

Lichen elemental content bioindicators for air quality in upper Midwest, USA: A model for large-scale monitoring



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ABSTRACT

Our development of lichen elemental bioindicators for a United States of America (USA) national monitoring program is a useful model for other large-scale programs. Concentrations of 20 elements were measured, validated, and analyzed for 203 samples of five common lichen species. Collections were made by trained non-specialists near 75 permanent plots and an expert near nine air monitoring sites. *Flavoparmelia caperata* (most frequent) and *Physcia aiopolia/stellaris* between them represented the full range of local forest cover and pollution load. *Evernia mesomorpha* (values saturated at intermediate pollution), *Parmelia sulcata*, and *Punctelia rudecta* (both difficult for non-specialists) were less useful. Conversion models (GLM or regression) rendered elemental data equivalent between species. Al, Cr, Cu, Fe, Hg, N, and S, plus composite indexes from them, were linked with local air pollution based on correlations with directly measured N and particulate matter as well as from PCA; elements were weakly correlated with modeled pollution estimates. Lichen Hg had no other useful surrogates. Invoking multiple causation and scale-dependence helped address several issues of interpretation, for instance conflicting bioindicator value of Al and Fe from literature.

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1. Introduction

Biomonitoring response to air pollution is an important component of evaluating the health of biological systems (e.g., for lichens Fenn et al., 2003; Nimis et al., 2002). Estimation of local air quality from elemental concentrations in lichens or mosses naturally growing *in situ* (passive monitoring as opposed to active monitoring with transplants: Garty, 2002) is a classic technique (Ferry et al., 1973; Martin and Coughtrey, 1982) with wide current usage (e.g. Donovan et al., 2016; McMurray et al., 2013; Paoli et al., 2014) to complement costly instrument monitoring.

Elemental biomonitoring well represents relative local pollution loads, as compared with instrument measurements (e.g. Bargagli and Mikhailova, 2002; Root et al., 2015), although accuracy of quantitative calibration varies (e.g. Bargagli, 2016; Bari et al., 2001; Boquete et al., 2015; Cercasov et al., 2002; Vestergaard et al., 1986). Elemental bioindicator species should be relatively

pollution-tolerant, widespread, and amenable to collection (e.g. Conti and Cecchetti, 2001; Puckett, 1988; Wolterbeek, 2002). Elemental accumulation rates can differ between lichen species; conversion between species (e.g. Karakas and Tuncel, 2004; Root et al., 2013; Sloof and Wolterbeek, 1993) can provide equivalent elemental data.

Our protocols were designed for large-scale studies where multiple target species are likely and simplicity plus cost-effectiveness facilitate implementation. Our general objective for the full project (located in upper Midwest, USA) was to recommend improved practices for elemental biomonitoring using *in-situ* mosses or lichens. Our objectives for this aspect of the full project were to: 1) evaluate elemental concentrations in multiple lichen species from non-specialist collectors; 2) develop conversion models between species; 3) calibrate lichen elemental data to measured values; and 4) evaluate lichen-indicated air quality across the study region. In two other aspects of this project, Will-Wolf et al. (2017a) assess the impact of variation in protocols and staff expertise on data quality, and Will-Wolf et al. (2017b) evaluate applications of protocols and bioindicators in eastern United States of America (E USA) for the United States Department of Agriculture Forest Service Forest Inventory and Analysis Program (FIA) program. Lichen elemental

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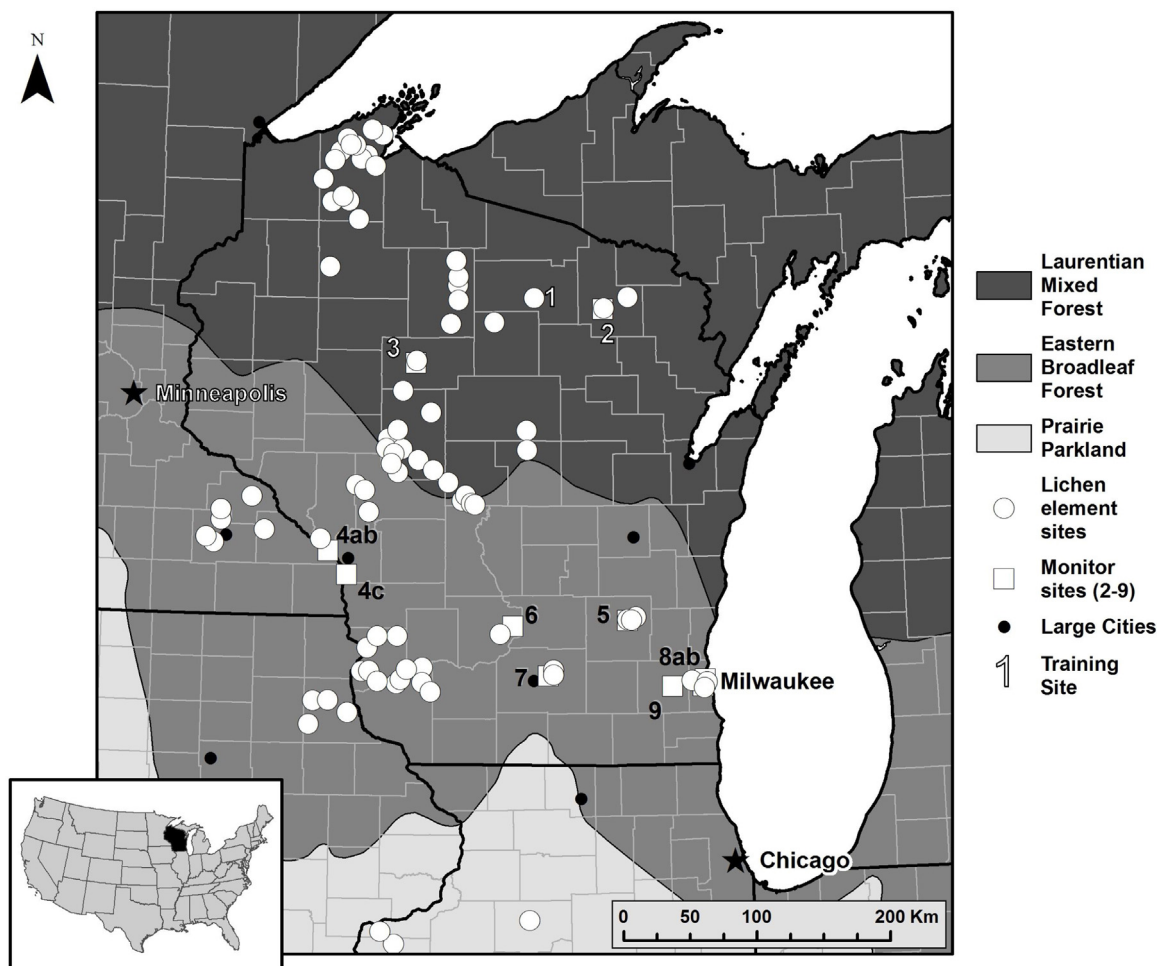


Fig. 1. Locations of lichen sites and instrument monitor sites. Wisconsin, marked in the USA inset, is in the center with Minnesota west, Iowa southwest, and Illinois south.

bioindicators for two other E USA projects (Will-Wolf et al., 2014, 2015b) used similar protocols, but only expert staff.

2. Methods

2.1. Study region and environmental variables

The 241,800 km² project area includes USA states Wisconsin and parts of adjacent Illinois, Iowa, and Minnesota (Fig. 1). It mostly extends across two Bailey ecoregions (Bailey et al., 1994; Cleland et al., 2007): Laurentian Mixed Forest (province 212) and Eastern Broadleaf

Forest (province 222); three southern (Illinois) plots are in Prairie Parkland (province 251). Laurentian Mixed Forest has high cover of mixed deciduous broadleaf-conifer forests; Eastern Broadleaf Forest has more fragmented deciduous broadleaf forests interspersed with agricultural land; Prairie Parkland now has sparse deciduous broadleaf forests in an agricultural landscape. Environmental variables represent location, climate, land cover, and modeled pollution deposition (Table 1; Supplementary Document 1). Lichen elemental concentrations and species distributions were correlated with land cover in a subset of the project area (Will-Wolf et al. 2011, 2015b); similar links were investigated at this larger spatial scale. Measured pollutants for 2008–2013 were from 11 instrument monitoring stations at eight monitor sites (2–9 on Fig. 1) linked with national networks (Supplementary Document 2).

2.2. Target species, field data collection, and measurement

Five macrolichen species common in eastern North America (E NA: Brodo et al., 2001) were collected: *Evernia mesomorpha* Nyl. (acronym Evemes; small/medium size fruticose growth form), *Flavoparmelia caperata* (L.) Hale (acronym Flacap: large foliose), *Parmelia sulcata* Taylor (acronym Parsul; medium foliose), *Physcia aipolia* (Ehrh. ex Humb.) Fűrnr. var. *aipolia* and *P. stellaris* (L.) Nyl. combined (acronym Physaip; small foliose, tightly appressed), and *Punctelia rudecta* (Ach.) Krog (acronym Punrud; large foliose). Flacap and Punrud were used in two recent E USA studies (Will-Wolf et al., 2014, 2015b) and all but Physaip have been used in other studies in the region (Will-Wolf et al., 2017a).

