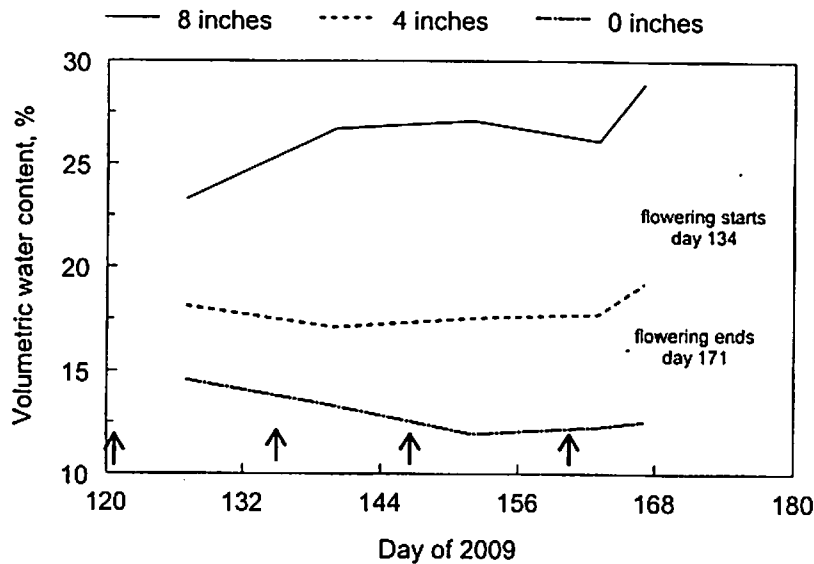


Figure 3. Soil volumetric water content for *Penstemon speciosus* over time in 2009. Soil volumetric water content is the combined average at the 8-, 20-, and 32-inch depths. Irrigations started on May 19 and ended on June 24. Arrows denote irrigations. *P. speciosus* was harvested on July 10 (day 191). Malheur Experiment Station, Oregon State University, Ontario, OR, 2009.

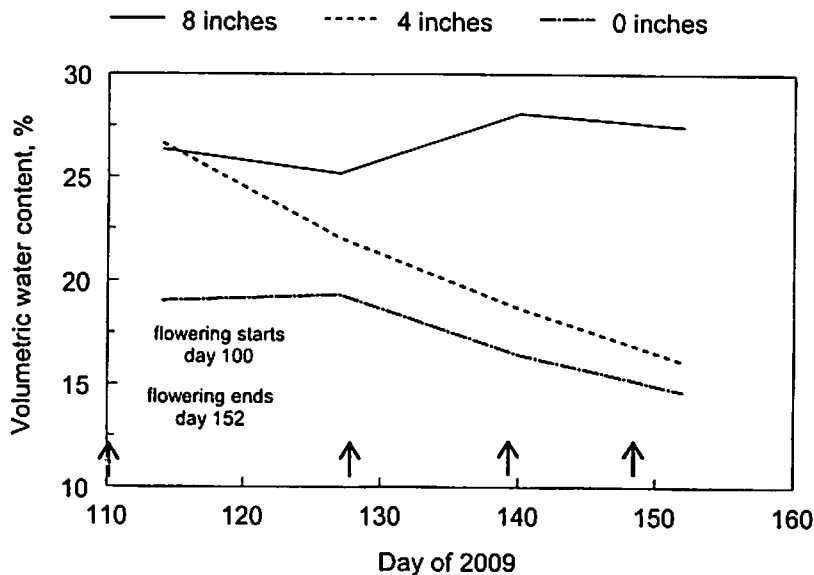


Figure 4. Soil volumetric water content for *Lomatium triternatum* over time in 2009. Soil volumetric water content is the combined average at the 8-, 20-, and 32-inch depths. Irrigations started on April 20 and ended on May 28. Arrows denote irrigations. *L. triternatum* was harvested on June 26 (day 177). Malheur Experiment Station, Oregon State University, Ontario, OR, 2009.

