

# Niche construction by two ectomycorrhizal truffle species (*Tuber aestivum* and *T. melanosporum*)

Luis G. García-Montero<sup>a,e,\*</sup>, Vicente J. Monleón<sup>b</sup>, Inmaculada Valverde-Asenjo<sup>c</sup>, Cristina Menta<sup>d</sup>, Thomas W. Kuyper<sup>e</sup>

<sup>a</sup> Centro para la Conservación de la Biodiversidad y el Desarrollo Sostenible (CBDS), E.T.S.I. Montes, Forestal y del Medio Natural, Universidad Politécnica de Madrid, 28040, Madrid, Spain

<sup>b</sup> Pacific Northwest Research Station, USDA Forest Service, 3200 SW Jefferson Way, Corvallis, OR, 97331, USA

<sup>c</sup> Dpt. of Chemistry in Pharmaceutical Sciences, Soil Science Unit, Pharmacy faculty, Universidad Complutense de Madrid, Plaza Ramón y Cajal s/n, 28040, Madrid, Spain

<sup>d</sup> Dpt. of Chemistry, Life Sciences, and Environmental Sustainability, University of Parma, Parco Area delle Scienze 11/A, 43124, Parma, Italy

<sup>e</sup> Soil Biology Group, Wageningen University & Research, P.O. Box 47, 6700, AA, Wageningen, the Netherlands

## ARTICLE INFO

### Keywords:

Ectomycorrhizal mutualistic symbiosis

Niche construction

Niche differentiation

*Tuber aestivum*

*Tuber melanosporum*

## ABSTRACT

Niche construction by biota has been frequently reported for animals and plants, but fungi have received less attention. For mycorrhizal fungi, mutualistic niche construction has been proposed where partners construct niches for each other. However, few eco-evolutionary mechanisms leading to mutualistic symbiotic feedback have been described. Here we report niche construction by two ectomycorrhiza-forming species of truffles, *Tuber aestivum* and *T. melanosporum*. The soils of 263 spots of these *Tuber*, which create bare patches (brûlés), have been monitored (9–10 years) in natural forests to analyze whether they meet the criteria to satisfy the mutualistic niche construction process. Soil habitat modification by these *Tuber* can be seen in their brûlés, where the vegetation of arbuscular mycorrhizal plants and soil organisms are largely suppressed. *Tuber melanosporum* brûlés were smaller more productive, and with lower vegetation cover than *T. aestivum*. Interactions among soil carbonates, pH, and brûlé size are associated with increased carpophore productivity, indicating positive feedback between habitat modification and fungal fitness. Both species modified the habitat differentially. *Tuber aestivum* brûlés had higher pH and lower total carbonate than soils outside. *Tuber melanosporum* brûlés had lower total carbonate and higher exchangeable  $\text{Ca}^{2+}$  and active carbonate than soils outside. In conclusion, brûlés show a mutualistic niche construction process because modifies their soil environment and increases the fitness of both *Tuber* spp. Enhanced carbonate weathering by truffles resulted in lower soil quality with a negative impact on plants, indicating that mutualistic niche construction does not equally benefit both partners.

## 1. Introduction

Ecological theory has traditionally focused on how organisms adapt to their environment. However, environmental modification by organisms and their legacy over time are processes that act as sources of modified selection and modified phenotypes through a process called niche construction (Day et al., 2003; Odling-Smee et al., 2003, 2013; Pausas and Bond, 2022). Matthews et al. (2014) proposed three criteria to identify a niche construction process (Criteria 1 and 2) and to determine when it affects evolution (Criterion 3).

- Criterion 1: An organism must significantly modify environmental conditions.
- Criterion 2: Organism-mediated environmental modifications must influence selection pressures on a recipient organism.
- Criterion 3: There must be an evolutionary response in at least one recipient population caused by environmental modification.

The term ecosystem engineering has frequently been used as an equivalent term, although that term focuses less on the evolutionary implications of these processes (Pausas and Bond, 2022). Niche construction is a broad concept and includes examples from animals (dam

\* Corresponding author. E.T.S.I. Montes, Forestal y del Medio Natural, Universidad Politécnica de Madrid, C/ José Antonio Nováis 10, Madrid, 28040, Spain.

E-mail addresses: [luisgonzaga.garcia@upm.es](mailto:luisgonzaga.garcia@upm.es) (L.G. García-Montero), [vicente.monleon@usda.gov](mailto:vicente.monleon@usda.gov) (V.J. Monleón), [mivalver@ucm.es](mailto:mivalver@ucm.es) (I. Valverde-Asenjo), [cristina.menta@unipr.it](mailto:cristina.menta@unipr.it) (C. Menta), [thom.kuyper@wur.nl](mailto:thom.kuyper@wur.nl) (T.W. Kuyper).

<https://doi.org/10.1016/j.soilbio.2023.109276>

Received 25 August 2023; Received in revised form 12 November 2023; Accepted 4 December 2023

Available online 11 December 2023

0038-0717/© 2023 Published by Elsevier Ltd.

building by beavers; landscape modification by fungus-growing termites), plants (dune-building by marram grass and their associated arbuscular mycorrhizal fungi), and microorganisms (through plant-soil feedbacks). Plants and their associated microbes bring changes in biological, chemical, and physical soil properties, and such environmental modifications, which at first sight might seem trivial, produce crucial selection pressures that increase fitness for some species and decrease fitness for others. Mutualistic symbioses, such as those formed between mycorrhizal fungi and plants, are prime examples of niche construction. However, few mutualistic niches have been described in detail, and many eco-evolutionary mechanisms leading to symbiotic feedback still need to be discovered (Peay, 2016). Habitats, considered suitable for certain plants, are sometimes not colonized without a suitable mycorrhizal fungal partner (Nuñez et al., 2009; Peay 2016). The differential modification of plant-soil feedback by different guilds of mycorrhizal fungi indicates that the mutualistic niche for one species might reduce the fitness of another species (Kadowaki et al., 2018).

A clear example of niche construction by ectomycorrhizal fungi is formed by truffles of the genus *Tuber* Micheli ex Wiggers. *Tuber* spp. are hypogeous ascomycetes growing in a mutualistic symbiosis with the roots of host plants such as *Quercus* L., *Corylus* L., *Populus* L., *Fagus* L., and *Cistus* L. Upon ripening, *Tuber* ascocarps emit distinctive species-specific volatile organic compounds that allow them to be located by mammals such as wild boar or pigs, trained dogs, or insects. After location, truffles are picked up by those animals, and through that mechanism, their spores are dispersed (Ceruti et al., 2003). Plants colonized by several *Tuber* species show an almost bare zone around the trunk of their host plants, called brùlé, where the growth of other non-host plants and soil organisms are inhibited (Fig. 1). There are several hypotheses about how those bare patches, which look like burnt sites, originate. The allelopathy hypothesis relates to the production of phytotoxic substances that affect seed germination, seedling establishment, and/or the growth of adult plants other than the host tree. An alternative parasitism hypothesis noted that these plant species, which are commonly colonized by arbuscular mycorrhizal fungi, are colonized by the truffle mycelium and potentially acquire carbon from these non-hosts without exchanging them for nutrients (Plattner and Hall, 1995; Gryndler et al., 2014; Schneider-Maunoury et al., 2018, 2020). The brùlé structure is mainly associated with three species of *Tuber*: The Mediterranean *T. melanosporum* Vittad. (Black truffle), the cosmopolitan *T. aestivum* Vittad. (Summer truffle), and the Chinese *T. indicum* Cooke and Massee (Streiblová et al., 2012). On the other hand, in natural forests, most studies on *T. aestivum* and *T. melanosporum* do not provide statistical data on brùlé development, carpophore production, and other

quantitative data associated with truffle ecology.

The brùlés created by *T. melanosporum* and *T. aestivum* provide attractive biological models in calcareous soils to analyze edaphic modification associated with these *Tuber* spp. (García-Montero et al., 2006, 2007, 2009a, 2009b, 2012; Menta et al., 2014). These studies demonstrated an impact of brùlé development on soil calcium carbonate and pH balances, and these soil chemical modifications, as a form of niche construction, could have feedback effects on the performance of these *Tuber* species and their host and non-host plants (García-Montero et al., 2009a, 2012). Bragato (2014) proposed that the brùlé could be a case of perturbational niche construction. His study in one brùlé created by an unidentified *Tuber* species demonstrated a significantly lower contribution of large macro-aggregates inside the brùlé compared to the adjacent area that was covered by grasses. Changes in soil aggregation were likely driven by the disappearance of grasses in the brùlé, which modified the impact of freeze-thaw cycles on soil aggregation and soil aeration that could have improved the conditions for sporocarp formation by the truffle due to lower resistance for sporocarp enlargement. Bragato (2014) put these observations explicitly in the niche construction framework of Odling-Smee et al. (2003, 2013).

The possibility that the edaphic chemical changes that occur during the development of brùlés are also an instance of niche construction has, to the best of our knowledge, not been studied before. The general objective of the present study was to evaluate, in the framework of niche construction theory, soil chemical changes caused by two truffle species, *T. melanosporum*, and *T. aestivum*, based on long-term observations of many wild truffle populations in Mediterranean forests located in Central Spain around the Natural Park of “Alto Tajo”. In our study, we focus on criteria 1 and 2 by Matthews et al. (2014) as sufficient to demonstrate niche construction.

## 2. Material and methods

### 2.1. Study area

Brùlés of *T. melanosporum* and *T. aestivum* were monitored in 9 municipalities of central Spain (Sigüenza and Peralejos de las Truchas, province of Guadalajara; Medinaceli, Arcos de Jalón and Chaorna, province of Soria; Masegosa, El Tobar, Cueva del Hierro and Beteta, province of Cuenca). The study area is within a 40 km radius around the town of Huertahernando (province of Guadalajara; geographic coordinates: 40° 49' 25" N, 2° 17' 12" W). The exact geographical coordinates of the *Tuber* brùlés are not given here as they were confidentially provided to the first author by professional truffle hunters who participated in this study and who wanted to preserve the exact locations that they exploited. The study area is a mountainous region with elevations ranging from 1000 to 1450 m.a.s.l. in a supra-Mediterranean bio-climatic belt. Its lithology is predominantly Jurassic and Cretaceous limestone and dolomite. Soils are frequently lithic and rendzic leptosols with considerable surface stoniness. The mean annual temperature is 10.8 °C, and the mean monthly minimum and maximum temperatures are 3.3 °C and 18.5 °C, respectively. The mean annual precipitation is 650 mm.