Lichens were collected by FIA field staff with no prior experience with lichens, who survey permanent plots on a preset 10-year rotation (USDA FS, 2015), and by professional lichenologist Susan Will-Wolf (the expert). Field staff were trained (at site 1, Fig. 1) for one day by the expert (details in Will-Wolf et al., 2017b). Field staff collected lichens September–November 2013; the expert collected in Summer–Fall 2013 from temporary sites near monitor sites 2–9 on Fig. 1 (Supplementary Document 2).

Collection areas were forest openings or edges with 20–50% canopy near permanent FIA plots (no destructive sampling on plots: USDA FS, 2015) or at temporary sites with similar characteristics. The moderately narrow range of relatively open canopy reduced habitat variability (e.g. Gandois et al., 2014), allowed penetration of air pollution, and still represented forest conditions. Areas affected by local chemical contamination (e.g. adjacent to active agricultural

Table 1

Site characteristics by ecoregion; most useful environmental variables. Variables in bold had the strongest relationships with lichen elemental data.

Environmental Variable	Laurentian Mixed Forest		Eastern Broadleaf Forest		Prairie Parkland	
	N = 40		N = 40		N = 3	
	avg	range	avg	range	avg	range
Geography, Climate						
Latitude, deg N	45.66	44.32–46.77	43.55	42.77–44.53	41.37	41.28–41.45
Longitude, deg W	–90.63	–91.51 to –88.57	–90.59	–92.55 to –87.90	–90.40	–90.89 to –89.54
Elevation, m	1227.90	442–1732	820.69	190–1200	740.33	713–785
Min annual temp, deg C	–0.55	–1.49 to 0.93	2.15	0.22–4.49	4.64	4.27–4.97
Max annual temp, deg C	11.30	9.93–13	13.24	12.4–14.4	15.58	15.53–15.66
Mean annual temp, deg C	5.38	4.27–6.93	7.70	6.34–8.99	10.11	9.91–10.32
Mean annual ppt, mm	822.80	773.4–897.6	870.34	830.7–943.6	932.27	930.8–933.1
Land cover, %						
Forest Cover	88.35	48.4–100	46.16	0.8–95.6	20.54	7.8–36.2
Agricultural Cover	5.80	0–50.7	30.83	0–81.5	74.23	58.5–87.9
Developed Land Cover	3.30	0–22.1	16.35	0.73–99.0	4.15	3.30–5.09
Modeled air pollution, kg/ha/yr						
SO₄wet deposition	6.33	5.47–8.55	7.81	6.10–9.96	7.95	7.51–8.78
Hg wet deposition	8.63	7.41–10.23	11.11	9.20–12.95	10.45	10.3–10.7
S total deposition	3.68	3.16–4.45	4.79	3.65–7.58	5.51	5.42–5.62
N total deposition	10.32	8.08–16.30	14.42	7.83–19.43	11.76	11.4–12.5
Pollution Indices						
Lichen N + S	1.09	0.51–2.54	1.63	0.97–2.64	1.22	0.44–2.10
Lichen 5 metals	1.28	0.66–2.65	1.45	0.89–2.50	1.00	0.63–1.63

fields, roads, or lawns) were avoided. Target species were collected from any natural standing (live or dead) woody stem or branch 0.5 m above the ground and within reach.

The field staff collection goal was two samples at each site; replicates of multiple species were collected at intervals. The expert collected replicate samples of each target species present at a site, plus a bulk Flacap sample for internal reference (Will-Wolf et al., 2017b). Samples for this study followed rigorous collecting and handling protocols (Will-Wolf et al., 2017a) to avoid contamination. Adequate samples included ≥ 1 gr air dry weight for one species, appearing healthy, and composited from ≥ 6 different substrates within a 200–500 m range (averages across within-site variation). Samples were handled in the field and lab (by the expert) wearing clean nitrile gloves, collected into certified chemically clean bags, air-dried, and kept cool and dry before measurement. Equipment was wiped with alcohol. Sample elements (listed by standard codes: IUPAC, 2014), plus a certified external standard (IAEA-336 pre-ground *Evernia prunastri* from rural Portugal: IAEA, 2014) and the internal reference, were measured at the Soil and Water Analysis Lab, Logan Forestry Sciences Lab, USFS Rocky Mountain Research Station, Logan, Utah, USA. C, N, S, and Hg were measured with combustion analysis, while 23 elements were measured by chemical digestion and ICP-OES (Supplementary Document 3).

2.3. Elemental data validation and conversion between species

Validation of elemental data was based on values below detection limit (BDL) and variability of replicates. Extreme outlier values ($>5\times$ the next higher value) were removed before validation (measurement error assumed). Replicate variability was calculated as relative standard deviation (rSD: $100 \times \text{standard deviation} \sigma / \text{mean } \mu$; e.g. Halleraker et al., 1998), equivalent to coefficient of variation (CV) when μ is positive. Average rSD was calculated for replicates of the external standard, the Flacap internal reference, Flacap from field sites, and post-grinding lab splits of all species. Validated elements had $<5\%$ of values BDL and average rSD $\leq 25\%$ for all sets of replicates. Cut-offs in the literature for data reliability are 10–50% rSD or CV (e.g. Frati et al., 2005; Gailey and Lloyd, 1986; Loppi et al., 2002). For validated elements, BDL values were replaced by $\frac{1}{2}$ the smallest accurately detected value.

Conversion of each species' data to equivalence with Flacap used data from all sites having both species. Linear regression of species averages for sites (Flacap the independent variable) and General Linear Model (GLM) univariate analysis of sample values with species and site as fixed independent factors were calculated in SPSS v23.0 (SPSS, 2015). Logarithm base 10 transformed ($\log_{10}$) data corrected for data distribution issues as necessary. Model strength was judged from homogeneity of variances (GLM) and probability (p) values. Flacap data violated the regression model assumption of no measurement error for the independent variable, so a GLM model (measurement error assumed) was selected when p values were similar, and a regression model with $p > 0.005$ was described as weak, in case measurement error of Flacap had inflated model strength. Models from regression and/or $\log_{10}$ data with notably lower p values were interpreted as indicating the relationship was non-linear. The second species' elemental concentrations were converted to equivalence with Flacap values using the strongest model. Site averages were calculated from converted data.

2.4. Elemental data analysis

Calibration of expert samples to measured data from air monitoring sites (2–9, Fig. 1) used monitor site variables for particulate matter (PM) and monitored elements. Each was often measured in different ways, for example both “Cadmium PM (particulate matter) 2.5 (particle size in microns) LC (local conditions)” and Cadmium PM10 LC might be measured. For the most frequent variable both measured and ranked values were tested. Values were also ranked for each type of variable for each element or compound; ranks were averaged for each monitor site to increase sample size (Supplementary Document 2). We then correlated (SPSS, 2015) composite monitor site variables with lichen elemental data from nearby sites.

Patterns of association between validated elements were explored with principal components analysis (PCA) and pairwise correlations. PCA of average site values for each element was conducted in PC-ORD v.6 (McCune and Mefford 2016); outcome was compared with randomized runs. Pairwise correlations between elements by sample or site, plus simple and partial correlations (SPSS, 2015) with environmental variables, also suggested sources

Table 2
Summary of models for conversion of data from other lichen species to equivalence with data from Flacap (total number of sites with both species in parentheses). Original data were used unless noted. Abbreviations: excl. = excluded; GLM = univariate general linear model; regress = regression model; wk = model p value $0.01 < p < 0.05$.

Element	Evemes to Flacap (N = 13)	Parsul to Flacap (N = 10)	Phyaip to Flacap (N = 14)	Punrud to Flacap (n = 13)
Al	GLM, 3 sites excl.	GLM, log10 data	Regress, 1 site excl.	Regress, 2 sites excl.
CC	GLM wk, 2 sites excl.	GLM	GLM	NOT NEEDED (GLM), 2 sites excl.
Ca	GLM, 2 sites excl.	GLM	GLM	GLM, log10 data, 3 sites excl.
Cd	GLM, log10 data, 2 sites excl.	Regress, log10 data	GLM, log10 data	NOT NEEDED (GLM), 2 sites excl.
Co	Regress, 2 sites excl.	GLM, log10 data	NOT NEEDED (GLM)	NOT NEEDED (GLM), 2 sites excl.
Cr	GLM, 2 sites excl.	GLM, log10 data	GLM, log10 data	Regress, 2 sites excl.
Cu	NOT NEEDED (GLM), log10 data, 4 sites excl.	GLM, log10 data	Regress	GLM, 2 sites excl.
Fe	GLM, 2 sites excl.	GLM	Regress, 1 site excl.	Regress, 3 sites excl.
Hg	GLM, 2 sites excl.	GLM	GLM	GLM, log10 data, 2 sites excl.
K	GLM, 2 sites excl.	GLM, log10 data	GLM, L10 data, 1 site excl.	GLM, 2 sites excl.
Mg	Regress	Regress	GLM, log10 data	Regress, 2 sites excl.
Mn	Regress	Regress	Regress, 1 site excl.	Regress wk, log10 data, 2 sites excl.
N	GLM, 2 sites excl.	Regress	Regress wk, log10 data, 1 site excl.	Regress, 2 sites excl.
Na	GLM wk, log10 data, 2 sites excl.	Regress	NOT NEEDED (GLM)	DO NOT CONVERT
Ni	GLM wk, L10 data, 2 sites excl.	GLM wk	NOT NEEDED (GLM), 1 site excl.	Regress, log10 data, 2 sites excl.
P	GLM wk, log10 data, 2 sites excl.	GLM, log10 data	GLM, L10 data	NOT NEEDED (GLM), 3 sites excl.
Pb	GLM, log10 data, 2 sites excl.	NOT NEEDED (GLM)	GLM, log10 data	Regress, 2 sites excl.
S	Regress, 2 sites excl.	GLM	GLM	NOT NEEDED (GLM), 3 sites excl.
Sr	GLM wk, 2 sites excl.	GLM, log10 data	GLM, log10 data	NOT NEEDED (GLM), 2 sites excl.
Zn	Regress wk, 3 sites excl.	GLM, log10 data	GLM wk, log10 data	Regress, 2 sites excl.
Data pattern	Nonlinear for 55% of elements	Nonlinear for 65% of elements	Nonlinear for 60% of elements	Nonlinear for 55% of elements

or arrival modes. Many statistical tests were performed but elements and variables were often correlated with each other. To qualitatively account for experiment-wide error, any result with p value $0.01 < p < 0.05$ was considered weak (includes species conversion models described in section 2.3)