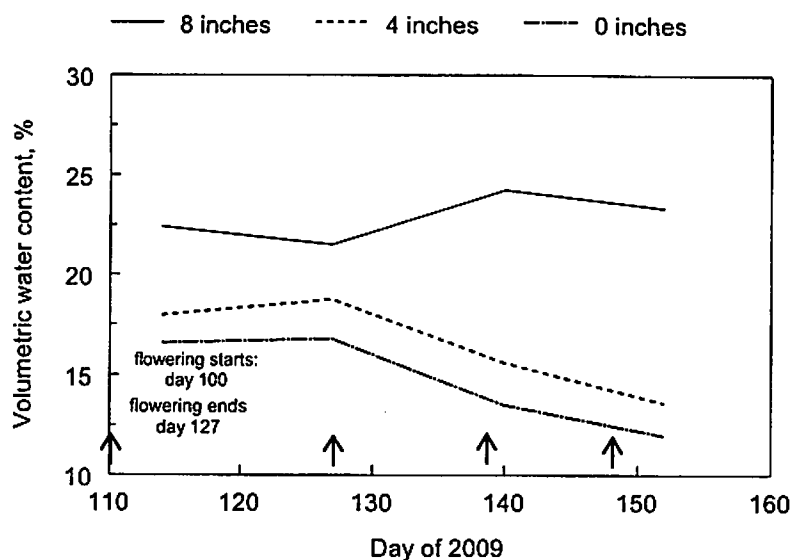


Figure 5. Soil volumetric water content for *Lomatium grayi* over time in 2009. Soil volumetric water content is the combined average at the 8-, 20-, and 32-inch depths. Irrigations started on April 20 and ended on May 28. Arrows denote irrigations. *L. grayi* was harvested on June 16 (day 167). Malheur Experiment Station, Oregon State University, Ontario, OR, 2009.

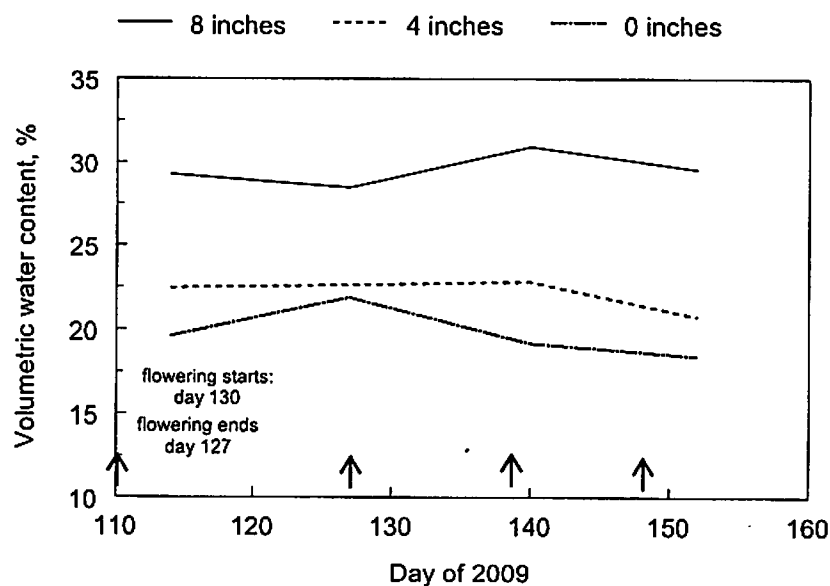


Figure 6. Soil volumetric water content for *Lomatium dissectum* over time in 2009. Soil volumetric water content is the combined average at the 8-, 20-, and 32-inch depths. Irrigations started on April 20 and ended on May 28. Arrows denote irrigations. *L. dissectum* was harvested on June 16 (day 167). Malheur Experiment Station, Oregon State University, Ontario, OR, 2009.