The vegetation of the study area falls within the Mediterranean Ibero-Levantine Province Castellano Maestrazgo-Manchega Sub-province geobotanical classification (Rivas-Martínez, 1987; Peinado et al., 2017). In the study area, two main and one intermediate forest types constitute the most important habitats where both truffle species abound. Both *Tuber* species form ectomycorrhizal associations with *Quercus faginea* Lam. and *Q. rotundifolia* Lam. (syn. *Q. ilex* L. subsp. *ballota* (Desf.) Samp.). The vegetation types are classified as (forest type 1) *Cephalantho rubrae* – *Quercetum fagineae* and (forest type 2) *Junipero thuriferae* – *Quercetum rotundifoliae*. Both *Quercus* species co-occur in the intermediate forest type (forest type 3). Compared to type 1, intermediate type 3 has a more xeric lithology with rocky soils exposed to the sun compared to type 1. Forest type 2 is the most xeric type.



Fig. 1. Brùlé formed by *T. aestivum* associated with *Q. rotundifolia* in a natural forest (Central Spain). Photo by T.W. Kuyper.

## 2.2. Brûlé characterisation

*Tuber melanosporum* has been harvested in these forests since the end of the 1960s, and *T. aestivum* since the early 2000s. With the help of seven professional truffle hunters who harvested those brûlés regularly, we monitored (i) 119 natural *T. aestivum* brûlés for 9 years (2007–2015) and (ii) 144 natural *T. melanosporum* brûlés for 10 years (1994–2000 and 2013–2015). Some of the brûlé data used in this study have been updated in the field during 2013–2015 from data initially published in García-Montero et al. (2007, 2014). Truffles were collected with the help of trained dogs and identified, according to Rioussset et al. (2001).

In each brûlé, we determined host tree identity, size (m<sup>2</sup>) of the brûlé, the percentage of the brûlé surface occupied by plants other than the host of *Tuber* spp., and sporocarp (truffle) production. The maximum annual production of sporocarps was calculated as the largest quantity of fresh sporocarps picked during a harvest season over a minimum period of 9 years. Maximum annual production or productivity (instead of average yield) was chosen to standardize and compare brûlés in the most favorable year for each site. At the beginning of the monitoring of each brûlé, we measured its longest and shortest axis and estimated brûlé area using an ellipse model. In brûlés with an irregular shape, we used a combination of several small ellipses. In 2015, at the end of the monitoring period, we measured the percentage of the brûlé surface occupied by plants other than the ectomycorrhizal hosts of *Tuber* spp. of the 34 brûlés of *T. melanosporum* in forest type 2, we could not measure the percentage of the brûlé surface occupied by plants in 32 *Tuber* brûlés, because access to this area in Beteta was unexpectedly restricted. For the same reason, we could not collect soil samples in Beteta. Our data set on *T. melanosporum* in forest type 2 is therefore restricted to 2 brûlés in the locality Cueva del Hierro.

## 2.3. Soils

Soil samples were taken in 90 brûlés, 45 of each species. Soil sampling inside brûlés had to be restricted because of the high economic value of these brûlés for truffle hunters and the disturbance caused by soil sampling. In all, 134 soil samples were taken: 90 inside vs 44 outside brûlés. We included the data of pH, total organic carbon (TOC), and total and active carbonate of 40 *T. melanosporum* brûlé soils (20 inside vs 20 outside) from García-Montero et al. (2009b) in the dataset. For economic reasons, not all parameters were analysed in the 134 soil samples. The sample size used in the statistical analysis of each soil parameter is shown in Supplementary Table 1. This set was used for testing significant differences between soils in brûlés of *T. aestivum*, soils in brûlés of *T. melanosporum*, and soils outside brûlés. These data also allowed testing for soil modification by each truffle species. For that, we compared 20 samples of brûlés of *T. aestivum* with 20 samples outside brûlés (at a distance greater than 2 m outside the visible brûlé boundary) and 20 samples of brûlés of *T. melanosporum* with 20 samples outside brûlés. Some comparisons (inside/outside) of *T. melanosporum* brûlés have been previously published in García-Montero et al. (2009b). Here we use an expanded set of brûlé characteristics (surface size, sporocarp productivity, plant cover in the brûlés, and soil characteristics), and extend the statistical analysis.

The uppermost 30 cm of each soil profile was sampled because both *Tuber* species fruit in this range (Verlhac et al., 1990). Soils were sampled using the same spade (0.20 m blade) and a tape measure. The samples included a mixture of 1 kg of various soil horizons. Soil horizons could not be recognized because of the constant mixing of horizons due to the continuous digging by the harvesters, their dogs, and wild animals. Sampling was done according to FAO (1990). The following soil properties were assessed: pH in water, total calcium carbonate (equivalent carbonate), total organic carbon (TOC), and exchangeable cations following the methods of ISRIC (1995). Active carbonate (carbonate extractable with ammonium oxalate) was determined according to AFNOR (1982). Active carbonate is the finely divided fraction of

carbonates measuring <50 µm; it is susceptible to rapid mobilization and is chemically very reactive. The amount and fraction of active carbonate are often used as a proxy for predicting iron chlorosis in the vineyard and orchard agroecosystems (Tagliavini and Rombolà, 2001).

## 2.4. Statistics

We first tested, through a two-way analysis of variance, whether the three forest types differed in size, plant cover, and sporocarp productivity for both truffle species, based on the sample of 263 brûlés. We repeated the analysis for 90 brûlés of which we had soil chemical data, to test for differences between forest types and truffle species in soil properties. Finally, we tested through a paired *t*-test for differences in soil properties inside and outside the brûlés, based on 20 paired samples for each species. In some of the analyses, the following extreme outlier values were discarded after testing (Grubbs test): the value of 150 m<sup>2</sup> of the size of a *T. melanosporum* brûlé; the values of 125.55 cmol<sup>+</sup> kg<sup>-1</sup> of exchangeable Ca<sup>2+</sup>, and 125.6 and

116.9 mg g<sup>-1</sup> of TOC inside the brûlés of *T. melanosporum*; and the value of 80 mg g<sup>-1</sup> of TOC inside the brûlé of *T. aestivum*. Soil data were additionally analysed through Principal Component Analysis (PCA).

Data were checked for normality (Shapiro-Wilk and Kolmogorov-Smirnov tests) and homogeneity of variances (Levene test). Transformations (log or square root) were made in case of departure from ANOVA assumptions, as in the values of brûlé size, soil pH, % total carbonate, % active carbonate, exchangeable Ca<sup>2+</sup>, Mg<sup>2+</sup>, and K<sup>+</sup>. In addition, non-parametric tests (Kruskal-Wallis test; Spearman test) were used when data did not comply with ANOVA assumptions, as in the values of the ratio soil % active/total carbonate and TOC. We also used analysis of covariance to compare *Tuber* productivity among forest types while controlling for brûlé size. Following Ramsey and Schaffer (1997), after log-transformation, we interpreted the results as multiplicative effects on the group median. The family-wise error rate was controlled with the Tukey-Kramer procedure. Most analyses were done in R v. 3.0.2 (R Core Team, 2013), but some complementary analysis was done with Statgraphics Centurion 18v Software (2019(1982–2018)).