3. Results

3.1. Elemental data collection and validation

In total 231 field samples were collected; 221 were measured for 26 elements in spring 2014. Field staff collected 146 samples near 70 of 81 permanent plots visited (86%) and 75 searched (93%), with two or more target species near 52 (74%) searched plots (ten not measured: only non-target species or too small). An additional 75 samples were collected and measured by field staff from the training site or by the expert at 13 temporary lichen sites near eight monitor sites. Plots equally represented the two most prevalent ecoregions and spanned the full (NE–SW) climate gradient and a long pollution gradient (Table 1), though they were unevenly spaced (Fig. 1). About 85% of 30 individuals of sampled Phyaip from five sites were assigned to *Physcia aipolia*; ~15% (from three of those sites) were assigned to *P. stellaris*. Vouchers for all species are deposited in WIS. Twenty elements were validated for analysis (Table 2). Six (As, B, Ba, Mo, Se, Si) were excluded, with >5% BDL and/or rSD > 25% from 7 to 8 replicates of the standard and/or internal reference. Average rSD for field replicates or lab splits confirmed those stronger results (Supplementary Document 3). S from digestion/ICP–OES was used for analyses; it had smaller rSD for field replicates and more sites with data than S from combustion.

This study analyzed data for 203 measured samples from 83 sites (original data and information on sites in Will-Wolf et al. (2017b), Will-Wolf et al. (2017a) excluded 18 samples from analyses because of poor quality validated data for several elements. Most excluded samples also had field sample quality issues: sample composited from too few substrates, very small, or with external contamination, including a novel possible soil dust signal from high Ca/Sr and low values of pollution elements (Supplementary Document 4, sections S4.1, S4.3).

3.2. Conversion of lichen species data and calculation of pollution indices

Most species and elements required data conversion for equivalence with Flacap (Table 2; model parameters and formulas in Supplementary Document 5). Relationships between species appeared nonlinear (regression and/or log10 data gave the strongest model) for most elements. The strongest model (of up to eight) had the lowest p value, the most homogeneous variances, and the weakest site $\times$ species interaction (GLM results). Conversion was from the difference between Estimated Marginal Means (EMMs) for a GLM model or the linear regression model formula applied to the second species. For Evemes, Phyaip, and Punrud a few of the sites with both species were excluded to achieve a stable conversion model: the most polluted Evemes sites excluded for most elements, a Phyaip site excluded for five elements, two to three moderately polluted Punrud sites excluded for most elements. No site was excluded for more than one species pair. Evemes pollution tolerance was exceeded at excluded sites; Phyaip and Punrud unexplained anomalous data drove site exclusion (Supplementary Document 5). Data excluded for conversion models were excluded for other analyses. Ranges of site averages with converted data were similar to Flacap alone (Supplementary Document 6). Within-site variability was higher for converted data, but exceeded 25% of average only for Cd, Mn, Na, Ni, Pb, Sr, and Zn. Range of within-site replicates after conversion was mostly <10% of full range.

Two combined pollution indices, one from lichen N + S, and one from lichen Al + Co + Cr + Cu + Fe, were calculated for each lichen site after data conversion, to serve as general indicators for relative local pollution load and to compensate for missing data on individual elements. Site values for other elements were rescaled to N values with a multiplier: average (across all lichen sites) elemental value/average N value (Table 3); rescaled elemental ranges were also similar to N. Rescaled values for each element were averaged for the site Index score. The Indices, while moderately correlated with each other ($r^2 = 0.465$), reflected somewhat different pollution patterns in the upper range of both Indices (scatter in Fig. 2).

Example scatterplots of elements by species (other scatterplots in Will-Wolf et al., 2017b) illustrate species differences from original data (Fig. 3, left graph), and interspersions of species after conversion (Fig. 3, right graphs). An element is graphed vs the Pollution Index with which it was more strongly correlated. Flacap

Table 3
Summary of calculations for combined lichen pollution indices.

	Original Site Values				Transformed Site Values			
	Avg	Max	Min	Multiplier	Avg	Max	Min	
Al	309.83	692.93	123.70	0.0044	1.350	3.019	0.5389	
Co	0.2121	0.4125	0.115	6.365	1.350	2.626	0.7320	
Cr	0.6334	2.1996	0.2820	2.131	1.350	4.687	0.6010	
Cu	3.127	7.402	1.222	0.4317	1.350	3.195	0.5272	
Fe	393.47	949.17	137.76	0.0034	1.350	3.256	0.4726	
N	1.350	2.645	0.4722	1.0	1.350	2.645	0.4722	
S	0.1190	0.2333	0.0392	11.342	1.350	2.646	0.4450	
Lichen Pollution Index, N + S				–	1.354	2.639	0.4429	
Lichen Pollution Index, 5 metals				–	1.350	2.650	0.6349	

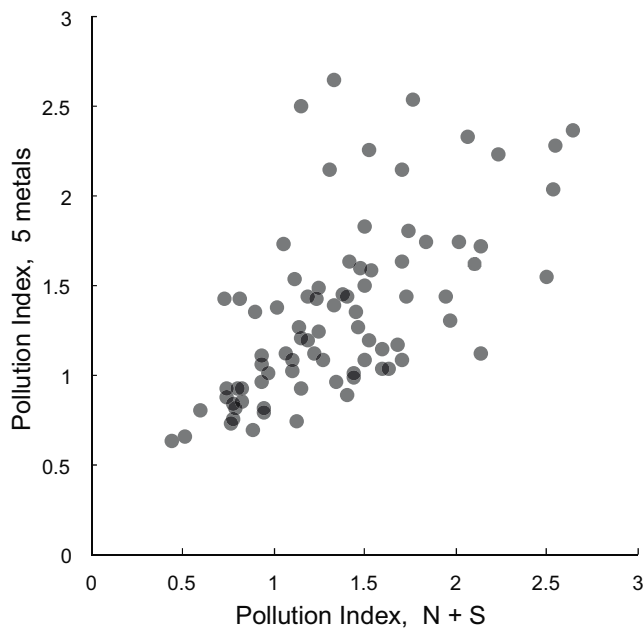


Fig. 2. Comparison of the two lichen elemental Pollution Indices. The "...5 metals" Index includes Al, Co, Cr, Cu, and Fe.

and Phyaip samples together spanned the full pollution range; only three sites lacked both species. For most species S (Fig. 3A, B), Cr (Fig. 3G, H), and Cu (Fig. 3I, J) elemental values increased with Pollution Index, a pattern also shown by Al, Co, Fe, and N. In contrast, original data for Evemes S, Cr, and Cu (Fig. 3A, G, I) leveled off at Pollution Index 1–1.2 and Evemes was absent from the upper Index ranges; pattern was similar for Al, Co, Fe, and N. Tight clustering of S, Cr, and Cu values partly reflects inclusion in their Index; wide scatter of Hg and Ca data reflects weak correlations with the Index (Supplementary Document 7). Original S (Fig. 3A) and Cr (Fig. 3G) values for species were similar at the same Index score, a pattern shared by Al, C, Fe, and N. Original values for species were more distinct for Hg (Fig. 3C) and Cu (Fig. 3I), a pattern shared by Cd, Co, K, Mg, Mn, Na, Ni, P, and Zn. Overlapping original values of Ca (Fig. 3E) for large foliose Flacap and Punrud contrasted with much lower and less variable values for Evemes, Parsul, and Phyaip; Pb and Sr shared that pattern. Original Evemes (24 samples) values for most pollution elements were below their peaks higher than Flacap values at the same Index score, while Parsul (23), Phyaip (47), and Punrud (25) values for those elements had varied patterns with Flacap (77). Variability increasing with Index score as for S, Cr, and Cu was common to most pollution elements. Monitor sites (covering ~75% of Index range in S, Cr, and Cu graphs) were for Fig. 3 assigned the average Pollution Index score of the temporary lichen sites near them.