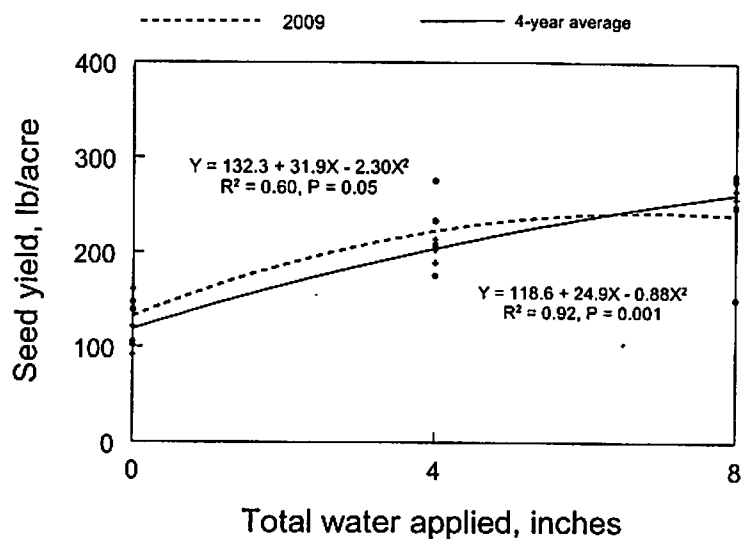


Figure 7. Average annual *Eriogonum umbellatum* seed yield response to irrigation water applied in 2009 and averaged over 4 years, Malheur Experiment Station, Oregon State University, Ontario, OR, 2009.

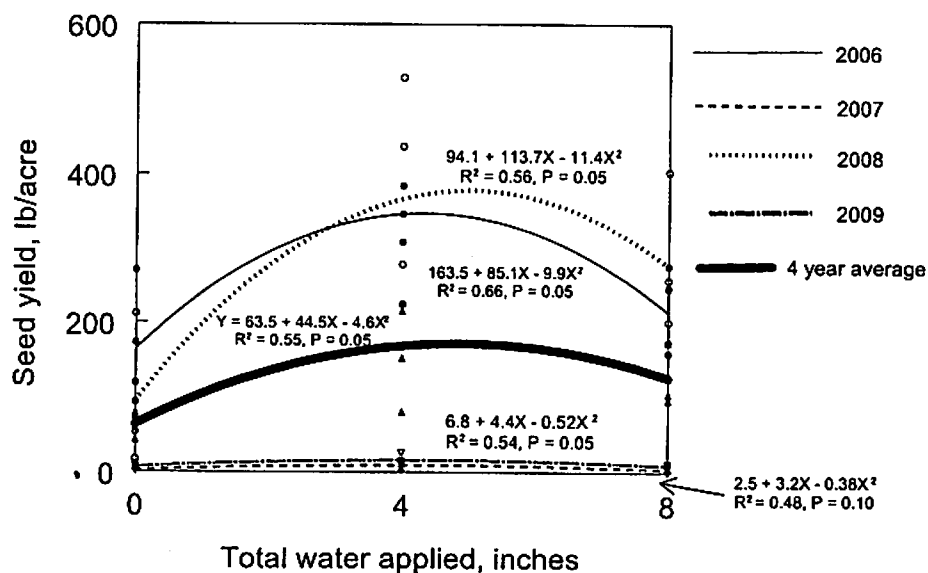


Figure 8. *Penstemon speciosus* seed yield response to irrigation water applied in 2009 and averaged over 4 years, Malheur Experiment Station, Oregon State University, Ontario, OR, 2009.