## 3. Results

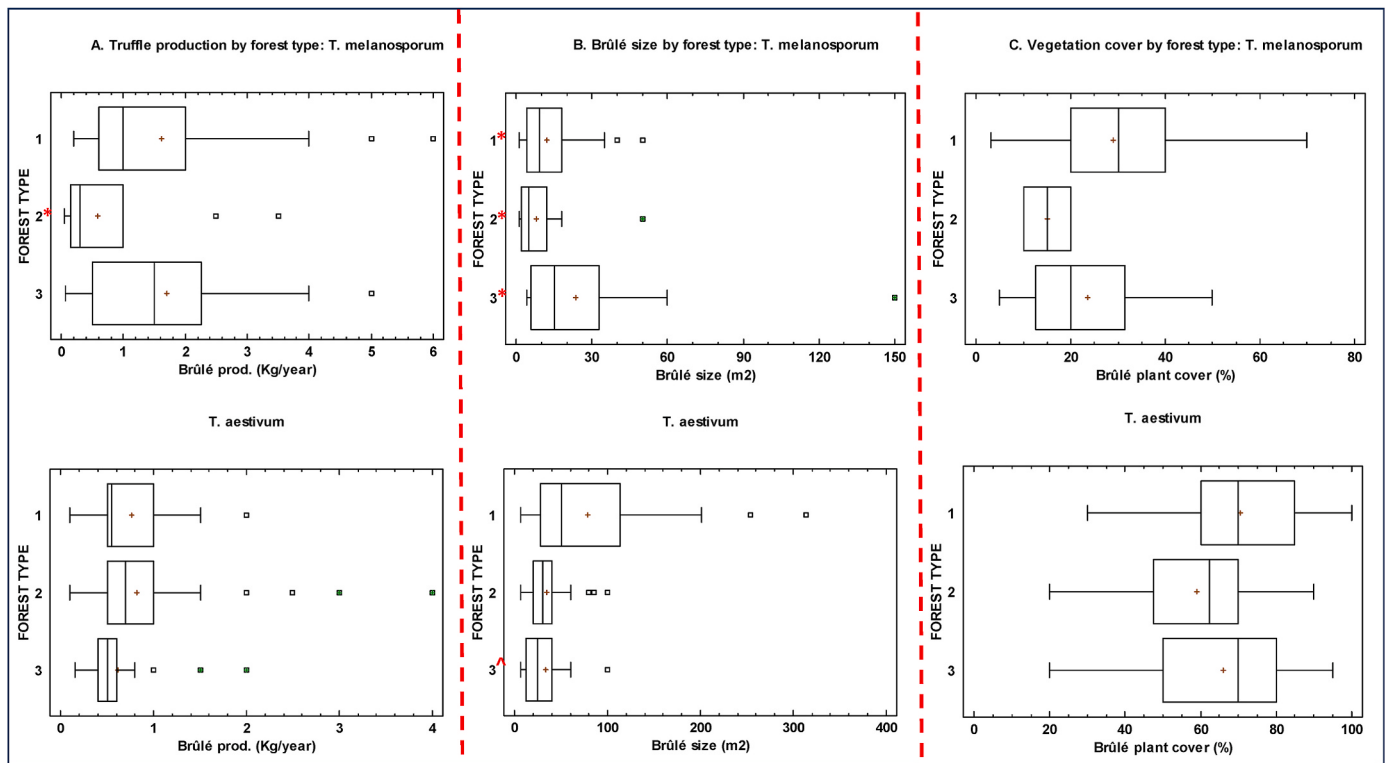
### 3.1. Characteristics of brûlés

Two-way analysis of variance showed that truffle species, forest type, and the interaction truffle species × forest type were significant sources of variation (*Tuber*-spp:  $F_{1,261} = 5.13$ ,  $p = 0.024$ ; forest type:  $F_{2,260} = 13.33$ ,  $p < 0.001$ ; *Tuber*-spp × forest type:  $F_{2,260} = 21.63$ ,  $p < 0.001$ ). Brûlés of *T. aestivum* (119 samples) and *T. melanosporum* (144 samples) showed significantly different characteristics in the three forest types (Fig. 2, Supplementary Table 2). There was a significant difference in *T. melanosporum* sporocarp productivity between forest types, with type 2 (the driest forest) being significantly less productive than the two other forest types.

The productivity of *T. aestivum* did not differ between forest types. *Tuber melanosporum* was much more productive than *T. aestivum* in forest types 1 and 3, but not in type 2 (Fig. 2A).

Two-way ANOVA for brûlé size showed that truffle species, forest type, and the interaction truffle species × forest type were significant sources of variation (*Tuber*-spp:  $F_{1,261} = 113.50$ ,  $p < 0.001$ ; forest type:  $F_{2,260} = 8.59$ ,  $p < 0.001$ ; *Tuber*-spp × forest type:  $F_{2,260} = 10.36$ ,  $p < 0.001$ ). Brûlés of *T. aestivum* were, on average, much larger than those of *T. melanosporum* (Fig. 2B). Brûlé size of *T. melanosporum* was smallest in forest type 2, and this coincided with the lowest productivity. Brûlé size of *T. melanosporum* was largest in the forest type 3, whereas it was smallest for *T. aestivum* in that forest type; therefore, species-specific differences in brûlé size were significant in both the most mesic (type 1) and xeric (type 2) forest type, but not in the intermediate type 3.

Two-way ANOVA showed that truffle species was a significant source



**Fig. 2.** Brûlé characteristics of *T. melanosporum* and *T. aestivum* in the different forest types. Sporocarp productivity (A), brûlé size (B), and vegetation cover (C). Forest type 1: formation of *Quercus faginea*, forest type 3, mixed forest with *Q. faginea* and *Q. ilex*, forest type 2: forest with *Q. ilex* [\* $p < 0.05$ ; + $p < 0.01$ ; \* $p < 0.001$ ].

of variation in brûlé vegetation cover, whereas forest type and the interaction truffle species  $\times$  forest type were not (*Tuber*-spp:  $F_{1,229} = 226.04$ ,  $p < 0.001$ ; forest type:  $F_{1,228} = 3.15$ ,  $p = 0.078$ ; spp  $\times$  forest type:  $F_{1,229} = 0.02$ ,  $p = 0.890$ ). *Tuber melanosporum* reduced vegetation cover in the brûlé much more than *T. aestivum* (Fig. 2C). Outside brûlés, vegetation cover was close to 100%.

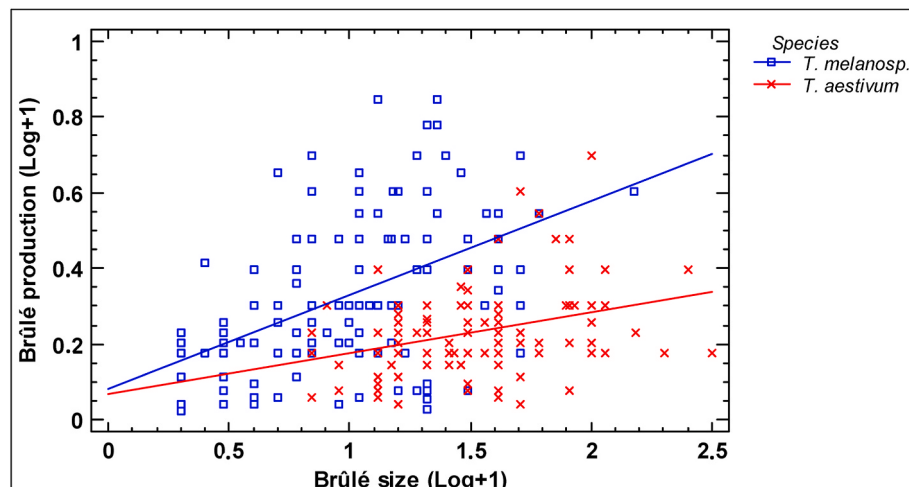
Brûlé size was significantly correlated with sporocarp productivity of both *T. melanosporum* ( $\log [\text{prod}] = -1.1732 + 0.4912 \log [\text{size}]$ ,  $r^2 = 0.24$ ,  $p < 0.001$ ) and *T. aestivum* species ( $\log [\text{prod}] = -1.3937 + 0.2691 \log [\text{size}]$ ,  $r^2 = 0.10$ ,  $p < 0.001$ ). This regression indicates that with a doubling of the brûlé area, the yield increases by 41% in *T. aestivum* ( $2^{0.4912} = 1.41$  times) and by 21% in *T. melanosporum* ( $2^{0.2691} = 1.21$  times). While in the same brûlé area, the productivity of *T. melanosporum* is much higher than that of *T. aestivum* (Fig. 3). Because of the significant

interaction between truffle species  $\times$  forest type for both productivity and

brûlé size, we repeated the analyses for that correlation for forest sites separately. For *T. aestivum*, the relationship between brûlé size and its productivity was significant but did not depend on forest type (forest type:  $F_{2,116} = 1.95$ ,  $p = 0.17$ ;  $\log [\text{size}]$ :  $F_{1,117} = 13.23$ ,  $p < 0.001$ ).

For *T. melanosporum*, the relationship between brûlé size and its productivity was significant and depended on forest type (forest-type:  $F_{2,141} = 14.85$ ,  $p < 0.001$ ;  $\log [\text{size}]$ :  $F_{1,142} = 28.66$ ,  $p < 0.001$ ;  $\log [\text{size}] \times$  forest type:  $F_{2,141} = 3.79$ ,  $p = 0.02$ ).

We analysed the relationship between (non-host) plant cover in brûlés and sporocarp productivity. In *T. aestivum*, the positive relationship between plant cover in brûlés and sporocarp productivity was significant and did not depend on forest type or the interaction between



**Fig. 3.** The relationship between brûlé size and maximum productivity for *T. aestivum* (red) and *T. melanosporum* (blue).

those two variables (forest type:  $F_{2,116} = 1.22$ ,  $p = 0.30$ ; plant cover:  $F_{1,117} = 5.50$ ,  $p = 0.02$ ; plant cover  $\times$  forest type:  $F_{2,116} = 0.49$ ,  $p = 0.61$ ). However, for *T. melanosporum*, the relationship between brûlé plant cover and the productivity was only significant if the effect of the forest type was included (forest type:  $F_{1,142} = 6.19$ ,  $p = 0.014$ ; plant cover:  $F_{1,142} = 0.26$ ,  $p = 0.61$ ; plant cover  $\times$  forest type:  $F_{1,142} = 6.72$ ,  $p = 0.01$ ). The significant interaction between plant cover and forest type was because, in forest type 1, productivity increased when plant cover decreased, whereas, in forest type 3, productivity increased when vegetation covers increased.

### 3.2. Soils inside and outside brûlés

In our analysis of soil modification by both truffle species we compared three categories, viz., brûlés of *T. aestivum*, brûlés of *T. melanosporum*, and soils outside brûlés (Fig. 4; Table 1). There was a significant difference in pH among the three groups (square root

transformed data of pH,  $F_{2,131} = 7.86$ ,  $p < 0.001$ ). There was no effect of forest type ( $p = 0.188$ ). Soils from *T. melanosporum* brûlés were, on average, 0.10 and 0.25 units of pH higher than soils of *T. aestivum* brûlés and soils outside brûlés, respectively (Fig. 4A). Both differences were significant. Soils from *T. aestivum* brûlés showed, on average, 0.14 units of pH higher than soils outside the brûlés, a difference that was also significant. Whereas forest type did not impact soil pH, the pH modification in *T. aestivum* brûlé showed significant interactions with brûlé size, with larger brûlés resulting in a stronger pH increase compared to neighboring soil ( $p = 0.030$ ). The effect of plant cover on brûlé pH was marginally significant ( $p = 0.080$ ). There was no relationship between sporocarp productivity and pH.