3.3. Calibration of lichen data to monitor station data

Correlations between monitor site PM and N variables for measured pollution and lichen elements Al, Co, Cr, Cu, Fe, N, and S (Table 4) or lichen Pollution Indices were reliably strong. Correlations of these lichen variables with monitor site variables for most other individual elements were less reliable and often weaker. In contrast, lichen Mn had negative correlations with monitor site variables including Mn. Most non-significant results in Table 4 and few positive correlations of a lichen element with its monitor site variable reflect few available monitor sites (Supplementary Document 2). Quantitative calibration of lichen elemental concentrations to measured levels of elements was not attempted because of small sample sizes.

3.4. Patterns of elemental association and relationship to environment

The strongest lichen elemental association pattern was the group of local pollution elements noted in Section 3.3 and Table 4. Axes 1 and 2 from PCA of sites by elements (Fig. 4) displayed 51% of variance; each was significant at $p=0.001$ (Supplementary Document 8). Similar patterns for Al, Co, Cr, Cu, Fe, N, and S whose pollution origins were supported from correlations with monitor site data (Table 4) were confirmed with PCA and direct simple pairwise correlations (Supplementary Document 7). All had strong negative correlation with PCA Axis 1 (vectors point left); all had strong pairwise correlations from both Flacap samples and site elemental averages. N and S were somewhat distinct from the five metals in all analyses (pattern stronger from correlations). Nutrient elements K and P were tightly linked both in the PCA (negatively correlated with Axis 2) and from correlations, with Mg loosely linked to them. Each had moderate to weak correlations with the seven local pollution elements. Most other elements were positively correlated with PCA Axis 2, inverse to C; they had weak to no correlations with the local pollution elements, nor with each other. (Supplementary Document 7). Positively correlated Sr and Ca had negative correlations with most other elements, a pattern not obvious in PCA, but associated with soil dust contamination (Supplementary Document 4).

Relationships of lichen elements with environmental variables in PCA (Fig. 4) were consistent with direct simple and partial correlations between elements and variables (Supplementary Document 1). Pollution Indices accurately reflected pollution element patterns on the PCA, though interpretation of Axis 1 as a pollution gradient was supported primarily by correlations of elements with monitor site variables (Table 4). Modeled N, representing in Fig. 4 all modeled pollution variables, was more strongly linked with geography, climate, and land cover than with lichen pollution elements both in the PCA (correlated more with Axis 2) and from direct pairwise correlations (Supplementary Document 1). Environmental variables were often correlated with each other, but only latitude and tem-

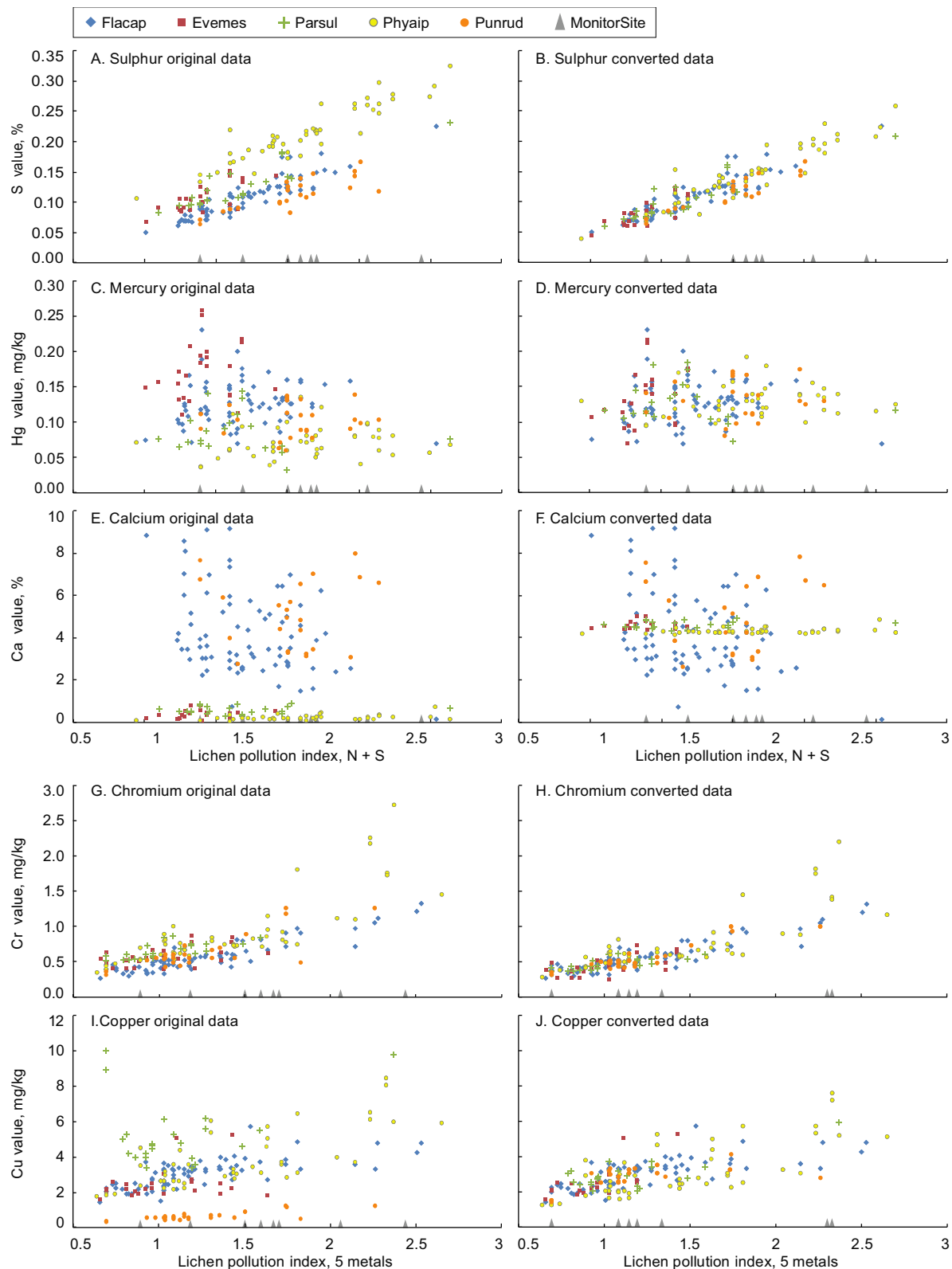


Fig. 3. Scatterplots for example elements S, Hg, Ca, Cr, and Cu of original sample data for all lichen species (left column) compared with Flacap data and converted sample data for other species (right column). Samples excluded for conversion were also excluded from graphs in the right column.

perature were so strongly correlated (negative: $r^2/\rho^2 = 0.86$ to 0.89) that they were not entered into the same partial correlations. Temperature and Forest Cover varied along a NE-SW gradient across ecoregions in the study area (Fig. 1), represented mostly on PCA axis 2. Moderate to weak correlations of lichen elements with

environmental variables were usually stronger than with modeled pollution variables. Land cover variables had few strong links with elemental values or lichen species, with two exceptions. Pb had positive simple and partial correlations at $r^2 > 0.75$ with Developed Land Cover. Flacap was absent from several sites (including

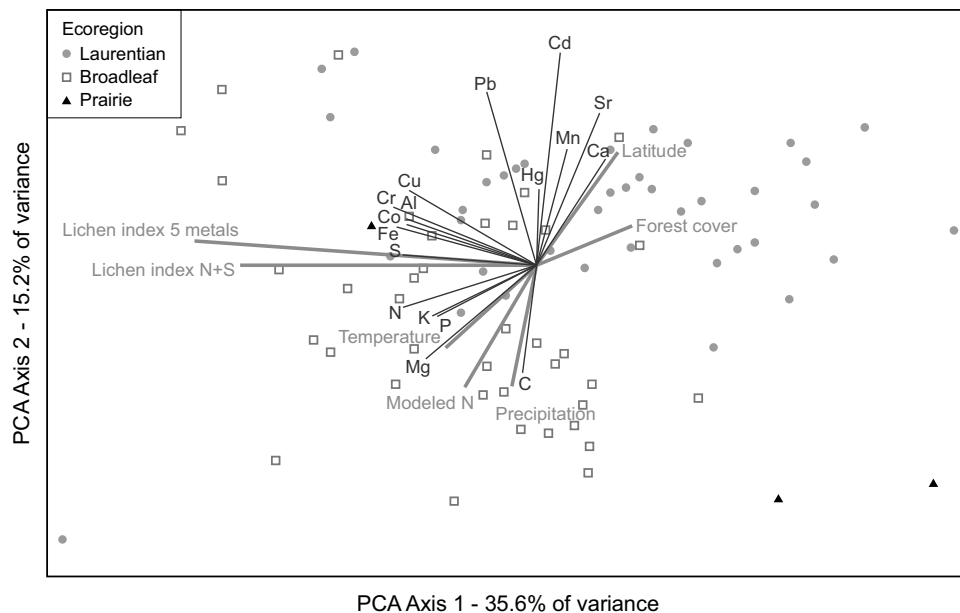


Fig. 4. Principal components analysis (PCA) of sites by elements. Correlations with Axis 1 of Lichen Index 5 metals ($r^2 = 0.77$) and Cr ($r^2 = 0.765$) are similarly strong. Vector scales differ for single elements (thin, blue lines) vs. secondary variables (thick, red lines). Full names of ecoregions in Fig. 1 legend.