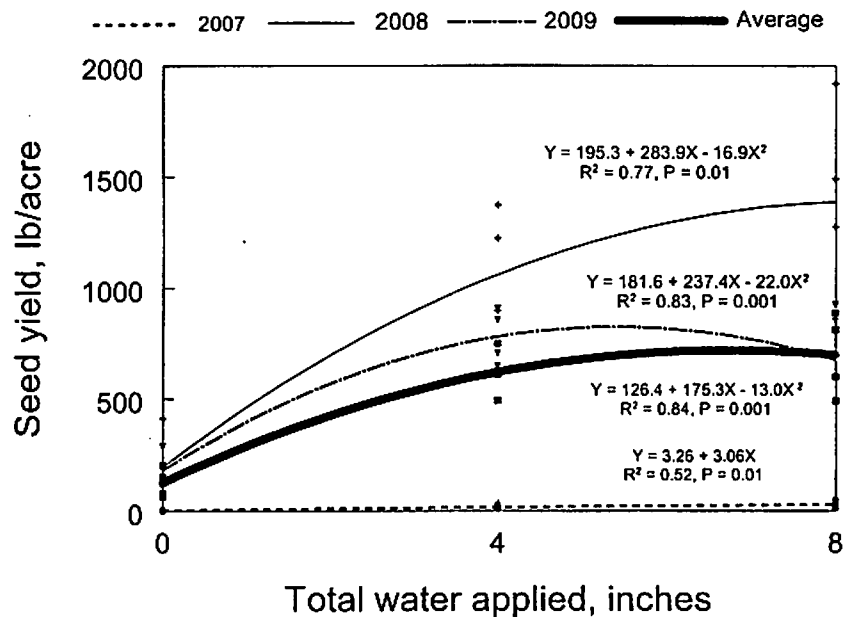


Figure 9. Annual and 3-year average *Lomatium triternatum* seed yield response to irrigation water applied, Malheur Experiment Station, Oregon State University, Ontario, OR, 2009.

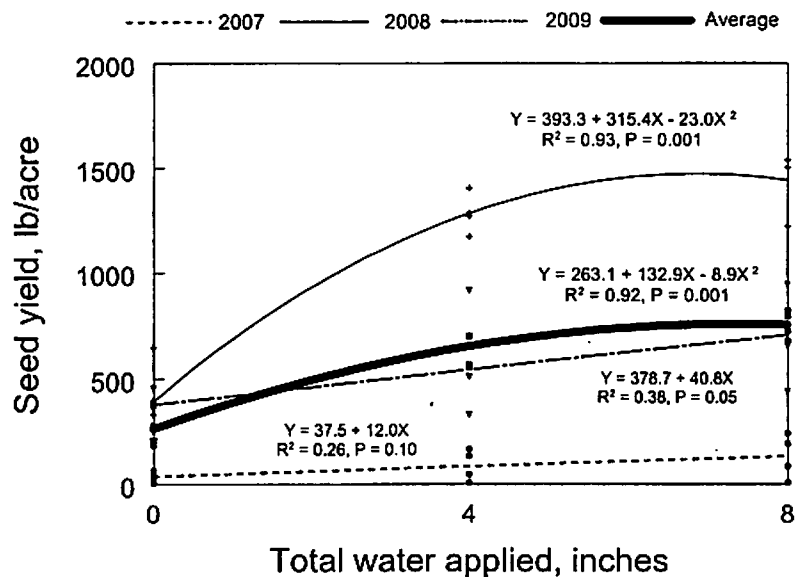


Figure 10. Annual and 3-year average *Lomatium grayi* seed yield response to irrigation water applied, Malheur Experiment Station, Oregon State University, Ontario OR, 2009.

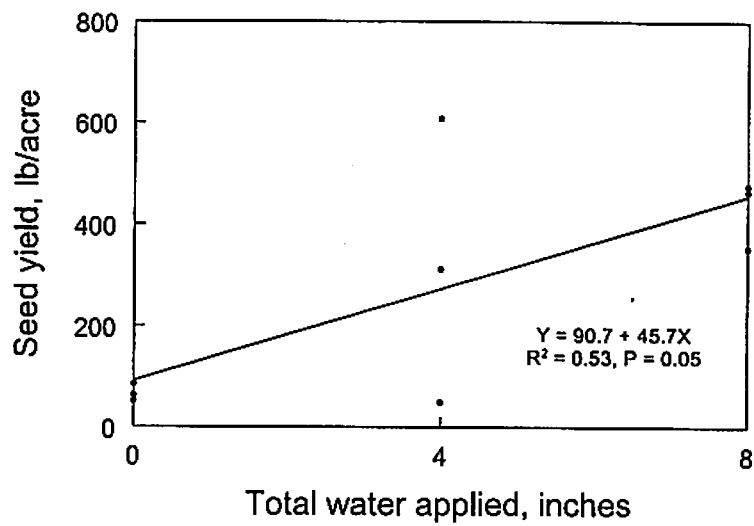


Figure 11. *Lomatium dissectum* seed yield response to irrigation water applied in 2009, Malheur Experiment Station, Oregon State University, Ontario OR, 2009.