Total carbonate content was significantly higher outside brûlés and inside brûlés of *T. melanosporum* than inside brûlés of *T. aestivum*, indicating modification of the carbonate pool by the latter truffle species (Fig. 4B). Two-way ANOVA showed that soil group and forest type were significant sources of variation for total carbonate content, whereas the

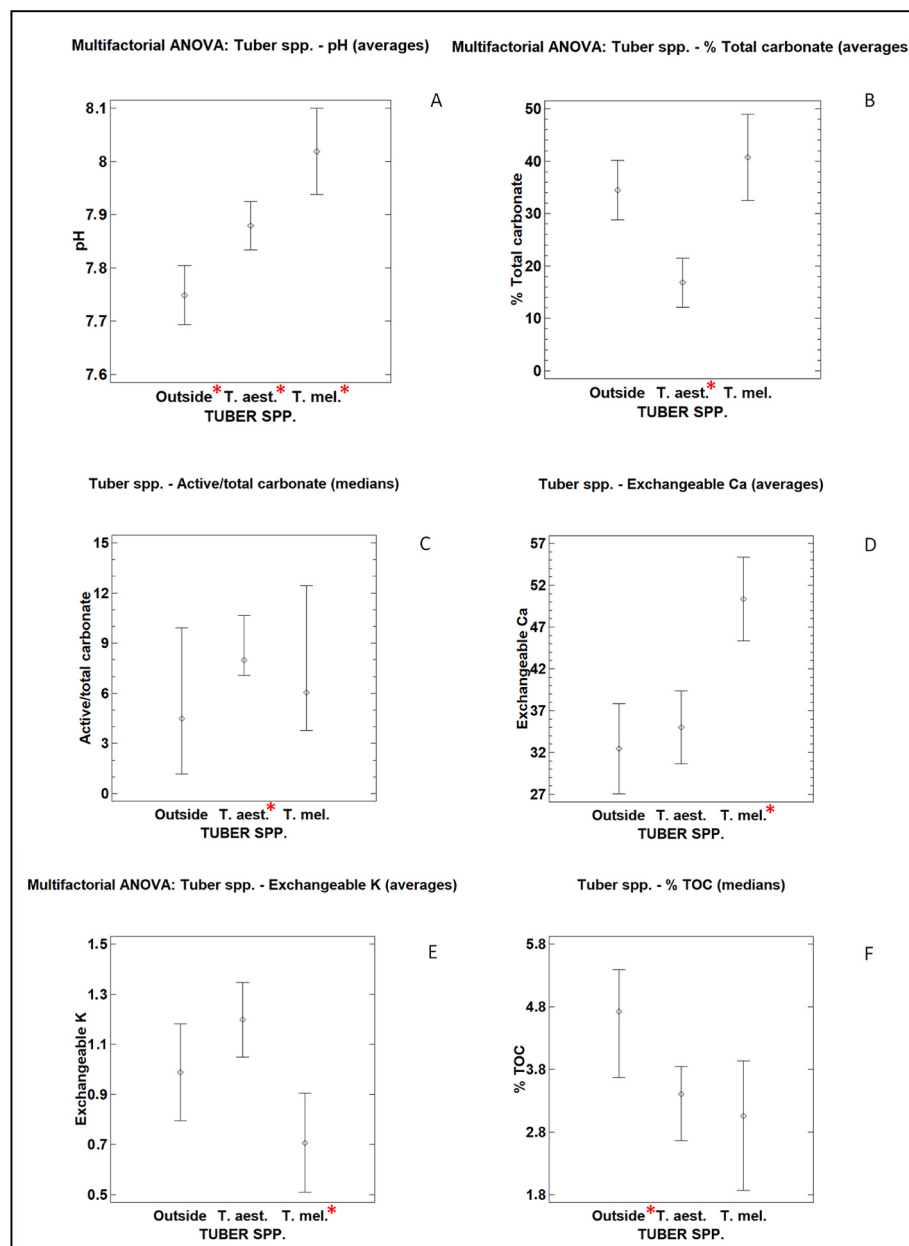


Fig. 4. Comparisons of average or median values of soil properties among the three groups of soils of *T. aestivum*, *T. melanosporum*, and soils outside the brûlés; pH (A), total carbonate (B), active/total carbonate ratio (C), exchangeable  $\text{Ca}^{2+}$  (D), exchangeable  $\text{K}^+$  (E), and total organic carbon (F) [ $*p < 0.05$ ].

**Table 1**

Soil properties of 45 brúles of *T. aestivum*, 45 brúles of *T. melanosporum*<sup>1</sup> and 44 Samples outside the brúles. [<sup>1</sup>Data partly published in García-Montero et al. (2007, 2014) added with data collected 2013–2015. <sup>2</sup>Brúle size in m<sup>2</sup>. <sup>3</sup>Maximum annual production of Tuber sporocarps in each brúle was calculated as the greatest quantity of fresh carpophores, in Kg, picked during a harvest season. <sup>4</sup>(g/100 g). <sup>5</sup>Exchangeable cations expressed in cmol + kg<sup>-1</sup>].

| <sup>1</sup> 134 Soil samples                 | Soils outside of the brúles |               |            | Soils inside <i>T. aestivum</i> brúles |               |             | Soils inside <i>T. melanosporum</i> brúles |               |             |
|-----------------------------------------------|-----------------------------|---------------|------------|----------------------------------------|---------------|-------------|--------------------------------------------|---------------|-------------|
|                                               | N                           | Average (SD)  | Max/Min    | N                                      | Average (SD)  | Max/Min     | N                                          | Average (SD)  | Max/Min     |
| <sup>2</sup> Brúle size                       | –                           | –             | –          | 45                                     | 45.69 (36.55) | 153/6.00    | 45                                         | 1.73 (1.47)   | 6.00/0.15   |
| <sup>3</sup> Brúle maximum production         | –                           | –             | –          | 45                                     | 0.65 (0.46)   | 2.00/0.10   | 45                                         | 12.90 (10.77) | 60.00/2.00  |
| Brúle plant cover: surface without plants (%) | –                           | –             | –          | 45                                     | 32.40 (19.35) | 80.00/5.00  | 45                                         | 72.84 (13.16) | 95.00/30.00 |
| pH                                            | 44                          | 7.77 (0.24)   | 8.17/7.00  | 45                                     | 7.88 (0.17)   | 8.16/7.48   | 45                                         | 7.96 (0.24)   | 8.35/7.10   |
| <sup>4</sup> Total carbonate                  | 44                          | 32.85 (25.08) | 94.51/0.21 | 45                                     | 16.83 (12.58) | 66.26/1.11  | 45                                         | 36.83 (28.01) | 85.76/1.46  |
| <sup>4</sup> Active carbonate                 | 44                          | 1.71 (2.06)   | 9.99/0.00  | 45                                     | 1.42 (1.01)   | 3.86/0.00   | 45                                         | 2.26 (2.13)   | 8.42/0.00   |
| <sup>4</sup> Active/total carbonate           | 44                          | 9.62 (18.44)  | 95.61/0.00 | 45                                     | 12.40 (11.78) | 46.31/0.00  | 45                                         | 19.48 (26.42) | 88.96/0.00  |
| <sup>4</sup> TOC                              | 44                          | 4.64 (2.36)   | 11.12/0.06 | 45                                     | 3.38 (1.23)   | 8.10/1.37   | 28                                         | 3.75 (3.10)   | 12.56/0.10  |
| <sup>5</sup> Ca <sup>2+</sup>                 | 30                          | 32.43 (17.40) | 72.43/9.62 | 45                                     | 35.01 (20.67) | 125.50/6.16 | 35                                         | 50.36 (24.17) | 122.55/8.64 |
| <sup>5</sup> Mg <sup>2+</sup>                 | 30                          | 5.56 (4.03)   | 16.10/0.42 | 45                                     | 6.56 (5.55)   | 19.62/0.21  | 35                                         | 5.43 (3.82)   | 17.48/0.41  |
| <sup>5</sup> K <sup>+</sup>                   | 30                          | 0.86 (0.45)   | 1.99/0.07  | 45                                     | 1.20 (1.07)   | 7.48/0.28   | 35                                         | 0.49 (0.28)   | 1.47/0.19   |

interaction soil group  $\times$  forest type was not (log-transformed data of soil carbonate content; soils:  $F_{2,131} = 6.59$ ,  $p = 0.019$ ; forest type:  $F_{2,131} = 4.48$ ,  $p = 0.013$ ; interaction soil group  $\times$  forest type:  $p = 0.724$ ).

Brúle characteristics of *T. aestivum* (size, productivity, vegetation cover) did not show significant relationships with total carbonate. In *T. melanosporum* brúles, there was a significant relationship between productivity and total carbonate, which more productive sites exhibiting lower carbonate content ( $\sqrt{\text{[total carbonate]}} = 7.33033 - 1.53143 \times \sqrt{\text{[brúle productivity]}}$ ,  $r^2 = 0.11$ ;  $p = 0.026$ ), indicating some modification of the total carbonate pool by that species.

There were significant no differences in active carbonate content among the three groups of soils due to large variations in active carbonate content between the three forest types, with very low contents in the most xeric forest type. Active carbonate content was highest in brúles of *T. melanosporum* and lowest in brúles of *T. aestivum*. The ratio active/total carbonate content was highest in brúles of *T. melanosporum* and lowest outside brúles, and the difference between these two soil groups was statistically significant (Fig. 4C). As this ratio is influenced by both effects of truffle species on total carbonate (a reduction by *T. aestivum*) and active carbonate (an increase by *T. melanosporum*), both truffle species showed a higher ratio compared to areas outside brúles. In the most xeric forest type (type 2), the ratio active/total carbonate content was very low, likely reflecting moisture limitations on carbonate dynamics. Brúle characteristics of *T. aestivum* did not have an impact on both parameters, whereas in brúles of *T. melanosporum* there were positive relationships between brúle size and productivity on the one hand and active carbonate and/or the ratio of active/total carbonate on the other hand. These relations were for brúle size ( $\log [\% \text{ active carbonate}] = -0.0202982 + 0.444812 \times \log [\text{brúle size} + 1]$ ;  $r^2 = 0.29$ ;  $p < 0.001$ ); and for the ratio of active/total carbonate ( $\log [\% \text{ active/total carbonate}] = 0.278966 + 0.64649 \times \log [\text{brúle size} + 1]$ ;  $r^2 = 0.10$ ;  $p = 0.031$ ). For brúle productivity, the relation was:  $\log [\text{act/tot carbonate}] = 0.558964 + 0.331396 \times \sqrt{\text{[prod]}}$ ;  $r^2 = 0.10$ ;  $p = 0.031$ ).