Prairie Parkland sites) having low forest cover and moderate Pollution Index scores, as well as from several high Index score urban sites with low forest cover, a pattern not reflected in elemental correlations.

Maps of selected variables and lichen elements (Fig. 5) indicate greater air pollution in the more southern and more populated Eastern Broadleaf Forest ecoregion. Modeled total N deposition (Fig. 5A; modeled Hg and S similar) depicts a smoother pattern of local air pollution than do lichen Pollution Indices for N + S or pollution metals (Fig. 5B, D–F). Maps confirm conclusions from Fig. 3 that the N + S (Fig. 5B) and pollution metals Indices (Fig. 5D–F) differ in geographic pattern. Spatial patterns of lichen Al (Fig. 5D) and Cu (Fig. 5F) showed the widest variation among those in the Pollution Index, 5 metals (Fig. 5E). Lichen Hg pattern (Fig. 5C) was notably different from other lichen pollution elements or modeled N.

4. Discussion

4.1. Bioindicator development as a model for large monitoring programs

Samples collected across the project area by mostly non-specialist field staff following rigorous protocols supported evaluation of all five lichen species as indicators for local pollution, achieving our first objective. Will-Wolf et al. (2017a) found that relaxed cleanliness protocols and collecting from too few substrates unacceptably reduced data quality. Sample collection time (~3/4 h/site), preparation time (1/2–1 h/sample), and elemental measurement cost (~2013 USA \$50/sample) were acceptable for large-scale application (Will-Wolf et al., 2017b). Validation of measured elements relied on replicates of reference samples as well as standard screening of single measurements; both protocol elements should be routinely included. Field site replicates and laboratory splits (not useful) contributed little to data validation. Data screening for general patterns as well as individual values identified entire samples for exclusion to support quality data analysis; this is recommended for routine analysis. For instance, evaluation of a Ca/Sr pattern suggested an unusual soil dust signal and a possible cause for data issues with some excluded samples (Supplementary

Document 4, section S4.1). Examination of scatterplots helped visualize conversion needs, samples with odd data to be considered for exclusion, and elements such as Ca whose accumulation patterns might be linked to lichen morphology.

Data conversion for equivalency among the five lichen species was also successful, achieving this objective for all species. Reliable models were supported by field site replicates of multiple target species at many sites across the pollution gradient. Exclusion of eligible sites was species-specific; no site was excluded for more than one species pair. Punrud data needed conversion for the fewest elements (13) to equivalence with morphologically similar Flacap. Original Ca, Pb, and Sr data overlapped between Punrud and Flacap but differed from the other morphologically distinct species. Elemental accumulation patterns linked with lichen morphology (also Bennett, 2008; Bennett and Wetmore, 1997) are consistent with accumulation through passive particle trapping (Bargagli and Mikhailova, 2002; Conti and Cecchetti, 2001; Sloof and Wolterbeek, 1993). Comparisons with two other studies using similar protocols (Will-Wolf et al., 2014, 2015b) suggest that species data conversion may be needed for more elements in large areas with multiple pollution sources (Supplementary Document 9). Specific conversion models were unique to each study, reinforcing their spatial scale- and context-dependence.

Flacap and Phyaip were the two most important target species, between them covering the project region. Easy species recognition by non-specialists helped; their identification success was ~95% for Evemes, Flacap, or Phyaip and ~70% for Parsul or Punrud (Will-Wolf et al., 2017a). Evemes data showed saturation of many elements at moderate air pollution loads as estimated from Indexes, and absence at higher loads; similar patterns with other species have been reported by Hauck et al. (2002), Sloof and Wolterbeek (1993), and Yemets et al., 2014. Evemes pollution sensitivity limits its usefulness for bioindication in the region, despite easy recognition. Parsul and Punrud were also less useful, linked to field staff identification problems; they had few samples, few unique sites, and many sample and data quality issues (Will-Wolf et al., 2017a). Punrud success from expert collectors in two similar studies (Supplementary Document 9) suggests collector expertise explains the contrast; the species is common across our project region (Jovan

Table 4
Correlations between monitor site variables and lichen elemental data, Indices, or modeled air pollution. Values of the latter were averages for nearby lichen sites.

	Monitor site measured variables (maximum number of sites in parentheses)									
Lichen element	PM(8)	N(6)	NOx(6)	Fe(5)	Mn(6)	Pb(6)	Ni(6)	Cr(6)	S(7)	
Co	0.976***	0.886*	1.0***	1.0***	1.0***	0.957*	0.906*	0.900*	0.900*	
Cu	0.952***	0.886*	1.0***	1.0***	1.0***	0.943**	0.970**	0.917*	0.900*	
S	0.952***	0.943**	1.0***	0.900*	0.943**	0.886*	ns	ns	ns	
Cr	0.952***	0.943**	1.0***	0.900*	0.943**	0.886*	ns	ns	ns	
N	0.929***	1.0***	0.960**	ns	ns	0.943**	ns	ns	0.900*	
Fe	0.919**	0.884*	0.934**	0.903*	ns	0.849*	0.952*	0.900*	ns	
Al	0.913**	ns	0.841*	ns	ns	ns	ns	ns	ns	
Na	0.905**	0.886*	0.943**	0.900*	0.900*	ns	0.838*	ns	ns	
Pb	0.833*	0.886*	0.943**	0.900*	0.900*	ns	ns	ns	ns	
Ni	0.794*	0.943**	ns	ns	ns	0.929**	ns	ns	0.929**	
K	ns	ns	0.913**	ns	ns	0.900*	ns	ns	0.941*	
Mn	−0.833*	ns	−1.0***	−1.0***	−1.0***	−0.900*	−0.928*	−0.943**	−0.900*	
Index, N + S	0.952***	0.971**	1.0***	0.900*	0.943**	0.888*	ns	ns	ns	
Index, 5 metals	0.945***	ns	0.951**	0.900*	0.943**	0.884*	1.0***	1.0***	ns	
Modeled air pollution										
S totdep	0.786*	ns		ns	ns	0.936**	ns	ns	1.0***	

Notes: Spearman rank correlations in standard type; Pearson correlations in italics; numbers in bold indicate a significant correlation between monitor site data and lichen data for the same element.

* = $p < 0.04$; ** = $p < 0.01$; *** = $p < 0.001$; ns = not significant.

See Table 1 for modeled air pollution variables Table 2 for lichen elements, and Supplementary Document 2 for monitor site variables and explanations.

Lichen elements and monitor site variables absent from this table had no or weak correlations with each other. S totdep had the strongest correlations of modeled pollutant deposition variables.

et al., 2017; Brodo et al., 2001). These results supported recommendations for best target species, and improvements to field staff training (Will-Wolf et al., 2017b).

Complementary distribution of Flacap and Phyaip sites had several possible causes. Flacap was indeed collected at more low pollution sites and Phyaip at more high pollution sites, consistent with reported sensitivities (e.g. McCune 1988; Showman 1997; Will-Wolf et al., 2015a), but other evidence suggested additional causes. Flacap data did not, like Evemes, exhibit saturation at higher pollution. Our Flacap maxima were lower for most pollution elements than in two similar studies (Supplementary Document 9). Flacap was also not collected, and Phyaip collected, at several study sites in low-forest landscapes with intermediate air quality. Flacap and Phyaip sample distributions in our study are more consistent with the classic “urban drought hypothesis” (Coppins 1973) and recent studies in Europe (e.g. Tretiach et al., 2012) invoking response of Flacap and other species to lower urban humidity (McCarthy et al., 2010). Flacap population declines in part of the study region (Will-Wolf et al., 2011) were more strongly linked to low-forest landscapes than to high air pollution. Links of lower Flacap abundance with forest fragmentation (perhaps related to dispersal limits as well as humidity) need broader evaluation.

We were less successful at achieving our objective to calibrate lichen elemental data to monitor site data. Lichen data for elements linked with air pollution were indeed positively correlated with ranks of PM2.5 and N variables from eight air monitor sites. This demonstrated, as others have found (e.g. McMurray et al., 2013; also see section 1), that lichen elements were good proxies to indicate relative local air quality. However, we lacked support to calibrate lichen elemental data quantitatively with instrument-measured data. Only PM2.5 was measured at all monitor sites, while correlations between lichen data and instrument-measured data for that element were seldom significant. Sparse availability of costly monitoring sites reinforces the value of lichen bioindicators to represent local air pollution.

Given average within-site variability <25% and within-site value ranges <10% of full range for converted data from most elements and all species (Supplementary Document 6), variation along a pollution gradient or change over time of 30–35% of the full gradient should be detectable in our project area by monitoring patterns and trends for the local pollution elements Co, Cr, Cu, Fe, N, and S, a level of precision useful for large-scale programs. Trends in Al, with higher within-site variability, would need to be stronger for detection. Somewhat weaker patterns and trends should be detectable with the combined Pollution Indices.

4.2. Indication of air quality in the study region

We successfully achieved our objective to evaluate lichen-indicated air quality across the study region. Lichen data from both individual elements and combined Pollution Indices clearly indicated a wide range of air quality conditions across the study region. Inclusion of all five lichen species in analyses contributed additional samples to average across within-site variability and better represent local air quality, even though data from only Flacap and Phyaip would have achieved the objective. Weak correlations of modeled air pollution variables with lichen elemental data, plus stronger correlations of the latter with monitor site data, suggest that lichen elemental data represented local air quality notably better than did regionally-modeled pollution variables, a conclusion consistent with other USA studies (Bari et al., 2001; Boquete et al., 2009; Geiser and Neitlich, 2007; Root et al., 2015; Will-Wolf et al., 2014). This is at least partly because the instrument monitoring stations that support air deposition models are widely scattered but over-represent areas of high pollution concern. Different patterns of local distribution for lichen N and S vs. pollutant metals Al, Co, Cr, Cu, and Fe

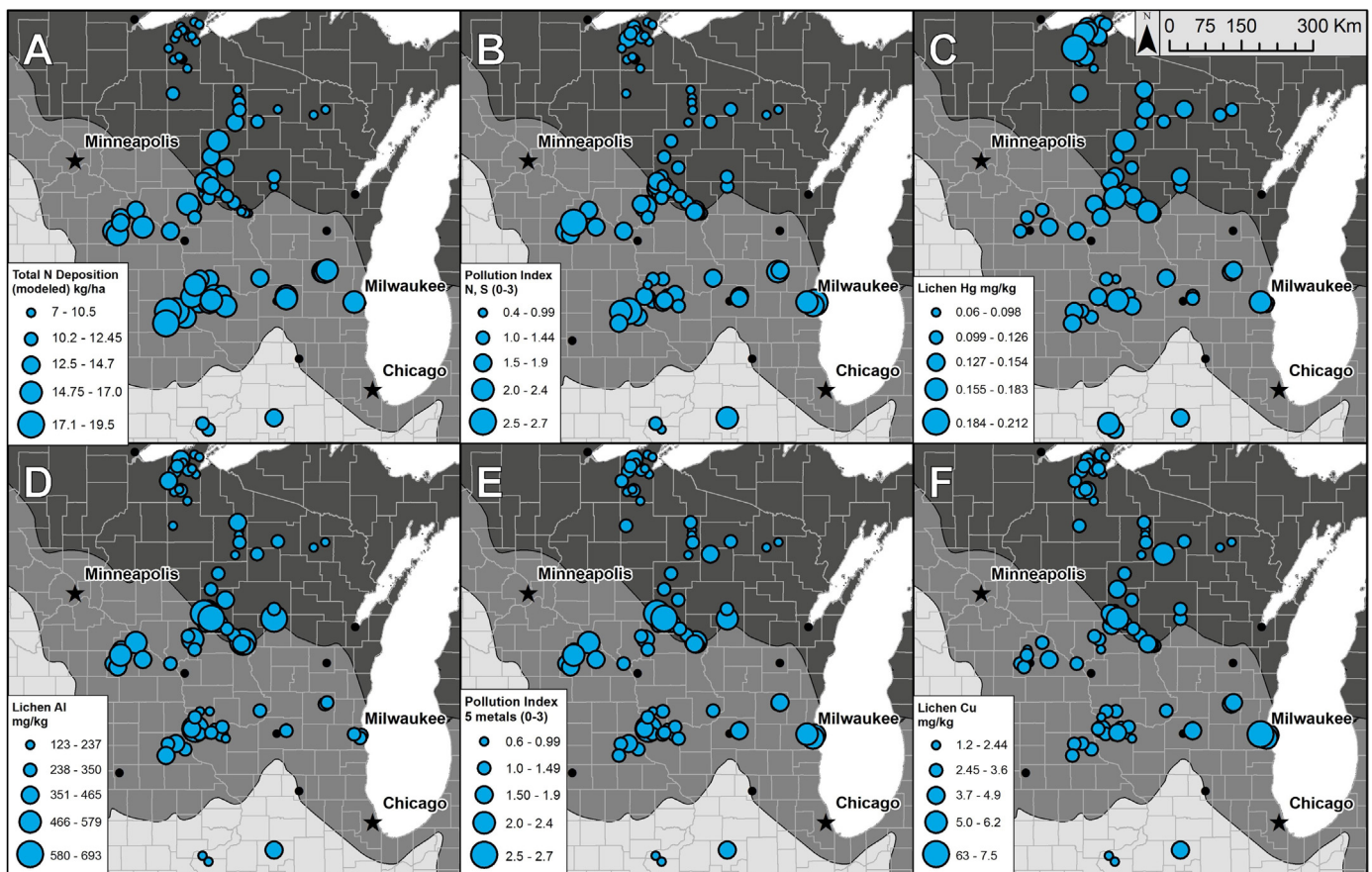


Fig. 5. Mapped modeled N deposition, lichen Pollution Indices, and selected lichen elements. A. Modeled total N deposition 2013; B. Lichen Pollution Index, N + S; C. Lichen Hg; D. Lichen Al; E. lichen Pollution Index, 5 metals; F. Lichen Cu. Scale bar inset (C) is for all maps. Solid dots are cities; ecoregion shading as in Fig. 1.

probably reflected different dispersal modes (N and S in aerosols vs pollution metals in particulates; e. g. [Bargagli and Mikhailova, 2002](#)); possibly also some undocumented differences in sources. Composite Pollution Indices captured these differences well and also averaged over data gaps, an advantage for biomonitoring.

Additional patterns for individual elements are summarized here and reviewed in detail in Supplementary Document 4. Lichen Hg was not well correlated with any other element or environmental variable including modeled Hg deposition, probably linked to known long residence of Hg in atmosphere as it disperses from its documented pollution sources ([Bargagli, 2016](#)). Thus this important pollution element that varies at a more regional scale had no other valid proxy in our study. Lichen Mn data were quite variable, not correlated with other lichen elements reflecting local pollution, and negatively correlated with monitor site data; we recommend, as did [Boquete et al. \(2011\)](#) for mosses, that Mn be excluded from lichen elemental biomonitoring. Lichen Pb appeared to specifically represent urban impact: it had strong correlations with cover of developed land, the only element strongly linked with land cover. Intensive mapping in urban areas of relative Pb load from lichen or moss elemental bioindicators could highlight specific subsections with elevated potential risk for more costly instrument monitoring, similar to the [Donovan et al. \(2016\)](#) study.

Our specific research objectives were achieved with classic, relatively simple, widely available, and equally broadly applied data analysis techniques. Their long history facilitates appropriate application and interpretation by applied researchers as well as academic scientists.

4.3. Applications of general ecological theory to biomonitoring practice

The ecological context- and scale-dependency (e.g. [Levin, 1992](#)) of lichen elemental bioindicators is generally accepted in most modern biomonitoring practice: specific applications and species conversion models are at least by implication recommended for the area and spatial scale used in the study. Performance of bioindicators in the ecological context of the sites studied, and the context of program limitations such as sample collection by non-specialists, are less often addressed in published literature. Including critical details that support interpretation of results in our extensive Supplementary Documents is an acknowledgement of context-dependency useful to include in literature, to support comparisons and improve applications.

A contrast in interpretation of the bioindicator applications of Al and Fe led us to more careful consideration of implications from ecological theory (details in Supplementary Document 4, section S4.2). Both Al and Fe are strongly supported from our study as local pollution indicators along with Co, Cr, Cu, N, and S: from correlations with monitor site data ([Table 4](#)) and between elements (Supplementary Document 7), and from PCA patterns ([Fig. 4](#)). In contrast, in European literature (e.g. [Bargagli and Mikhailova, 2002](#)) Al and Fe are often interpreted as bioindicators for soil dust contamination. Differences in soil chemistry or air pollutant types do not seem satisfactory explanations, since Europe and E USA have similar temperate forest environments and industrial history. Comparisons with two other E USA studies supported two alternate hypotheses. Invoking simple spatial scale-dependency, Al and Fe

might have longer residency in the atmosphere than do Co, Cr, Cu, N, and S, and so vary less across small areas (Will-Wolf et al., 2014, 2015b), consistent with the well-documented regional pattern of Hg distribution (Bargagli 2016). Invoking both multiple causation (e.g. for lichens Ellis and Coppins, 2010; Will-Wolf et al., 2006) and more subtle scale-dependency, Al and Fe might have both soil dust and pollution sources: the dominant signal seen might for instance depend on size of the area studied relative to breadth of the pollution and soil chemistry gradients.

Evaluation in Section 4.1 of multiple factors linked with strong spatial complementarity of Flacap and Phyaip in our study pointed to nearby forest cover as possibly a stronger driver of the pattern than pollution load. Modern European “urban drought” studies suggested a non-pollution mechanism for absence of Flacap from cities with low forest cover (e.g. Tretiach et al., 2012). This might also apply somewhat to small and isolated rural forests, and dispersal limitation, interacting with response to pollution and humidity, is another possible mechanism linking lichens with landscape pattern (e.g. Ellis and Coppins, 2010) that could be further investigated for bioindicator species. Species in the genus *Physcia* are well documented as thriving in high nitrogen habitats and microhabitats (e.g. Conti and Cecchetti, 2001; Fenn et al., 2003; Jovan et al., 2012). Our study also suggested *Physcia* thrives in low-forest landscapes, another pattern deserving more investigation. In another example, proportion of developed land cover was identified in our study as the strongest environmental correlate of Pb pattern, pointing to urban areas as the most important targets for bioindication with Pb (Supplementary Document 4). The importance in our study of evaluating links with environmental drivers other than pollution, including some like local land cover whose relevance is not obvious, likely parallels circumstances for any large monitoring program.