There was no significant difference in exchangeable Mg<sup>2+</sup> between the three soil types. Exchangeable Ca<sup>2+</sup> was significantly higher in *T. melanosporum* than in *T. aestivum* brúles or soils outside brúles (Fig. D;  $F_{2,107} = 7.54$ ,  $p < 0.001$ ), an effect consistent with the stronger modification of calcium carbonate, as assessed through the ratio active/total carbonate. For both Mg<sup>2+</sup> and Ca<sup>2+</sup>, both truffle species did not exhibit significant relationships with brúle characteristics.

There were significant differences between the three soil groups in soil exchangeable K<sup>+</sup> (Fig. 4E;  $F_{2,107} = 8.83$ ,  $p < 0.001$ ). *Tuber aestivum* brúles showed significantly higher K<sup>+</sup> than *T. melanosporum* brúles. For both truffle species, there were no relationships between brúle characteristics and K<sup>+</sup>.

TOC content was significantly higher outside brúles than inside brúles of both *Tuber* species (Fig. 4F; Kruskal-Wallis test), indicating an impact on both soil inorganic and soil organic components. There were

no relationships between TOC content and *T. aestivum* and *T. melanosporum* brúle characteristics.

Soil properties were summarized using Principal Components Analysis. The PCA that included all soil samples that were completely surveyed (91 cases) is shown in Supplementary Fig. 1. The first axis, PC<sub>1</sub>, accounted for 29% of the variance, PC<sub>2</sub> 23%, and PC<sub>3</sub> 16% (Bartlett's Test:  $p = 0.0$ ,  $n = 91$ ). The first axis separated soils with high contents of active carbonates, Ca<sup>2+</sup>, and a high active/total carbonate ratio from soils with high contents of Mg<sup>2+</sup> and represents a separation between brúle soils and soils outside brúles. PC<sub>2</sub> separates sites with high pH, total carbonate, and K<sup>+</sup> and separates forest types, with the most xeric forest being recognized by higher pH and total carbonate content. PC<sub>3</sub>, which explains 16% of the variance, separates sites with high TOC content and high total carbonate content from sites where both soil properties are low. This likely coincides with brúles created by *T. melanosporum* and *T. aestivum*.

### 3.3. Differential niche construction by *T. aestivum* and *T. melanosporum*

Soil properties outside brúles of *T. aestivum* and *T. melanosporum* did not exhibit significant differences, except for pH, which was much higher in soils outside the brúles of *T. aestivum* than in soils outside the brúles of *T. melanosporum* (Table 2).

Soils of brúles of *T. aestivum* had significantly higher pH than soils

**Table 2**

Soil properties inside and outside brúles of *T. aestivum* and *T. melanosporum*<sup>1</sup>.  $n$  = number of pairwise comparisons in a paired  $t$ -test. [ns: not significant; \*:  $0.01 < p < 0.05$ ; \*\*:  $0.001 < p < 0.01$ ; \*\*\*:  $p < 0.001$ ]. [<sup>1</sup>(g/100 g); <sup>2</sup>Exchangeable cations expressed in cmol + kg<sup>-1</sup>. <sup>3</sup>Part of the soil parameters of *T. melanosporum* brúles have been published before in García-Montero et al. (2009b)].

| <i>T. aestivum</i> brúles                  | n  | Inside | Outside | p    |
|--------------------------------------------|----|--------|---------|------|
| pH                                         | 20 | 7.91   | 7.63    | ***  |
| <sup>1</sup> Total carbonate               | 20 | 16.96  | 36.23   | **   |
| <sup>1</sup> Active carbonate              | 20 | 1.62   | 2.58    | n.s. |
| <sup>1</sup> Active/total carbonate        | 20 | 11.6   | 12.6    | n.s. |
| <sup>1</sup> TOC                           | 20 | 3.15   | 4.12    | *    |
| <sup>2</sup> Ca <sup>2+</sup>              | 20 | 33.9   | 34.8    | n.s. |
| <sup>2</sup> Mg <sup>2+</sup>              | 20 | 6.15   | 6.26    | n.s. |
| <sup>2</sup> K <sup>+</sup>                | 20 | 1.09   | 0.85    | n.s. |
| <sup>3</sup> <i>T. melanosporum</i> brúles | n  | Inside | Outside | p    |
| pH                                         | 20 | 7.89   | 7.91    | n.s. |
| <sup>1</sup> Total carbonate               | 20 | 21.66  | 33.63   | ***  |
| <sup>1</sup> Active carbonate              | 10 | 3.17   | 1.67    | **   |
| <sup>1</sup> Active/total carbonate        | 10 | 44.3   | 12.5    | **   |
| <sup>1</sup> TOC                           | 10 | 2.89   | 4.94    | n.s. |
| <sup>2</sup> Ca <sup>2+</sup>              | 10 | 44.0   | 27.8    | *    |
| <sup>2</sup> Mg <sup>2+</sup>              | 10 | 3.24   | 4.18    | n.s. |
| <sup>2</sup> K <sup>+</sup>                | 10 | 0.62   | 0.87    | n.s. |

outside brûlés and significantly lower contents of total carbonate and TOC. Soils of brûlés of *T. melanosporum* had significantly lower carbonate and significantly higher active carbonate, active/total carbonate ratio, and  $\text{Ca}^{2+}$  than soils outside brûlés (Table 2).

#### 4. Discussion

This study demonstrated significant soil modification by two truffle species, *T. aestivum* and *T. melanosporum*. Modifications are already visible from the creation of small to large bare patches with little or no vegetation cover (Fig. 1), and these sites, known as brûlés, are known by truffle hunters as the locations where they collect their highly-priced bounty. Brûlés differ from surrounding soils in several soil chemical properties, including a higher content of active carbonates (and hence a higher active/total carbonate ratio),  $\text{Ca}^{2+}$ , and a lower content of TOC. Our data also showed significant differences in habitat modification by both truffle species, with differences in brûlé area, the extent to which vegetation of non-host plants is reduced, pH, and  $\text{Ca}^{2+}$ .

Are these habitat modifications an instance of niche construction by ectomycorrhizal fungi? Matthews et al. (2014) listed two criteria that enable the recognition of niche construction. The first criterion of significant modification of environmental conditions is clearly met by both species, as shown by Tables 1 and 2, and the PCA in Supplementary Fig. 1. In addition, the second criterion is also met by both species because the formation of brûlés increased the fitness of these truffles. Truffles are hypogeous fungi that form sporocarps below-ground and therefore need active dispersal, often by mammals that are attracted by truffle aroma. Truffle spores are not digested in the stomachs of these mammals, therefore survive the gut passage, and their dung then acts as truffle inoculum (Ori et al., 2018; Piattoni et al., 2014). If we consider sporocarp production a proxy for spore production and hence for fungal fitness (Pringle and Taylor, 2002), the positive relationship between brûlé area (a proxy for its age) and sporocarp productivity implies increasing fitness with larger areas of modified habitat. The result by Liu et al. (2016) and Oliach et al. (2022) showing an increase in the number of truffle sequences upon ageing of plantations also supports the claim that brûlé formation enhances fungal fitness by reducing competition with other ectomycorrhizal fungi and partly eliminating arbuscular mycorrhizal fungi and plants, unless these plant species are favorable companion species.

Regarding the second criterion of Matthews et al. (2014), which implies that environmental modifications must influence selection pressures on recipient species, the creation of brûlés selects herbaceous plant species that are better able to withstand the conditions created by the truffles. González-Armada et al. (2010) noted that inside brûlés there is a larger representation of annual plant species. Among the species listed as characteristic for brûlés they also mentioned plants that are non-mycorrhizal or only sparsely colonized by arbuscular mycorrhizal fungi. In our sites, *Digitalis obscura* L. was often present in brûlés, a species known for its ability to grow on sites with high exchangeable  $\text{Ca}^{2+}$ , low exchangeable  $\text{Mg}^{2+}$ , and low P availability (Roca-Pérez et al., 2005). Taschen et al. (2020) compared six perennial plant species for their sensitivity to truffle brûlés and potential positive impact on truffle production. From this limited survey, it seems that sensitivity and positive impact are negatively correlated, suggesting that truffles could 'select' those plant species that have a subsequent beneficial impact on truffle production. However, a study by Barou et al. (2023) suggested that *Thymus vulgaris* L. suppresses rather than enhances truffle production, especially when it is arbuscular mycorrhizal. Menta et al. (2014) and Pinto et al. (2017) described how *T. aestivum* brûlés caused changes in soil fauna, with some collembolans benefitting from the changes in soil chemical conditions, while other collembolan species declined.

Truffle brûlés also exert a selective impact on arbuscular mycorrhizal and ectomycorrhizal fungi. Mello et al. (2015) observed a reduced arbuscular mycorrhizal fungal species richness inside *T. melanosporum* brûlés, and Taschen et al. (2020) observed decreased abundance of

arbuscular mycorrhizal fungi in these brûlés. Napoli et al. (2010) and Mello et al. (2011) also noted a reduced species richness of ectomycorrhizal fungi, especially of members of the Basidiomycota, inside brûlés, equally indicating the selection exerted by truffles when creating brûlés. This shift toward a larger representation of Ascomycota in brûlés than outside brûlés was also noted by Taschen et al. (2015). There is almost certainly a selection on the ectomycorrhizal host tree of truffles as well. The habitat modification caused by truffles increased the content of active carbonates and could increase the risk for iron chlorosis in the host trees (Hall et al., 2007). In this regard, the amount of active carbonate is often used as a proxy for predicting iron chlorosis in agroecosystems (Tagliavini and Rombolà, 2001). The brûlé could therefore transform the mutualistic relationship into unbalanced interactions, as proposed by García-Montero et al. (2009a, 2012). Soils with a high content of active carbonate in the brûlés lead chlorosis, which favors increased mycorrhization by *Tuber* spp., which in turn promotes the formation of new amounts of active carbonate according to a feedback model. The concept of the mutualistic niche, as a generalized case of mycorrhizal fungal niche construction (Peay, 2016), needs to be nuanced, as the niche construction by the ectomycorrhizal does not inevitably enhance the fitness of the tree. Our studies confirm that brûlé construction generates a low-quality productive environment with negative effects on soil quality (Innangi et al., 2020). Streiblová et al., (2012) concluded that *T. melanosporum* and *T. aestivum* brûlés had established an efficient environmental strategy to gain and control the space needed for their survival and reproduction, where they suppress the richness and biodiversity of indigenous organisms. It is interesting that Mamoun and Olivier (1993) observed increased growth of hazel but reduced mycorrhizal colonization by *T. melanosporum* after the addition of iron. Niche construction by truffles has been well-known by truffle cultivators. Dessolas et al. (2008) and Chevalier and Palenzona (2011) proposed that *Tuber* spp. "prepare their nest to produce carpophores". Our report on niche construction by truffles, therefore, expands the earlier suggestion by Bragato (2014) on the perturbational niche construction by truffles by changing aggregate sizes and soil porosity.