5. Conclusions

Bioindication with elemental data from *in situ* naturally-occurring lichens allows both broader coverage and more intense sampling per total program cost than other options, while it represents local air quality better than regionally modeled variables. Biomonitoring with lichen transplants (Bargagli and Mikhailova, 2002; Garty, 2002), not practically feasible for large-scale monitoring because it requires costly multiple visits per site and single-source transplants not suitable for long climate gradients, has contributed much research about the variation of within-site responses to be averaged across (considered noise in this context) for generating reliable site-wide signals of pollution load. Reliable site-wide signals support monitoring the health and environmental response of biotic communities across a large region. For instance, estimation of N critical loads by calibration of site lichen community response to atmospheric N has been more successful in W USA supported by widespread site-level *in situ* lichen elemental bioindication (Fenn et al., 2003; Geiser et al., 2010; Jovan et al., 2012) than in E USA (Cleavitt et al., 2015; Pardo et al., 2011, 2015) without such wide support.

Considering multiple environmental factors and appropriate ecological theory such as context- and scale-dependence of patterns at all levels of bioindicator design and interpretation improves application of results. Considering cost-effectiveness is more important for large-scale scientific studies and applications to monitoring, than for small-scale studies. These considerations are important to both support quality science and facilitate continued use of lichen elemental bioindication in large monitoring programs (Ferretti and Erhardt, 2002).

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Appendix A. Supplementary information

Supplementary information associated with this article can be found, in the online version, at <http://dx.doi.org/10.1016/j.ecolind.2017.03.017>.

References

- Bailey, R.G., Avers, P.E., King, T., McNab, W.H., 1994. Ecoregions of the United States. In: Map. Scale 1:7,500,000. (+ titles of Subregions). Prepared for the USDA Forest Service, <http://www.fs.fed.us/land/pubs/ecoregions/ecoregions.html>. Accessed December 2014.
- Bargagli, R., Mikhailova, I., 2002. Accumulation of inorganic contaminants. In: Nimis, P.L., Scheidegger, C., Wolseley, P. (Eds.), *Monitoring with Lichens – Monitoring Lichens*. NATO Science Series. Kluwer Academic Publishers, The Hague, NL, pp. 65–84.
- Bargagli, R., 2016. Moss and lichen biomonitoring of atmospheric mercury: a review. *Sci. Total Environ.* 572, 216–231.
- Bari, A., Rosso, A., Minciardi, M.R., Troiani, F., Piervittori, R., 2001. Analysis of heavy metals in atmospheric particulates in relation to their bioaccumulation in explanted *Pseudevernia furfuracea* thalli. *Environ. Monit. Assess.* 69, 205–220.
- Bennett, J.P., Wetmore, C.M., 1997. Chemical element concentrations in four lichens on a transect entering Voyageurs National Park. *Environ. Exp. Bot.* 37, 173–185.
- Bennett, J.P., 2008. Discrimination of lichen genera and species using element concentrations. *Lichenologist* 40, 135–151.
- Boquete, M.T., Fernández, J.A., Aboal, J.R., Real, C., Carballeira, A., 2009. Spatial structure of trace elements in extensive biomonitoring surveys with terrestrial mosses. *Sci. Total Environ.* 408, 153–162.
- Boquete, M.T., Fernández, J.A., Aboal, J.R., Carballeira, A., 2011. Are terrestrial mosses good biomonitors of atmospheric deposition of Mn? *Atmos. Environ.* 45, 2704–2710.
- Boquete, M.T., Fernández, J.A., Aboal, J.R., Carballeira, A., 2015. Relationship between trace metal concentrations in the terrestrial moss *Pseudoscleropodium purum* and in bulk deposition. *Environ. Pollut.* 201, 1–9.
- Brodo, I.M., Sharnoff, S.D., Sharnoff, S., 2001. *Lichens of North America*. Yale University Press, New Haven, Connecticut.
- Cercasov, V., Pantelici, A., Sălăgean, M., Caniglia, G., Scarlat, A., 2002. Comparative study of the suitability of three lichen species to trace-element air monitoring. *Environ. Pollut.* 119, 129–139.
- Cleavitt, N.L., Hinds, J.W., Poirot, R.L., Geiser, L.H., Dibble, A.C., Leon, B., Perron, R., Pardo, L.H., 2015. Epiphytic macrolichen communities correspond to patterns of sulfur and nitrogen deposition in the northeastern United States. *Bryologist* 118, 304–324.
- Cleland, D.T., Freeouf, J.A., Keys Jr., J.E., Nowacki, G.J., Carpenter, C., McNab, W.H., 2007. Ecological subregions: sections and subsections of the conterminous United States [1:3,500,000] [CD-ROM]. In: Sloan, A.M. (Ed.), *Cartog. General Technical Report WO-76*. U.S. Department of Agriculture, Forest Service, Washington, DC.
- Conti, M.E., Cecchetti, G., 2001. Biological monitoring: lichens as bioindicators of air pollution assessment—a review. *Environ. Pollut.* 114, 471–492.
- Coppins, B.J., 1973. The ‘drought hypothesis’. In: Ferry, B.W., Baddeley, M.S., Hawksworth, D.L. (Eds.), *Air Pollution and Lichens*. University of Toronto Press, Toronto, Ontario, CA, pp. 124–142.
- Donovan, G.H., Jovan, S.E., Gatzolis, D., Burstyn, I., Michael, Y.L., Monleon, V.J., Amacher, M.C., 2016. Using an epiphytic moss to identify previously unknown sources of atmospheric cadmium pollution. *Sci. Total Environ.* 569, 84–93.
- Ellis, C.J., Coppins, B.J., 2010. Partitioning the role of climate, pollution and old-growth woodland in the composition and richness of lichens epiphytes in Scotland. *Lichenologist* 42, 601–614.
- Fenn, M.E., Baron, J.S., Allen, E.B., Rueth, H.M., Nydick, K.R., Geiser, L., Bowman, W.D., Sickman, J.O., Meixner, T., Johnson, D.W., Neitlich, P., 2003. Ecological effects of nitrogen deposition in the western United States. *Bioscience* 53, 404–420.