Matthews et al. (2014) and Laland et al. (2016) indicated that fulfilling these two criteria suffices for meeting the definition of niche construction as an ecological process. They added a third criterion to evaluate the evolutionary consequences of niche construction, viz. that there must be an evolutionary response in at least one of the species that has been caused by the environmental modification. We did not address that evolutionary question in our study. However, we consider it likely that there are evolutionary responses in those arbuscular mycorrhizal plant species that are both suitable companion species that enhance truffle productivity and escape from the detrimental effects that truffles exert on the surrounding vegetation and/or on the ectomycorrhizal host that must cope with increased risk of chlorosis. The contrary data reported by Taschen et al. (2020) and Barou et al. (2023) on the ecological impact of *Thymus* species would be consistent with local differentiation and evolution.

Whereas our data show that both *T. aestivum* and *T. melanosporum* construct their own niche, both species do so in somewhat different ways. This differential effect can be clearly seen from the fact that soils outside brûlés of both species are largely similar, whereas the brûlés are dissimilar. While these differences may reflect habitat choice by both truffle species in what would seem homogeneous sites, we would argue that these differences reflect problems in delimiting brûlés.

Both truffle species co-occur in calcareous regions of southern Europe and then compete depending on environmental conditions. Habitat characteristics of *T. melanosporum* and *T. aestivum* were provided by Jaillard et al. (2016) and Robin et al. (2016), respectively. However, these authors did not report on habitat modification by truffles. Wehrlen et al. (2009) indicated that *T. aestivum* is less climatically demanding and has a wider distribution area in Europe than *T. melanosporum*. *Tuber aestivum*, as opposed to *T. melanosporum*, can produce sporocarps in soils with more variable water levels and can

withstand flooding (Thomas, 2021). It is also able to better withstand dry conditions, as the br  le size of that species was not lower in the most xeric forest type, whereas that of *T. melanosporum* was significantly reduced. A study by Pi  uela et al. (2021) indicated that under irrigation *T. melanosporum* replaced *T. aestivum*, confirming higher drought tolerance of the latter species. Replacement of *T. melanosporum* by *T. aestivum* has been reported by Turgeman et al. (2012) and Garc  a-Montero et al. (2014). Both species seem to benefit from climate change (higher temperatures), and areas suitable for cultivation will likely increase in central Europe (  ejka et al., 2020).

In most cases, *T. aestivum* br  les were larger and non-host plant cover was greater than *T. melanosporum* br  les. Other differences between both species are visible in the PCA (Supplementary Fig. 1), Tables 1 and 2, and Supplementary Table 2. Soils of *T. melanosporum* br  les had higher pH than those of *T. aestivum*, but the difference in pH in properties inside and outside br  les was larger in the latter species. There was also a significant positive relationship between *T. aestivum* br  le size and soil pH. These data fit with observations by Gryndler et al. (2017) that liming increased the mycelial biomass of *T. aestivum*, and observations by Hilszcza  ska et al. (2018) that calcium carbonate content was the best predictor for *T. aestivum* productivity. The active/total carbonate ratio was higher in br  les of *T. melanosporum* and was significantly positively correlated with br  le area, suggesting the ongoing formation of active carbonates from the total carbonate pool. Higher exchangeable  $\text{Ca}^{2+}$  in br  les of *T. melanosporum* would fit with ongoing modification of the total carbonate pool. Nevertheless, in both species, the exact mechanisms of br  les formation are still poorly known.

Br  les of both species showed significantly lower contents of TOC, demonstrating that niche construction has consequences for both the organic and inorganic components of the soil. It had been hypothesized that truffle species possess some saprotrophic activity during sporocarp formation. However, a study by Zeller et al. (2008) demonstrated that truffles do not exhibit saprotrophy during sporocarp development. The mechanism for soil organic C loss may be due to a lower input of fresh litter (through the reduction in vegetation cover). However, the amounts of C lost are much larger than can be explained by differential inputs of fresh C. It is more likely that the creation of br  les results in environments that are more conducive to decomposition. In that respect, it may be noteworthy that Innangi et al. (2020), who also observed lower TOC inside br  les of *T. aestivum* than outside, reported no changes in peroxidase activity inside br  les compared to outside br  les. A hydrolytic enzyme, related to the breakdown of more labile C sources, was significantly lower in br  les than outside. One hypothesis is that inside br  les, where total carbonate content was reduced, organic C that was protected by those carbonates was released and became available for decomposition (Virto et al., 2018). This mechanism could equally explain the observations by Bragato (2014) on the decline of the fraction of macroaggregates in br  les and the protection of organic carbon in these macro aggregates. Transformation of carbonates may be caused by organic-acid production by truffle species, but the high carbonate content of these soils would buffer against pH decrease. Poitou (1988) showed that, *in vitro*, *T. melanosporum* mycelia acidify the environment and dissolve calcium carbonate. Callot (1999) reported an experiment on the growth conditions of *T. melanosporum* mycelia where pH values of 5.5–6 were reached in all cultures. However, to the best of our knowledge, there are no reports of oxalate or citrate exudation by truffle mycelia.

Truffle species can persist for at least twenty years on the same tree (Oliach et al., 2022). Truffle hunters even provided anecdotal evidence that br  les have persisted for over fifty years. However, habitat modification cannot go on indefinitely, and with the exhaustion of the carbonate pool, the productivity of br  les should decline. Queralt et al. (2017) showed that *T. melanosporum* extraradical mycelium biomass was more abundant in productive br  les than in non-productive ones. With the decline of truffle biomass, habitat modification declines. It may

be possible that the br  le is the arena for an oxalate-carbonate cycle, where fungi dissolve carbonate through the exudation of oxalic acid, and bacteria subsequently oxidize this calcium oxalate into carbonate (Verrecchia et al., 2006), but the mechanism has not often been demonstrated (Cromack et al., 1979).

In conclusion, br  le is an example of niche construction by ectomycorrhizal fungi in a limited spatial sense, where habitat modification is visible, and measurable in terms of soil chemistry, soil physics (Bragato, 2014), and soil biology -Mello et al. (2011) reported significant differences in fungal species composition inside and outside br  les; Mello et al. (2013) described differences in bacterial communities inside and outside br  les-. However, it is still necessary to deepen the br  le niche construction processes by analyzing other soil parameters, e.g., N, which likely will be reduced after the disappearance of soil organic matter (Table 1), P, micronutrients like Fe and Zn (whose bioavailability will decrease due to increased active carbonate) could further demonstrate soil habitat modification. Therefore, additional research on br  le niche construction is required, focusing, for example, on quantitative relations between truffle mycelial biomass and the extent of habitat modification or on the legacy effects in br  les after the truffles have disappeared.

On the other hand, habitat modification by fungi could go beyond br  le, and this phenomenon could be more widespread. Niche construction by ectomycorrhizal fungi, which had already been mentioned as a keyword by Peay (2016), is ubiquitous, as these fungi pave the way for tree establishment. This is visible in the case of pine and eucalyptus co-invasion and their associated ectomycorrhizal fungi, which sometimes show significant ecosystem effects. For example, Chapela et al. (2001) reported massive losses of soil organic carbon after pines with their associated ectomycorrhizal fungi invaded paramo grasslands in Ecuador, and Dickie et al. (2010) linked such co-invasions to invasion meltdown. Another prominent example of niche construction by ectomycorrhizal fungi is that formed by mat-forming fungi (Cromack et al., 1979). Trappe et al. (2012) listed some changes in soil chemical properties caused by these mat-forming fungi. Another example is the structure called shiro formed by the ectomycorrhizal fungus *Tricholoma matsutake* (S. Ito & S. Imai) Singer. Lian et al. (2006) describe changes in fungal communities after shiro formation. Therefore, a more general review of niche construction by ectomycorrhizal fungi would be necessary, where their edaphic habitat modifications can be discussed in more detail. A better understanding of mutualistic niche construction processes by mycorrhizal fungi could contribute to improving sustainable forestry, environmental, conservation, and agricultural engineering and management, such as proposed by Garc  a-Montero et al. (2017) through the "rhizoculture" approach.

## Author contributions

LGGM, TWK, and VJM designed the study. LGGM, IVA, and CM performed the field and laboratory tasks. VJM, TWK, and LGGM performed and discussed the statistical analyses. LGGM, TWK, and VJM wrote the manuscript draft. All authors edited the manuscript and approved the final version.

## Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

## Data availability

The data and code used to generate the results of this study are available at <https://zenodo.org/deposit/8025273>

## Acknowledgements

We thank Domingo Moreno (truffle-hunter), the late Dave Myrold (OSU), Jim Trappe (USDA F.S.), Dan Luoma (OSU), Jennifer Parke (OSU), Markus Kleber (OSU), Eric Verrecchia (UL), Rachel Creamer (WUR), Pepe Díaz (UCM), Xavier Parladé (IRTA), Carlos Colinas (ULL), Dani Oliach (ULL), the late José Miguel Barea (CSIC), Amaya Álvarez-Lafuente (El Serranillo), Marcos Morcillo (MICOFORA), the late Juan Hernando (UCM), Maribel Hernando (UCM), José Luis Manjón (UAH), José Ramón Quintana (UCM), Ana de Santiago (UCM), Cristina Pascual (UPM), Paz Andrés (UPM), Susana Martín (UPM), Esperanza Ayuga (UPM) and Luis Benito-Matías, for their viewpoints and scientific support during this lengthy study, and Margarita and Luis, Warren and Leslie for their personal support. The authors would like to give special thanks to other “truffle-hunters”: the late Justo, the late Domingo (padre), the late Emilio, José Mari, Pepe, Toño, and other truffle collectors as Abel, Gonzalo, and Vicente; and all the people and Council of “Peralejos de las Truchas” -especially to Dña. Pura-for their invaluable contribution to this study. We thank the Alto Tajo Nature Reserve Institution. From 1994 to the present, this work was supported by the financed by the I.N.I.A. Project SC94-129 (1994–1996), the I.M.I.A. Project FP-01-41 (2001), the I.T.D.A./I.M.I.A Research Agreement CO00-TN (2001–2002), the Grant of Spanish Education Ministry “Movilidad de Profesorado Salvador de Madariaga” (PR2011-0548) (2012), the UPM fundings of R + D Group DeMN (VAGI22LGGM) (2014–2023), the University of Alcalá Project CM/JIN/2019-002 (2020–2021), and the support of the research line on Biofertilizers promoted from the CORTEVA AGRISCIENCE-UPM Chair University-Company (Cod. CAT2013200335A) (2020–2023).

## Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.soilbio.2023.109276>.

## References

- AFNOR, 1982. Determination du calcaire actif. In: AFNOR (Ed.), *Qualité des sols*. AFNOR, Saint-Denis, pp. 31–106.
- Barou, V., Rincón, A., Calvet, C., Camprubí, A., Parladé, J., 2023. Aromatic plants and their associated arbuscular mycorrhizal fungi outcompete *Tuber melanosporum* in compatibility assays with truffle-oaks. *Biology* 12, 628.
- Bragato, G., 2014. Is truffle brûlé a case of perturbational niche construction? *Forest Systems* 23, 349–356.
- Callot, G., 1999. La truffe, la terre, la vie. INRA, Versailles.
- Cejka, T., Trnka, M., Krusic, P.J., Stobbe, U., Oliach, D., Václavík, T., Tegel, W., 2020. Predicted climate change will increase the truffle cultivation potential in central Europe. *Scientific Reports* 10, 21281.
- Ceruti, A., Fontana, A., Nosenzo, C., 2003. European Species of the Genus *Tuber*. A Historical Revision. Museo Regionale di Scienze Naturali, Torino.
- Chapela, I.H., Osher, L.J., Horton, T.R., Henn, M.R., 2001. Ectomycorrhizal fungi introduced with exotic pine plantations induce soil carbon depletion. *Soil Biology and Biochemistry* 33, 1733–1740.
- Chevalier, G., Palenzona, M., 2011. Les implications de 40 années de recherche sur les truffes: perspectives d'avenir. In: Donnini, D., Baciarelli Falini, L., Bencivenga, M., Di Massimo, G. (Eds.), *Atti di 3° Congresso Internazionale di Spoleto sul Tartufo*. Comunità Montana dei Monti Martani Serano e Subasio, pp. 531–550. Valtopina.
- Cromack Jr., K., Sollins, P., Graustein, W.C., Speidel, K., Todd, A.W., Spycher, G., Li, C. Y., Todd, R.L., 1979. Calcium oxalate accumulation and soil weathering in mats of the hypogeous fungus *Hysterangium crassum*. *Soil Biology and Biochemistry* 11, 463–468.
- Day, R.L., Laland, K.N., Odling-Smee, F.J., 2003. Rethinking adaptation: the niche-construction perspective. *Perspectives in Biology and Medicine* 46, 80–95.
- Dessolas, H., Chevalier, G., Pagnery, J.C., 2008. Nouveau manuel de trufficulture. Périgieux. FR: Misenpage.
- Dickie, I.A., Bolstridge, N., Cooper, J.A., Peltzer, D.A., 2010. Co-invasion by *Pinus* and its mycorrhizal fungi. *New Phytologist* 187, 475–484.
- FAO, 1990. Guideline for Soil Description. FAO, Roma.
- García-Montero, L.G., Casermeiro, M.A., Hernando, J., Hernando, I., 2006. Soil factors that influence the fruiting of *Tuber melanosporum* (black truffle). *Australian Journal of Soil Research* 44, 731–738.
- García-Montero, L.G., Manjón, J.L., Pascual, C., García-Abril, A., 2007. Ecological patterns of *Tuber melanosporum* and different *Quercus* Mediterranean woods: quantitative production of truffles, brûlé sizes and soil studies. *Forest Ecology and Management* 242, 288–296.
- García-Montero, L.G., Manzano, P., Alwanney, D., Valverde-Asenjo, I., Álvarez-Lafuente, A., Benito-Matías, L.F., Parladé, X., Ortuño, S., Morcillo, M., Gascó, A., Calderón-Guerrero, C., Mauro, F., Méndez, M., Sánchez-Medina, A., Andrés, M.P., Quintana, J.R., Menta, C., Pinto, S., Pinto, L., Pita, P., Turkmen, C., Pascual, C., Ayuga, E., Torrent, F., Robredo, J.C., Martín-Ortega, P., Pera, J., Gómez, L., Almendros, G., Colinas, C., Verrecchia, E.P., 2017. Towards an integrated understanding of the rhizosphere phenomenon as an ecological driver: can rhizoculture improve agricultural and forestry systems? In: Lukac, M., Grenni, P., Gamboni, M. (Eds.), *Soil Biological Communities and Ecosystem Resilience*. Springer International Publishing, Midtown Manhattan, pp. 43–75.
- García-Montero, L.G., Moreno, D., Monleon, V.J., Arredondo-Ruiz, F., 2014. Natural production of *Tuber aestivum* in central Spain: *Pinus* spp. versus *Quercus* spp. brûlés. *Forest Systems* 23, 394–399.
- García-Montero, L.G., Quintana, A., Valverde-Asenjo, I., Díaz, P., 2009a. Calcareous amendments in truffle culture: a soil nutrition hypothesis. *Soil Biology and Biochemistry* 41, 1227–1232.
- García-Montero, L.G., Valverde-Asenjo, I., Díaz, P., Pascual, C., 2009b. Statistical patterns of carbonates and total organic carbon on soils of *Tuber rufum* and *T. melanosporum* (black truffle) brûlés. *Australian Journal of Soil Research* 47, 206–212.
- García-Montero, L.G., Valverde-Asenjo, I., Moreno, D., Díaz, P., Hernando, I., Menta, C., Tarasconi, K., 2012. Influence of edaphic factors on edible ectomycorrhizal mushrooms: new hypotheses on soil nutrition and C sinks associated to ectomycorrhizae and soil fauna using the *Tuber brûlé* model. In: Zambonelli, A., Bonito, G. (Eds.), *Edible Ectomycorrhizal Mushrooms*. Springer-Verlag, Berlin, pp. 83–104.
- González-Armeda, B., De Miguel, A.M., Caverio, R.Y., 2010. Ectomycorrhizae and vascular plants growing in brûlés as indicators of below and above ground microecology of black truffle production areas in Navarra (Northern Spain). *Biodiversity & Conservation* 19, 3861–3891.
- Gryndler, M., Černá, L., Bukovská, P., Hřelová, H., Jansa, J., 2014. *Tuber aestivum* association with non-host roots. *Mycorrhiza* 24, 603–610.
- Gryndler, M., Šmilauer, P., Štovíček, V., Nováková, K., Hřelová, H., Jansa, J., 2017. Truffle biogeography-A case study revealing ecological niche separation of different *Tuber* species. *Ecology and Evolution* 7, 4275–4288.
- Hall, I.R., Brown, G.T., Zambonelli, A., 2007. Taming the Truffle. The History Lore and Science of the Ultimate Mushroom. Timber Press, Portland.
- Hilszczańska, D., Rosa-Gruszecka, A., Gawrys, R., Horak, J., 2018. Effect of soil properties and vegetation characteristics in determining the frequency of Burgundy truffle fruiting bodies in Southern Poland. *Écoscience* 10, 1530327.
- Innangi, M., Fioretto, A., Lozano Fondón, C., García-Montero, L.G., Marzaioli, R., Pinto, S., Rutigliano, F.A., Menta, C., 2020. The complex interrelationship between *Tuber aestivum* and soil chemical and biological properties. *Pedobiologia* 80, 150648.
- ISRIC, 1995. Procedures for Soil Analysis. FAO, Wageningen.
- Jailard, B., Oliach, D., Sourzat, P., Colinas, C., 2016. Soil characteristics of *Tuber melanosporum* habitat. In: Zambonelli, A., Iotti, M., Murat, C. (Eds.), *True Truffle (Tuber spp.) in the World*. Springer International Publishing, Midtown Manhattan, pp. 169–190.
- Kadowaki, K., Yamamoto, S., Sato, H., Tanabe, A., Hidaka, A., Toju, H., 2018. Mycorrhizal fungi mediate the direction and strength of plant–soil feedbacks differently between arbuscular mycorrhizal and ectomycorrhizal communities. *Communications Biology* 1, 196.
- Laland, K., Matthews, B., Feldman, M.W., 2016. An introduction to niche construction theory. *Evolutionary Ecology* 30, 191–202.
- Lian, C., Narimatsu, M., Nara, K., Hogetsu, T., 2006. *Tricholoma matsutake* in a natural *Pinus densiflora* forest: correspondence between above-and below-ground genets, association with multiple host trees and alteration of existing ectomycorrhizal communities. *New Phytologist* 171, 825–836.
- Liu, B., Fischer, C., Bonet, J.A., Castaño, C., Colinas, C., 2016. Shifts in soil fungal communities in *Tuber melanosporum* plantations over a 20-year transition from agriculture fields to oak woodlands. *Forest Systems* 25, eSC05.
- Mamoun, M., Olivier, J.M., 1993. Effect of iron amendment on the development of the ectomycorrhizal fungus *Tuber melanosporum* and the rhizoplane bacteria. *European Journal of Soil Biology* 29, 83–90.
- Matthews, B., De Meester, L., Jones, C.G., Ibelings, B.W., Bouma, T.J., Nuutinen, V., Van De Koppel, J., Odling-Smee, J., 2014. Under niche construction: an operational bridge between ecology, evolution, and ecosystem science. *Ecological Monographs* 84, 245–263.
- Mello, A., Ding, G.C., Piceno, Y.M., Napoli, C., Tom, L.M., DeSantis, T.Z., Andersen, G.L., Smalla, K., Bonfante, P., 2013. Truffle brûlés have an impact on the diversity of soil bacterial communities. *PLoS One* 8, e61945.
- Mello, A., Lumini, E., Napoli, C., Bianciotto, V., Bonfante, P., 2015. Arbuscular mycorrhizal fungal diversity in the *Tuber melanosporum* brûlé. *Fungal Biology* 119, 518–527.
- Mello, A., Napoli, C., Murat, C., Morin, E., Marceddu, G., Bonfante, P., 2011. ITS-1 versus ITS-2 pyrosequencing: a comparison of fungal populations in truffle-grounds. *Mycologia* 103, 1184e1193.
- Menta, C., García-Montero, L.G., Pinto, S., Conti, F.D., Baronia, G., Maresi, M., 2014. Does the natural “microcosm” created by *Tuber aestivum* affect soil microarthropods? A new hypothesis based on Collembola in truffle culture. *Applied Soil Ecology* 84, 31–37.
- Napoli, C., Mello, A., Borra, A., Vizzini, A., Sourzat, P., Bonfante, P., 2010. *Tuber melanosporum*, when dominant, affects fungal dynamics in truffle grounds. *New Phytologist* 85, 237–247.
- Núñez, M.A., Horton, T.R., Simberloff, D., 2009. Lack of belowground mutualisms hinders Pinaceae invasions. *Ecology* 90, 2352–2359.