- Ferretti, M., Erhardt, W., 2002. Key issues in designing biomonitoring programmes. In: Nimis, P.L., Scheidegger, C., Wolseley, P. (Eds.), *Monitoring with Lichens – Monitoring Lichens*, NATO Science Series. Kluwer Academic Publishers, The Hague, NL, pp. 111–142.
- Ferry, B.W., Baddeley, M.S., Hawksworth, D.L., 1973. *Air Pollution and Lichens*. University of Toronto Press, Toronto, Ontario, CA.
- Frati, L., Brunialti, G., Loppi, S., 2005. Problems related to lichen transplants to monitor trace element deposition in repeated surveys: a case study from central Italy. *J. Atmos. Chem.* 52, 221–230.
- Gailey, F.A.Y., Lloyd, O.L., 1986. Methodological investigations into low technology monitoring of atmospheric metal pollution: part 3—the degree of replicability of the metal concentrations. *Environ. Pollut.* 12, 85–109 (Series B).
- Gandois, L., Agnan, Y., Leblond, S., Séjalon-Delmas, N., Le Roux, G., Probst, A., 2014. Use of geochemical signatures including rare earth elements, in mosses and lichens to assess spatial integration and the influence of forest environment. *Atmos. Environ.* 95, 96–104.
- Garty, J., 2002. Biomonitoring heavy metal pollution with lichens. In: Kranner, I., Beckett, R.P., Varma, A.K. (Eds.), *Protocols in Lichenology. Culturing, Biochemistry, Ecophysiology and Use in Biomonitoring*. Springer-Verlag, Berlin, Germany, pp. 458–482.
- Geiser, L.H., Neitlich, P., 2007. Air pollution and climate gradients in western Oregon and Washington indicated by epiphytic macrolichens. *Environ. Pollut.* 145, 203–218.
- Geiser, L.H., Jovan, S.E., Glavich, D.A., Porter, M.K., 2010. Lichen-based critical loads for atmospheric nitrogen deposition in Western Oregon and Washington Forests, USA. *Environ. Pollut.* 158, 2412–2421.
- Halleraker, J.H., Reimann, C., de Caritat, P., Finne, T.E., Kashulina, G., Niskaavaara, H., Bogatyrev, I., 1998. Reliability of moss (*Hylocomium splendens* and *Pleurozium schreberi*) as a bioindicator of atmospheric chemistry in the Barents region: interspecies and field duplicate variability. *Sci. Total Environ.* 218, 123–139.
- Hauck, M., Mulack, C., Paul, A., 2002. Manganese uptake in the epiphytic lichens *Hypogymnia physodes* and *Lecanora conizaeoides*. *Environ. Exp. Bot.* 48, 107–117.
- International Atomic Energy Association (IAEA), 2014. Reference products for environment and trade. In: Reference Materials. Trace Elements & Methyl Mercury, Website <<http://nucleus.iaea.org/rpst/ReferenceProducts/ReferenceMaterials/TraceElementsMethylmercury/index.htm>> Accessed October 2014.
- IUPAC, 2014. Periodic Table of the Elements, <<http://www.iupac.org/>> Accessed October 2014.
- Jovan, S., Riddell, J., Padgett, P.E., Nash III, T.H., 2012. Eutrophic lichens respond to multiple forms of N: implications for critical levels and critical loads research. *Ecol. Appl.* 22, 1910–1922.
- Jovan, S., Will-Wolf, S., Geiser, L., Dillman, K., 2017. User Guide for the National FIA Lichen Database (beta). Draft Gen. Tech. Rep. PNW-GTR-XXX. Portland, OR: U.S. Department of Agriculture, Forest Service, Pacific Northwest Research Station, XXp.
- Karakas, S.Y., Tuncel, S.G., 2004. Comparison of accumulation capacities of two lichen species analyzed by instrumental neutron activation analysis. *J. Radioanal. Nucl. Chem.* 259, 113–118.
- Levin, S.A., 1992. The problem of pattern and scale in ecology: the Robert H. MacArthur award lecture. *Ecology* 73, 1943–1967.
- Loppi, S., Giordani, P., Brunialti, G., Isocrono, D., Piervittori, R., 2002. Identifying deviations from natural diversity for bioindication purposes. In: Nimis, P.L., Scheidegger, C., Wolseley, P. (Eds.), *Monitoring with Lichens – Monitoring Lichens*, NATO Science Series. Kluwer Academic Publishers, The Hague, NL, pp. 281–284.
- Martin, M.H., Coughtrey, P.J., 1982. *Biological Monitoring of Heavy Metal Pollution*. Applied Science Publishers, London, UK.
- McCarthy, M.P., Best, M.J., Betts, R.A., 2010. Climate change in cities due to global warming and urban effects. *Geophys. Res. Lett.* 37, L09705, <http://dx.doi.org/10.1029/2010GL042845>.
- McCune, B., Mefford, M.J., 2016. PC-ORD. Multivariate Analysis of Ecological Data. Version 6.21. MjM Software Design, Gleneden Beach, OR, pp. 6, <<https://www.pcord.com/pcordwin.htm>> Accessed June 201.
- McCune, B., 1988. Lichen communities along O₃ and SO₂ gradients in Indianapolis. *Bryologist* 91, 223–228.
- McMurray, J.A., Roberts, D.W., Fenn, M.E., Geiser, L.H., Jovan, S., 2013. Using epiphytic lichens to monitor nitrogen deposition near natural gas drilling operations in the Wind River range, WY, U. S. A. *Water Air Soil Pollut.* 224, 1487–1501.
- Nimis, P.L., Scheidegger, C., Wolseley, P. (Eds.), 2002. *Kluwer Academic Publishers, The Hague, Netherlands*.
- Paoli, L., Guttová, A., Grassi, A., Lackovicová, A., Senko, D., Loppi, S., 2014. Biological effects of airborne pollutants released during cement production assessed with lichens (SW Slovakia). *Ecol. Indic.* 40, 127–135.
- Pardo, L.H., Fenn, M.E., Goodale, C.L., Geiser, L., Driscoll, C.T., Allen, E.B., Baron, J.S., Bobbink, R., Bowman, W.D., Clark, C.M., Emmett, B., Gilliam, F.S., Greaver, T.L., Hall, S.L., Lilleskov, E.A., Liu, L., Lynch, J.A., Nadelhoffer, K.J., Perakis, S.S., Robin-Abbott, M.J., Stoddard, J.L., Weathers, K.C., Dennis, R.L., 2011. Effects of nitrogen deposition and empirical nitrogen critical loads for ecoregions of the United States. *Ecol. Appl.* 21, 3049–3082.
- Pardo, L.H., Robin-Abbott, M.J., Fenn, M.E., Goodale, C.L., Geiser, L., Driscoll, C.T., Allen, E.B., Baron, J.S., Bobbink, R., Bowman, W.D., Clark, C.M., Emmett, B., Gilliam, F.S., Greaver, T.L., Hall, S.L., Lilleskov, E.A., Liu, L., Lynch, J.A., Nadelhoffer, K.J., Perakis, S.S., Stoddard, J.L., Weathers, K.C., Dennis, R.L., 2015. Effects and empirical critical loads of nitrogen for ecoregions of the United States. ch. 5. In: de Vries, W., Hetteling, J.-P., Posch, M. (Eds.), *Critical Loads and Dynamic Risk Assessments*, Environmental Pollution 25. Springer Science + Business Media, Dordrecht, The Netherlands, pp. 129–169, http://dx.doi.org/10.1007/978-94-017-9508-1_5.
- Puckett, K.J., 1988. Bryophytes and lichens as monitors of metal deposition. In: Nash, T.H., Wirth, V. (Eds.), *Lichens, Bryophytes and Air Quality*, 30. Bibliotheca Lichenologica, pp. 231–267.
- Root, H.T., Geiser, L.H., Fenn, M.E., Jovan, S., Hutten, M.A., Ahuja, S., Dillman, K., Schirokauer, D., Berryman, S., McMurray, J.A., 2013. A simple tool for estimating throughfall nitrogen deposition in forests of western North America using lichens. *For. Ecol. Manage.* 306, 1–8.
- Root, H.T., Geiser, L.H., Jovan, S., Neitlich, P., 2015. Epiphytic macrolichen indication of air quality and climate in interior forested mountains of the Pacific Northwest, USA. *Ecol. Indic.* 53, 95–105.
- SPSS, 2015. IBM SPSS Statistics. Version 23.0.0., pp. 1989–2015.
- Showman, R.E., 1997. Continuing lichen recolonization in the upper Ohio River Valley. *Bryologist* 100, 478–481.
- Sloof, J.E., Wolterbeek, B.T., 1993. Interspecies comparison of lichens as biomonitors of trace-element air pollution. *Environ. Monit. Assess.* 25, 149–157.
- Tretiach, M., Pavanetto, M., Pittao, E., Sanità di Toppi, L., Piccotto, M., 2012. Water availability modifies tolerance to photo-oxidative pollutants in transplants of the lichen *Flavoparmelia caperata*. *Oecologia* 168, 589–599.
- United States Department of Agriculture, Forest Service [USDA FS], 2015. Field Guides for Standard (Phase 2) Measurements. V.7.0. U.S. Department of Agriculture, Forest Service, Forest Inventory and Analysis, FIA, Library, Field Guides, Methods, and Procedures, <http://http://www.fia.fs.fed.us/library/field-guides-methods-proc/index.php> Accessed June 2016.
- Vestergaard, N.K., Stephansen, V., Resmussen, L., Pilegaard, K., 1986. Airborne heavy metal pollution in the environment of a Danish steel plant. *Water Air Soil Pollut.* 27, 263–377.
- Will-Wolf, S., Geiser, L.H., Neitlich, P., Reis, A., 2006. Comparison of lichen community composition with environmental variables at regional and subregional geographic scales. *J. Veg. Sci.* 17, 171–184.
- Will-Wolf, S., Nelsen, M.P., Trest, M.T., 2011. Does morphological response of four common lichen species to pollution, shade, and landscape pattern predict long-term changes in distribution? In: Bates, S., Bungartz, F., Elix, J., Herrera, M., Lücking, R., Zambrano, A. (Eds.), *Biomonitoring, Ecology, and Systematics of Lichens*, 106. Festschrift Thomas H. Nash III. Bibliotheca Lichenologica, 375–102.
- Will-Wolf, S., Jovan, S., Nelsen, M.P., Trest, M.T., Rolih, K., Reis, A., 2014. Lichen community indexes for response to climate and air quality in the Midatlantic states, USA. In: Internal USDA FIA Report, October 2014., pp. 2014. Available from the authors.
- Will-Wolf, S., Jovan, S., Neitlich, P., Peck, J.L., Rosentreter, R.R., 2015a. Lichen-based indexes evaluate responses to climate and air pollution across northeastern U.S.A. *Bryologist* 118, 59–82.
- Will-Wolf, S., Makhholm, M.M., Nelsen, M.P., Trest, M.T., Reis, A., Jovan, S., 2015b. Element analysis of two common macrolichens supports bioindication of air pollution and lichen response in rural midwestern U.S.A. *Bryologist* 118, 371–384.
- Will-Wolf, S., Jovan, S., Amacher, M.C., 2017a. Lichen elements as environmental indicators: evaluation of methods for large monitoring programs. *The Lichenologist* 26 (January), 2017. Accepted for publication.
- Will-Wolf, S., Jovan, S., Amacher, M.C., 2017b. Lichen elemental indicators for air pollution in eastern U.S.A. forests; a pilot study in the upper Midwest. In: Submitted for a USDA Forest Service Pacific Northwest Research Station General Technical Report. PNW-GTR-XXX.
- Wolterbeek, B., 2002. Biomonitoring of trace element air pollution: principles: possibilities and perspectives. *Environ. Pollut.* 120, 11–21.
- Yemets, O.A., Solhaug, K.A., Gauslaa, Y., 2014. Spatial dispersal of airborne pollutants and their effects on growth and viability of lichen transplants along a rural highway in Norway. *Lichenologist* 46, 809–823.