- Odling-Smee, F.J., Erwin, D., Palkovacs, E.P., Feldman, M.W., Laland, K.N., Thomson, J., 2013. Niche construction theory: a practical guide for ecologists. *The Quarterly Review of Biology* 88, 3–28.
- Odling-Smee, F.J., Laland, K.N., Feldman, M.W., 2003. *Niche Construction: the Neglected Process in Evolution*. Princeton Univ. Press, Princeton.
- Oliach, D., Castaño, C., Fischer, C.R., Barry-Etienne, D., Bonet, J.A., Colinas, C., Oliva, J., 2022. Soil fungal community and mating type development of *Tuber melanosporum* in a 20-year chronosequence of black truffle plantations. *Soil Biology and Biochemistry* 165, 108510.
- Ori, F., Trappe, J., Leonardi, M., Iotti, M., Pacioni, G., 2018. Crested porcupines (*Hystrix cristata*): mycophagist spore dispersers of the ectomycorrhizal truffle *Tuber aestivum*. *Mycorrhiza* 28, 561–565.
- Pausas, J.G., Bond, W.J., 2022. Feedback in ecology and evolution. *Trends in Ecology & Evolution* 37, 637–644.
- Peay, K.G., 2016. The mutualistic niche: mycorrhizal symbiosis and community dynamics. *Annual Review of Ecology, Evolution, and Systematics* 47, 143–164.
- Peinado, M., Aguirre, J.L., Aparicio, A., 2017. The Iberian ranges and highlands. In: Loidi, J. (Ed.), *The Vegetation of the Iberian Peninsula*. Springer, Cham, pp. 349–512.
- Piattoni, F., Amicucci, A., Iotti, M., Ori, F., Stocchi, V., Zambonelli, A., 2014. Viability and morphology of *Tuber aestivum* spores after passage through the gut of *Sus scrofa*. *Fungal Ecology* 9, 52–60.
- Pinto, S., Gatti, F., García-Montero, L.G., Menta, C., 2017. Does soil fauna like truffles just as humans do? One-year study of biodiversity in natural brûlés of *Tuber aestivum* Vittad. *Science of the Total Environment* 584–585, 1175–1184.
- Piñuela, Y., Alday, J.G., Oliach, D., Castaño, C., Bolaño, F., Colinas, C., Bonet, J.A., 2021. White mulch and irrigation increase black truffle soil mycelium when competing with summer truffle in young truffle orchards. *Mycorrhiza* 31, 371–382.
- Plattner, L., Hall, I., 1995. Parasitism of non-host plants by the mycorrhizal fungus *Tuber melanosporum*. *Mycological Research* 99, 1367–1370.
- Poitou, N., 1988. Cas particulier des sols truffiers. *Bulletin FNPT-CTIFL* 10, 11–16.
- Pringle, A., Taylor, J.W., 2002. The fitness of filamentous fungi. *Trends in Microbiology* 10, 474–481.
- Queralt, M., Parladé, J., Pera, J., De Miguel, A.M., 2017. Seasonal dynamics of extraradical mycelium and mycorrhizas in a black truffle (*Tuber melanosporum*) plantation. *Mycorrhiza* 27, 565–576.
- R Core Team, 2013. *R: a Language and Environment for Statistical Computing*. R Foundation for Statistical Computing, Version 3.0.2. <http://www.R-project.org/>.
- Ramsey, F.L., Schaffer, D.W., 1997. *The Statistical Sleuth*. Duxbury Press, Belmont.
- Riouiset, L., Riouiset, G., Chevalier, G., Bardet, M.E., 2001. Truffles d'Europe et de Chine. INRA, Dijon.
- Rivas-Martínez, S., 1987. Memoria del mapa de series de vegetación de España. ICONA, Madrid.
- Robin, C., Goutal-Pousse, N., Le Tacon, F., 2016. Soil characteristics for *Tuber aestivum* (syn. *T. uncinatum*). In: Zambonelli, A., Iotti, M., Murat, C. (Eds.), *True Truffle (Tuber spp.) in the World*. Springer International Publishing, Midtown Manhattan, pp. 211–231.
- Roca-Pérez, L., Pérez-Bermúdez, P., Gavidia, I., Boluda, R., 2005. Relationships among soil characteristics, plant macronutrients, and cardenolide accumulation in natural populations of *Digitalis obscura*. *Journal of Plant Nutrition and Soil Science* 168, 774–780.
- Schneider-Maunoury, L., Clément, C., Coves, H., Lambourdière, J., Leclercq, S., Richard, F., Selosse, M.-A., Taschen, E., 2018. Is *Tuber melanosporum* colonizing the roots of herbaceous, non-ectomycorrhizal plants? *Fungal Ecology* 31, 59–68.
- Schneider-Maunoury, L., Deveau, A., Moreno, M., Todesco, F., Belmondo, S., Murat, C., Courty, P.-E., Jazkalski, M., Selosse, M.-A., 2020. Two ectomycorrhizal truffles, *Tuber melanosporum* and *T. aestivum*, endophytically colonise roots of non-ectomycorrhizal plants in natural environments. *New Phytologist* 225, 2542–2556.
- Statgraphics Centurion 18 Software. 1982–2018. *Statistical Software Version 18.1.12 (64-bits)*. Statgraphics Technologies, The Plains.
- Streiblová, E., Gryndlerová, H., Gryndler, M., 2012. Truffle brûlé: an efficient fungal life strategy. *FEMS Microbiology Ecology* 80, 1–8.
- Tagliavini, M., Rombolà, A., 2001. Iron deficiency and chlorosis in orchard and vineyard ecosystems. *European Journal of Agronomy* 15, 71–92.
- Taschen, E., Sauve, M., Taudiere, A., Parladé, J., Selosse, M.A., Richard, F., 2015. Whose truffle is this? Distribution patterns of ectomycorrhizal fungal diversity in *Tuber melanosporum* brûlés developed in multi-host Mediterranean plant communities. *Environmental Microbiology* 17, 2747–2761.
- Taschen, E., Sauve, M., Vincent, B., Parladé, J., van Tuinen, D., Aumeeruddy-Thomas, Y., Assenat, B., Selosse, M.-A., Richard, F., 2020. Insight into the truffle brûlé: tripartite interactions between the black truffle (*Tuber melanosporum*), holm oak (*Quercus ilex*) and arbuscular mycorrhizal plants. *Plant and Soil* 446, 577–594.
- Thomas, P.W., 2021. Ectomycorrhiza resilience and recovery to extreme flood events in *Tuber aestivum* and *Quercus robur*. *Mycorrhiza* 31, 511–517.
- Trappe, M.J., Cromack Jr., K., Caldwell, B.A., Griffiths, R.P., Trappe, J.M., 2012. Diversity of mat-forming fungi in relation to soil properties, disturbance, and forest ecotype at Crater Lake National Park, Oregon, USA. *Diversity* 4, 196–223.
- Turgeman, T., Sitrit, Y., Danai, O., Luzzati, Y., Bustan, A., Roth-Bejerano, N., Kagan-Zur, V., Masaphy, S., 2012. Introduced *Tuber aestivum* replacing introduced *Tuber melanosporum*: a case study. *Agroforest Systems* 84, 337–343.
- Verlhac, A., Giraud, M., Leteinturier, J., 1990. *La Truffe Guide Pratique*. Centre technique interprofessionnel des fruits et légumes CTIFL, Paris.
- Verrecchia, E.P., Braissant, O., Cailleau, G., 2006. The oxalate–carbonate pathway in soil carbon storage: the role of fungi and oxalotrophic bacteria. In: Gadd, G.M. (Ed.), *Fungi in Biogeochemical Cycles*. Cambridge University Press, Cambridge, pp. 289–310.
- Virto, I., Antón, R., Apesteguía, M., Plante, A., 2018. Role of carbonates in the physical stabilization of soil organic matter in agricultural Mediterranean soils. In: Muñoz, M. A., Zornoza, R. (Eds.), *Soil Management and Climate Change*. Academic Press, Cambridge, pp. 121–136.
- Wehrle, L., Chevalier, G., Besancon, G., Frochet, H., 2009. Truffle Cultivation-Forestry: a New Strategy of Producing the Burgundy Truffle (*Tuber uncinatum* Chatin). *Acta Bot Yunnanica* XVI, pp. 97–99.
- Zeller, B., Bréchet, C., Maurice, J.P., Le Tacon, F., 2008. Saprotrophic versus symbiotic strategy during truffle ascocarp development under holm oak. A response based on  $^{13}\text{C}$  and  $^{15}\text{N}$  natural abundance. *Annals of Forest Science* 65, 607–614.